Manual of Clinical Dialysis

Second Edition

Manual of Clinical Dialysis

Second Edition

Suhail Ahmad

University of Washington, Scribner Kidney Center, Northwest Kidney Centers, Seattle, Washington, USA



Suhail Ahmad, MD Professor, Medicine University of Washington Scribner Kidney Center Northwest Kidney Centers Seattle, Washington sahmad@u.washington.edu

ISBN: 978-0-387-09650-6 e-ISBN: 978-0-387-09651-3

DOI: 10.1007/978-0-387-09651-3

Library of Congress Control Number: 2008928866

© Springer Science+Business Media, LLC 2009

All rights reserved. This work may not be translated or copied in whole or in part without the written permission of the publisher (Springer Science+Business Media, LLC, 233 Spring Street, New York, NY 10013, USA), except for brief excerpts in connection with reviews or scholarly analysis. Use in connection with any form of information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed is forbidden.

The use in this publication of trade names, trademarks, service marks, and similar terms, even if they are not identified as such, is not to be taken as an expression of opinion as to whether or not they are subject to proprietary rights

While the advice and information in this book are believed to be true and accurate at the date of going to press, neither the authors nor the editors nor the publisher can accept any legal responsibility for any errors or omissions that may be made. The publisher makes no warranty, express or implied, with respect to the material contained herein.

Printed on acid-free paper

springer.com

Grateful thanks to my family, Vimli, Saba, and Zeba, and friends for their strong support and to Arlene for all of her help. This work is dedicated to my teachers, namely Dr. Scribner, and all of my patients.

Foreword (For First Edition)

As the next millennium begins, we hope that in the years ahead the need for dialysis will be decreased by better preventive care, especially the control of hypertension during the early stages of chronic renal disease. An increase in the number of donated kidneys and a decrease in their rejection rate also seems possible. In the meantime, it is our goal as dialysis professionals to do the very best job we can to make dialysis treatments as effective as possible in terms of patient survival and rehabilitation.

Despite the excellence of this manual, in terms of dialysis dose, one conclusion is inescapable: the current recommendation for dialysis dose, although recently revised upward, is still too low to support the well-being needed for rehabilitation. Indeed, at a urea reduction ratio of 65%, which is the current minimum set by Medicare, patients remain chronically uremic. The author does not say this, but if you read between the lines, he is trying to tell the reader that it is true. Furthermore, this dosage is based on observed (often malnourished) body weight, whereas it should be based on ideal body weight to reflect more accurately the needed dose.

Equally bad for patient well-being is the fact that there is no margin of safety built into this minimum. I believe a margin of safety is essential since the delivered dose is not checked with every dialysis; yet every aspect of dialysis procedures works against delivering the prescribed dialysis dose. For example, if adverse intradialytic events occur during a session, the time lost is seldom made up.

It is important to point out that the higher the weekly dose of dialysis the better. No adverse effects have been encountered no matter how high the dose. Pierratos has shown, with seven nights per week of home dialysis, a marked improvement in well-being, using a dose so large that phosphate had to be added to the dialysate.

Access to the circulation still is the "Achilles heel" of hemodialysis, and recirculation is a major cause of under-dialysis. The native Cimino fistula remains the gold standard. Vein grafts should never be used in patients in whom any natural vein is usable.

In the case of peritoneal dialysis, the danger of under-dialysis is ever present. Since the contribution by the native kidney in controlling uremia is more important in this group of patients, loss of residual renal function puts patients at risk for

severe under-dialysis. In this group the close monitoring of the dose, including that supplied by residual renal function, is particularly critical to avoid adverse patient outcome.

The basic constraint that Dr. Ahmad has to live with in order to be realistic in his dosage recommendations is the "standard" $3^{1}/_{2}$ h, three times a week dialysis schedule. Both dialysis professionals and their patients must come to understand the basic fact that it is not possible, except in small patients, to give enough dialysis in $3^{1}/_{2}$ h, three times a week, to support rehabilitation. The current dismally low rehabilitation rate supports this contention. As suggested in the text, there are many ways to get beyond this session time constraint, and I hope these suggestions will increase interest in pursuing them according to the needs of the individual patient because under-dialysis is a major cause of failure to rehabilitate dialysis patients.

Of course, as soon as ways are found to break through this $3^{1}/_{2}$ h, three times a week session time barrier, other benefits begin to fall into place, such as correction of chronic acidosis and especially the ability to control blood pressure. It is my opinion, which is based on a huge amount of practical experience and published material, that antihypertensive medications are totally ineffective in controlling blood pressure in the dialysis population. In addition, their use increases the incidence of hypotensive episodes, especially during short dialyses. Indeed, antihypertensive medications must be discontinued before the patient's extracellular volume can be reduced to the level of dry weight. I define dry weight as that which reflects an extracellular volume small enough to render the dialysis patient normotensive and unable to tolerate antihypertensive medications. However, all this important information cannot be applied unless at least 12–15 h per week are devoted to being on dialysis.

Even if professional staff optimize every aspect of dialysis according to the guidelines in this manual, there still remains a key task. Staff must convince the patients of the vital importance to their well-being of receiving the highest possible dose of dialysis. Physicians in particular have proven to be poor advocates and teachers of this crucial objective. Patients must be made to understand that the higher the weekly dialysis dose, the better they will feel. Then it is up to the patients to decide whether it is a worthwhile trade off to spend extra time on dialysis in exchange for better sense of well-being, without which rehabilitation is very difficult if not impossible.

Seattle, Washington, USA July, 1999 Belding H. Scribner, MD

Acknowledgements

Table 2.4 is adapted with permission from: ANSI/AAMI RD62: 2006 with permission of the Association for the Advancement of Medical Instrumentation, Inc. (C) 2006 AAMI www.aami.org. All rights reserved.

Figure 4.7 is adapted with permission from: Uldall R. Hemodialysis access. Part A: Temporary. In *Replacement of Renal Function by Dialysis, 4th Edition*. Edited by C Jacobs, CM Kjellstrand, KM Koch and JF Winchester. Dordrecht: Kluwer, 1996, pp. 277–293. © Kluwer.

Figure 5.6 is adapted with permission from: Golper TA, Wolfson M, Ahmad S et al. Multicenter trial of L-carnitine in maintenance hemodialysis patients. I. Carnitine concentrations and lipid effects. *Kidney Int* 1990, 38:904–911.

Figure 6.4 is adapted with permission from: 14. Daugirdas JT, Depner TA. A nomogram approach to hemodialysis urea modeling. *Am J Kidney Dis* 1994, 23:33–40.

Tables 7.2 and 7.8 are reproduced with permission from: Kaplan AA. Continuous arteriovenous hemofiltration and related therapies. In *Replacement of Renal Function by Dialysis. 4th Edition*. Edited by C Jacobs, CM Kjellstrand, KM Koch and JF Winchester. Dordrecht: Kluwer Academic Publishers, 1993:390–418. © Kluwer.

Figure 8.1 is adapted with permission from: Twardowski ZJ. Physiology of Peritoneal Dialysis. In *Clinical Dialysis*, *3rd Edition*. Edited by AR Nissensson, RN Fine, and DE Gentile. Norwalk: Appleton and Lange,1995, pp. 322–342.

Figure 8.2 is reproduced with permission from: Nolph KD, Miller F, Rubin J et al. New directions in peritoneal dialysis concepts and applications. *Kidney Int* 1980, 18(Suppl 10):111–116.

Table 7.1 reproduced with permission from Mehta RL, McDonald BR, Aguilar MM, et al. Regional citrate anticoagulation for continuous arteriovenous hemodialysis in critically ill patients. Kidney Int 1990, 38:976–981

Tables 7.3 and 7.4 adapted with permission from Daugirdas JT, Blake PG, Ing TS: Handbook of Dialysis, 4th ed. Philadelphia, PA: Lippincott, William & Wilkins, 2007

Table 7.6 reproduced with permission from Swartz R, et al. Improving the delivery and continuous renal replacement therapy using regional citrate anticoagulation. Clin Nephrol 2004, 61:134–143

Table 9.5 is reproduced with permission from: Diaz-Buxo JA. Clinical use of peritoneal dialysis. In *ClinicalDialysis*, *3rd Edition*. Edited by AR Nissenson, RN Fine and DE Gentile. Norwalk: Appleton & Lange, 1995, pp. 376–425.

Figure 10.1 is reproduced with permission from: Diaz-Buxo JA. Chronic peritoneal dialysis prescription. In *Handbook of Dialysis, 2nd Edition*. Edited by JT Daugirdas and TS Ing. Philadelphia: Lippincott-Raven, 1993, pp. 310–327. © Lippincott, Williams & Wilkins.

Figure 10.2 is adapted with permission from: Diaz-Buxo JA. Clinical use of peritoneal dialysis. In *Clinical Dialysis*, *2nd Edition*. Edited by AR Nissenson, RN Fine, and DE Gentile. Norwalk: Appleton & Lange, 1990, 13:256–300.

Figure 10.3 is adapted with permission from: Twardowski ZJ et al. Peritoneal equilibration test. *Perit Dial Bull* 1987, 7:138–140.

Table 11.5 is reproduced with permission from: Gokal R. Peritoneal infections, hernias and related complications. In *Replacement of Renal Function by Dialysis*, *4th Edition*. Edited by C Jacobs, CM Kjellstrand, KM Kochand JF Winchester. Dordrecht: Kluwer, 1996, pp. 657–688. © Kluwer.

Table 11.6 is reproduced with permission from: Bargman JM. Noninfectious complications of peritoneal dialysis. In *The Textbook of Peritoneal Dialysis*. Edited by R Gokal and KD Nolph. Dordrecht: Kluwer, 1994:555–591. © Kluwer.

Figure 12.1 is reproduced with permission from: Lowrie EG, Lew LN. Death risk in hemodialysis patients: the predictive value of commonly measured variables and an evaluation of death rate differences between facilities. *Am J Kidney Dis* 1990, 5:458–482.

Table 12.3 is reproduced with permission from: Fouque D, Kopple JD. Malnutrition and dialysis. In *Replacement of Renal Function by Dialysis. 4th Edition*. Edited by C Jacobs, CM Kjellstrand, KM Koch and JF Winchester. Dordrecht: Kluwer, 1996:1271–1290. © Kluwer.

Figures 13.3 and 13.4 are reproduced with permission from: Charra B, Calemard E, Cuche M et al. Control of hypertension and prolonged survival on maintenance hemodialysis. *Nephrol* 1983, 33:96–102.

Figure 13.2 reproduced with permission from Eknoyan G, Beck GJ, Breyer JA et al. Design and preliminary results of the mortality and morbidity of hemodialysis (MMHD) pilot study [Abstract]. J Am Soc Nephrol 1994, 5:513.

Figure 13.9 is adapted with permission from: Buonchristiani U, Fagugli RM, Pinciaroli MR et al. Reversal of left ventricular hypertrophy in uremic patients by treatment of daily hemodialysis (dhd). *Contrib Nephrol* 1996, 119:152–156.

Table 13.3 is adapted with permission from: London G, Marchais S, Guerin AP. Blood pressure control in chronic hemodialysis patients. In *Replacement of Renal Function by Dialysis, 4th Edition*. Edited by C Jacobs, CM Kjellstrand, KM Koch and JF Winchester. Dordrecht: Kluwer, 1996:966–990. © Kluwer.

Figure 15.1 is reproduced with permission from: Rose BD, Rennke HG. Signs and symptoms of chronic renal failure. In *Renal Pathophysiology*. Edited

Acknowledgements xi

by BD Rose and HG Rennke. Baltimore: Williams & Wilkins, 1994:276-300.

© Lippincott, Williams & Wilkins.

Published by Science Press Ltd, 34–42 Cleveland Street, London W1P 6LB, UK. $^{\circledR}$ 1999 Science Press Ltd.

First published 1999. Reprinted 2000, 2003.

http://www.science-press.com/

All rights reserved. No part of this publication may be reproduced, stored in a retrieval system or transmitted in any form or by any means electronic, mechanical, photocopying, recording or otherwise without prior written permission of the publishers.

British Library Cataloguing in Publication Data.

A catalogue record for this book is available from the British Library.

ISBN 1-85873-345-6

Although every effort has been made to ensure that drug doses and other information are presented accurately in this publication, the ultimate responsibility rests with the prescribing physician. Neither the publishers nor the authors can be held responsible for errors or for any consequences arising from the use of information contained herein. Any product mentioned in this publication should be used in accordance with the prescribing information prepared by the manufacturers. No claims or endorsements are made for any drug or compound at present under clinical investigation.

Project editor: Mark Knowles Illustrator: Stuart Molloy Typesetter: Simon Banister Designer: Claire Huntley Production: Adrienne Hanratty

Printed in Singapore by Stamford Press

Contents

F O	rewor	d (For 1	First Edition)	V11	
Ac	knowl	edgeme	ents	ix	
Ab	brevia	ations .		xxi	
1	Brie	ef Histo	ry of Clinical Dialysis: The Seattle Experience	1	
	1.1		tion of Dialysis	3	
	1.2	Mecha	anisms Involved in Molecular Movement	3	
		1.2.1	Diffusion	4	
		1.2.2	Ultrafiltration	4	
		1.2.3	Osmosis	5	
		1.2.4	Convection	5	
	1.3	Cleara	nnce	5	
		1.3.1	Blood vs Plasma Clearance	6	
		1.3.2	Clinical Factors Influencing Dialysis Urea Clearance	6	
	Refe	erence.		6	
2	Hemodialysis Technique				
	2.1	Blood	Flow Rate	7	
	2.2	Dialys	sate Flow Rate	7	
	2.3	Dialyz	zer Efficiency and Mass Transfer Area Coefficient (KoA)	8	
	2.4	Differ	ent Hemodialysis Techniques	8	
		2.4.1	Traditional Hemodialysis	8	
		2.4.2	Hemofiltration	9	
		2.4.3	Hemodiafiltration	10	
		2.4.4	Slow Low Efficiency Dialysis (SLED)	10	
		2.4.5	Ultrafiltration	10	
	2.5	Hemo	dialysis Setup	10	
		2.5.1	Blood Circuit	11	
		2.5.2	Dialysate Circuit	18	
	Refe	erences		2.7	

xiv Contents

3	Ant	icoagul	ation	29
	3.1		in Anticoagulation	
		3.1.1	Systemic Standard Heparinization	31
		3.1.2	Low-Dose Heparinization	31
		3.1.3	Low Molecular Weight Heparin	
	3.2		ems with Heparin Anticoagulation	32
	3.3		atives to Heparin	
		3.3.1	Citrate Anticoagulation	33
	3.4	No Ar	nticoagulation	35
	Refe			36
	₹7			27
4			ccess	37
	4.1		nent Access	37
		4.1.1	Preparation	
		4.1.2	Arteriovenous Fistula	38
		4.1.3	Arteriovenous Graft	41
		4.1.4	Diagnosis and Management of Arteriovenous	
			Dialysis Access	43
		4.1.5	Dual-Lumen Catheters with Dacron Cuff	45
		4.1.6	Special Arteriovenous Shunts	48
	4.2		prary Access	49
		4.2.1	General Technique	
		4.2.2	Complications of Temporary Access	
		4.2.3	Comparison of the Three Access Sites	
	4.3	-	t of Access	55
		4.3.1	Access Surveillance	56
	Refe	erences		58
5	Con	nplicati	ons of Hemodialysis	59
	5.1		al Complications	
		5.1.1	Hypotension	59
		5.1.2	Cardiac Arrhythmias	68
		5.1.3	Intradialytic Hypertension	69
		5.1.4	Muscle Cramps	69
		5.1.5	Carnitine and Intradialytic Hypotension, Arrhythmias,	0)
		3.1.5	and Muscle Cramps	70
		5.1.6	Nausea and Vomiting	71
		5.1.7	Headache	71
		5.1.8	Serious, Less Common Complications	72
	5.2		ne-Related Complications	75
		5.2.1	Air Embolism	75
		5.2.2	Hemolysis	
	Refe	erences	•	76

Contents xv

6	Dos	e of Her	modialysis	79
	6.1	Histori	ical Background	79
		6.1.1	Dialysis Index	80
		6.1.2	Urea Clearance	80
		6.1.3	Urea as a Marker for Uremic Toxins	81
		6.1.4	Current Methods of Measuring Dialysis Dose	81
	6.2	Potent	ial Problems with the Calculation of Dialysis Dose	83
		6.2.1	Influence of the Single-Pool Model	83
	6.3	Detern	nining Adequate Dialysis	91
		6.3.1	Acceptable Kt/V Values	92
		6.3.2	Frequency of Dose Measurement	92
	Refe	erences .		92
7	Con	tinnons	Therapies	95
•	7.1		iew	95
	7.2		of Continuous Therapies	95
	,	7.2.1	Continuous Arteriovenous Hemofiltration (CAVH)	95
		7.2.2	Continuous Venovenous Hemofiltration (CVVH)	96
		7.2.3	Continuous Venovenous Hemodialysis (CVVHD)	98
		7.2.4	Continuous Venovenous Hemodiafiltration (CVVHDF)	99
		7.2.5	Slow Low-Efficiency Diffusion Hemodialysis (SLEDD)	99
		7.2.6	Slow Continuous Ultrafiltration (SCUF)	
		7.2.7	Newer Technologies	
	7.3	Compo	onents of Continuous Therapies	
		7.3.1	Vascular Access	
		7.3.2	Tubing	
		7.3.3	Filter	
		7.3.4	Replacement Fluid	
		7.3.5	Dialysis Fluid	
		7.3.6	Machines	
	7.4	Dialys	ate Flow and Ultrafiltration Rates	110
	7.5	Antico	pagulation	112
		7.5.1	Heparin	
		7.5.2	Low Molecular Weight Heparin	113
		7.5.3	Citrate	113
		7.5.4	Prostacyclin	114
		7.5.5	Argatroban	
		7.5.6	Lepirudin	115
		7.5.7	Danaparoid	
		7.5.8	Fondaparinux	
		7.5.9	Nafamostat	
		7.5.10	No Anticoagulation	
	7.6	Drug F	Removal During CRRT	117

xvi Contents

	7.7 7.8 Refe	Intraoperative Dialysis	119
8	Dori	toneal Dialysis	123
0	8.1	Historical Background	
	8.2	Anatomy and Physiology	
	8.3	Kinetics of Peritoneal Transport	
	0.5	8.3.1 Diffusion	
		8.3.2 Ultrafiltration	
	Refe	erences	
9	Tech	nnique of Peritoneal Dialysis	129
	9.1	Peritoneal Dialysis Catheters	
		9.1.1 Description	
		9.1.2 Catheter Insertion Technique	
	9.2	Peritoneal Dialysis Fluid	
		9.2.1 Osmotic Agents	
	9.3	Delivery Mechanism	
	9.4	Peritoneal Dialysis Techniques	
		9.4.1 Continuous Ambulatory Peritoneal Dialysis (CAPD)	140
		9.4.2 Automated Peritoneal Dialysis (APD)	140
	Refe	erences	143
10	Dose	e of Peritoneal Dialysis	145
	10.1	Weekly Creatinine Clearance	145
		10.1.1 Residual Glomerular Filtration Rate	
		10.1.2 Peritoneal Creatinine Clearance	146
		10.1.3 Correction for Body Surface Area	
		10.1.4 Total Weekly Creatinine Clearance Calculation	148
	10.2	Urea Clearance Concept (Kt/V _{urea})	148
		10.2.1 Volume of Distribution of Urea	148
	10.3	Recommended Dose of Dialysis	149
		10.3.1 Potential Problem with Dose Measurements	
		10.3.2 Frequency of Dose Determination	
	10.4	Peritoneal Function Test	
		10.4.1 Traditional Peritoneal Equilibration Test	
		10.4.2 Fast Peritoneal Equilibration Test	
		10.4.3 Results of the Peritoneal Equilibration Test	152
	10.5	Use of Fast Peritoneal Equilibration Test Results in Selecting	
		a Peritoneal Dialysis Regimen	153
		10.5.1 Selection of Technique	
	Refe	prences	155

Contents xviii

11	Complications of Peritoneal Dialysis	. 157
	11.1 Peritonitis	. 157
	11.1.1 Clinical Diagnosis	. 157
	11.1.2 Therapy	. 160
	11.2 Exit Site and Tunnel Infection (also see Chapter 9)	
	11.3 Under-dialysis	. 165
	11.4 Malnutrition	. 166
	11.5 Membrane Failure	
	11.6 Cardiovascular Complications	
	11.7 Intra-Abdominal Pressure	
	11.8 Hemoperitoneum	
	References	. 167
12	Nutritional Issues	. 169
	12.1 Protein Calorie and Nutritional Status of Dialysis Patients	
	12.2 Significance of Nutritional Status	
	12.2.1 Hemodialysis	
	12.2.2 Peritoneal Dialysis	
	12.3 Factors Causing Malnutrition	
	12.3.1 Uremia	
	12.3.2 Other Factors	
	12.4 Assessment of Nutritional Status	
	12.4.1 Dietary Intake	
	12.4.2 Anthropometry and Body Weights	
	12.4.3 Bioelectric Impedance Analysis	
	12.4.4 Dual Energy X-Ray Absorptiometry	. 175
	12.4.5 Subjective Global Assessment	
	12.4.6 Biochemical Assessment	. 176
	12.5 Nutritional Requirements	. 176
	12.5.1 Protein	. 178
	12.5.2 Caloric Intake	. 178
	12.5.3 Lipids	. 178
	12.5.4 Fatty Acids, Lipids, and Carnitine	. 178
	12.5.5 Vitamins and Trace Elements	. 179
	12.5.6 Additional Nutritional Support	
	12.5.7 Metabolic Acidosis	. 180
	References	. 181
13	Hypertension	. 183
-	13.1 Prevalence	
	13.2 Control of Hypertension	
	13.3 Significance of Hypertension Control	
	13.4 Pathogenesis	
	13.4.1 Sodium Excess	
	13.4.2 Other Factors	

xviii Contents

	13.5 Treatment of Hypertension	188
	13.5.1 Sodium and Volume Control	
	13.5.2 Ultrafiltration	
	13.5.3 Pharmacological Agents	
	13.6 Conclusions	
	References	
14	Anemia	
	14.1 Pathogenesis	
	14.1.1 Erythropoietin	
	14.1.2 Uremic Factors	
	14.1.3 Other Factors	
	14.2 Treatment of Anemia	201
	14.2.1 Erythropoietin-Stimulating Agents (ESA)	• • •
	and Administration	
	14.3 Iron Status	
	14.3.1 Tests to Evaluate Iron Status	
	14.3.2 Iron Supplementation	
	14.4 Carnitine	
	14.5 Other Measures to Improve Hematocrit Response	
	References	208
	1010101000	200
15		
15	Renal Osteodystrophy	211
15	Renal Osteodystrophy	211
15	Renal Osteodystrophy	211 211
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention	211 211 211
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone	211 211 211 212
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy	211 211 211 212 212 214
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease	211 211 211 212 212 214 215
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism. 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone. 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease.	211 211 211 212 212 214 215
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease	211 211 212 212 214 215 215
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy	211 211 212 212 214 215 215 216
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy 15.4 Metastatic Calcification	211 211 212 212 214 215 216 216 216
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy 15.4 Metastatic Calcification 15.5 Laboratory Findings and Management of ROD	211 211 212 212 214 215 216 216 217 218
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy 15.4 Metastatic Calcification 15.5 Laboratory Findings and Management of ROD 15.5.1 Laboratory Findings	211 211 212 212 214 215 216 216 216 218
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy 15.4 Metastatic Calcification 15.5 Laboratory Findings and Management of ROD 15.5.1 Laboratory Findings 15.5.2 High Turnover Disease	211 211 212 212 214 215 216 216 217 218 218
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy 15.4 Metastatic Calcification 15.5 Laboratory Findings and Management of ROD 15.5.1 Laboratory Findings	211 211 212 212 214 215 216 216 217 218 218 218 2121
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy 15.4 Metastatic Calcification 15.5 Laboratory Findings and Management of ROD 15.5.1 Laboratory Findings 15.5.2 High Turnover Disease 15.6 Low-Turnover Disease 15.6.1 Aluminum Control	211 211 212 212 214 215 216 216 217 218 218 218 218 221
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy 15.4 Metastatic Calcification 15.5 Laboratory Findings and Management of ROD 15.5.1 Laboratory Findings 15.5.2 High Turnover Disease	211 211 212 212 214 215 216 216 217 218 218 218 2124 224
15	Renal Osteodystrophy 15.1 Pathophysiology of Renal Osteodystrophy 15.1.1 Vitamin D Metabolism 15.1.2 Phosphorus Retention 15.1.3 Parathyroid Hormone 15.2 Histological Classification of Renal Osteodystrophy 15.2.1 High-Turnover Bone Disease 15.2.2 Low-Turnover Bone Disease 15.2.3 Mixed (Uremic) Bone Disease 15.3 Clinical Manifestation of Renal Osteodystrophy 15.4 Metastatic Calcification 15.5 Laboratory Findings and Management of ROD 15.5.1 Laboratory Findings 15.5.2 High Turnover Disease 15.6 Low-Turnover Disease 15.6.1 Aluminum Control 15.6.2 Low Parathyroid Hormone	211 211 212 212 214 215 216 216 217 218 221 224 224 224

Contents xix

16	Atypical Dialysis Circumstances	227
	16.1 Pregnancy	
	16.1.1 Dialysis	227
	16.1.2 Associated Conditions	229
	16.2 Drug Removal in Overdose Situations	229
	16.2.1 Peritoneal Dialysis	230
	16.2.2 Hemodialysis	230
	16.2.3 Hemoperfusion	230
	16.2.4 Specific Examples	
	Reference	
17	The Future	233
	17.1 Increasing Financial Pressure	233
	17.2 Changing Population	233
	17.3 Patient Outcome Measures	233
	17.4 Limited Transplantation Options	234
	17.5 Low Rates of Rehabilitation	234
	17.6 Ideal Renal Replacement Therapy	234
	17.7 Simpler Machines and Daily Dialysis	235
	17.7.1 Diffusion vs Convection	
	17.8 Mechanical Artificial Kidney	235
	17.8.1 Implantable Mechanical, Artificial Kidney	
	Reference	
Ind	ex	239

Abbreviations

AII Angiotensin II

AAMI Association for the Advancement of Medical Instrumentation

ACE Angiotensin-converting enzyme

ACT Activated clotting time

AMAC Arm muscle area circumference

AN6 Acrylonitrile-6

ARB Angiotensin (II) receptor blockers

AV Arteriovenous

AVP Arginine-vasopressin

BIA Bioelectric impedance analysis

BP Blood pressure
BSA Body surface area
BUN Blood urea nitrogen
(i)Ca (Ionized) calcium

CAAPD Continuous automated ambulatory peritoneal dialysis

CAPD Continuous ambulatory peritoneal dialysis
CAVH Continuous arteriovenous hemofiltration
CAVHD Continuous arteriovenous hemodiafiltration

CCB Calcium channel blocker

CCPD Continuous cycling peritoneal dialysis CHF (Slow and) continuous hemofiltration

CTS Carpal tunnel syndrome

CVVH Continuous venovenous hemofiltration
CVVHD Continuous venovenous hemodiafiltration

DDS Dialysis disequilibrium syndrome DEXA Dual-energy X-ray absorptiometry

DFO Deferoxamine DI Dialysis index

DOQI Dialysis Outcome Quality Initiative

DPI Dietary protein intake ECF Extracellular fluid

xxii Abbreviations

ESRD End-stage renal disease

Epo Erythropoietin
Eto Ethylene oxide
FBV Fiber bundle volume
GFR Glomerular filtration rate

GU Glucose uptake

HCO₃ Hydrogen bicarbonate

Hct Hematocrit HD Hemodialysis HDF Hemodiafiltration

HDL High-density lipoprotein

HF Hemofiltration ICF Intracellular fluid

IHF Intermittent hemofiltration
IDPN Intradialytic parenteral nutrition

IJ Internal jugular

IPD Intermittent peritoneal dialysis
IUF Intermittent ultrafiltration

K_{urea} Urea clearance

 $\begin{array}{lll} \mbox{(e)Kt/V} & \mbox{(Equilibrated) dose of dialysis} \\ \mbox{KoA} & \mbox{Mass transfer coefficient} \\ \mbox{K}_{ru} & \mbox{Residual renal urea clearance} \\ \mbox{K}_{uf} & \mbox{Ultrafiltration coefficient} \\ \end{array}$

kd Kilodalton

LDL Low-density lipoprotein
LMWH Low molecular weight heparin
LVH Left ventricular hypertrophy
MAK Mechanical artificial kidney
MCV Mean corpuscular volume

MM Middle molecule

MMHD Morbidity in Maintenance Hemodialysis Study

NCDS National Co-operative Dialysis Study NIPD Nocturnal intermittent peritoneal dialysis

NKF National Kidney Foundation

NO Nitric oxide

NTx Cross-linked N-terminal telopeptide of type I collagen

PAN Polyacrylonitrile

(n)PCR (Normalized) protein catabolic rate pClCr Peritoneal creatinine clearance pCl_{urea} Peritoneal urea clearance

PD Peritoneal dialysis

PET Peritoneal equilibration test

PGI₂ Prostacyclin

PICP Procollagen type I C-terminal peptide

pKt Peritoneal urea clearance rate

Abbreviations xxiii

PMMA Polymethylmethacrylate

(n)PNA (Normalized) protein equivalent of nitrogen appearance rate

PO₄ Phosphate

PTFE Polytetrafluoroethylene
PTH Parathyroid hormone
PTT Prothrombin time
PTX Parathyroidectomy

pre-BUN Predialysis concentration of blood urea nitrogen post-BUN Postdialysis concentration of blood urea nitrogen

PS Polysulfone
PV Plasma volume
Qb Blood flow rate
Qd Dialysate flow rate
RBC Red blood cell

rClCr Renal creatinine clearance rClU Renal urea clearance RO Reverse osmosis ROD Renal osteodystrophy Renal replacement therapy RRT **SCUF** Slow and continuous UF Subjective global assessment SGA Slow low efficiency dialysis **SLED**

SM Small molecule

SUF Sequential ultrafiltration and dialysis

t Time

TBW Total body water

TIBC Total iron-binding capacity
TMP Transmembrane pressure
TPN Total parenteral nutrition
TPD Tidal peritoneal dialysis
TPR Total peripheral resistance
TSFT Triceps skin fold thickness

UF(R) Ultrafiltration (rate)
UKM Urea kinetic modeling
UNA Urea/nitrogen appearance
URR Urea reduction ratio

V Volume of distribution of body fluid

VDR Vitamin D receptors

V_{urea} Volume of distribution of urea

WBC White blood cell

W(p)ClCr Weekly (peritoneal) creatinine clearance

Chapter 1 Brief History of Clinical Dialysis: The Seattle Experience

Although it was not until the 1960s that long-term dialysis in a clinical setting became a reality, dialysis as a treatment for renal failure had been the focus of interest for some time. By the end of the 1950s, Dr. B. H. Scribner had established an acute dialysis program at the University of Washington. In 1960, a uremic comatosed man who was thought to have acute renal failure was brought back to almost normal active life with intermittent hemodialysis. However, he was found to have chronic irreversible renal disease and had to be sent home to die; it became clear to the Seattle team that if long-term vascular access could be maintained, long-term dialysis would become a reality. This led to the development of the Scribner Shunt and the advent of chronic hemodialysis.

The Seattle team developed an entire program to care for a population of patients who had a chronic disease and who were being kept alive on a new form of treatment. New equipment and systems were developed and refined and solutions for unexpected problems had to be devised—specifically, treatment of hyperphosphatemia, renal osteodystrophy, and hypertension. To make the treatment more practical, by reducing the bulk of the dialysate through the use of concentrated dialysate, a proportioning system had to be developed and a substitute for bicarbonate was used to prevent the precipitation of calcium carbonate. This was achieved by using acetate. However, when acetate-related problems started to appear (due to the use of more efficient dialyzers, in the mid-1970s), a double proportioning system was developed to enable the use of bicarbonate again. As is often the case, the resolution of one problem often led to other unexpected difficulties. The commitment and ingenuity of the pioneers of dialysis treatment, however, meant that these hurdles were overcome and the success of dialysis as a treatment for end-stage renal disease (ESRD) was assured. Later in the 1980s, another Scribner fellow, Joseph Eschbach, developed and used recombinant erythropoietin, and anemia-related issues became history. The pioneering work continues today; the most recent modification in dialysate was the development of a citric acid-based acid concentrate for dialysate. This is proving to be more beneficial to the patients than the currently used acetic acid-based acid concentrate.

The shortage of resources in the early days of dialysis necessitated the founding of a patient selection committee to decide which of the needy patients would be accepted into the program. This committee (thought by many to be the foundation for the development of medical ethics) forced several actions with far-reaching consequences, one of which was the development of home dialysis.

A young high-school student was found to have ESRD but was not accepted for dialysis by the patient selection committee. The team decided that home dialysis was a viable alternative if they could develop a smaller hemodialysis machine that could be used at home. The collaborative effort of Dr. Scribner's clinical team and the engineering team of Dr. Albert L. Babb succeeded in building a home hemodialysis machine in only 3 months. This home machine became the prototype of machines in use currently.

In early 1960s, Dr. Fred Boen joined the Seattle group and began treating a patient using peritoneal dialysis (PD), with a closed system containing 20-l (and later 40-l) bottles. Henry Tenckhoff, a research fellow with Dr. Boen, treated patients at home using Boen's repeated puncture technique. This technique, however, required aseptic access to the peritoneal cavity with a catheter each time dialysis was needed, and meant that Dr. Tenckhoff had to visit each patient's home at least three times a week to insert the access device. Eventually, Dr. Tenckhoff developed the indwelling peritoneal catheter and a sterile technique for its insertion, which made it possible to use the new form of dialysis on a larger scale.

A detailed analysis of the Seattle experience with intermittent PD (IPD) revealed the potential risk of under-dialysis and poor "technique survival rates" [1], suggesting that the dialysis dose needed to be increased. In 1965, Dr. Robert Popovich while in Seattle had become involved in the kinetics of the "middle molecule" across the peritoneal membrane before moving to Texas and becoming a pioneer of the continuous ambulatory PD (CAPD) technique. This continuous therapy improved the dialysis dose and made PD a viable technique of renal replacement therapy (RRT).

Encountering a patient who was dying of malnutrition due to bowel disease, Dr. Scribner saw an opportunity to apply the group's expertise in vascular access to another area of medicine. The development of Broviac (and later on Hickman) catheters and the "total parenteral nutrition" (TPN) program (operated by the nephrology team at the University of Washington) was a result of the vision and dedication of Dr. Scribner and his co-workers.

This very brief account of the Seattle experience shows that the commitment of Dr. Scribner, his team, their collaborators, and community members accomplished more than the development of a dialysis access device. Their efforts led to the development of systems for dialysis, central venous catheters, parenteral nutrition, long-term care of ESRD patients, community-based dialysis centers, home dialysis programs, an early concept of dialysis dose calculation, and continued technological improvement. The development of the dialysis program established nephrology as a subspecialty and has also had far-reaching implications in the fields of bowel disease, organ transplantation, oncology, and for all acutely ill patients. It is now

difficult to imagine that less than 50 years ago, patients with ESRD had only one prognosis—death—and that patients with renal failure were connected to patients with liver failure so that each could be kept alive by the healthy organ of the other.



1.1 Definition of Dialysis

In broad terms, the process of dialysis involves bidirectional movement of molecules across a semipermeable membrane. Clinically, this movement takes place in and out of blood, across a semipermeable membrane. If the blood is exposed to an artificial membrane outside of the body, the process is called hemodialysis (HD) or hemofiltration (HF). If the exchange of molecules occurs across the peritoneal membrane, the process is called peritoneal dialysis (PD).

1.2 Mechanisms Involved in Molecular Movement

The movement of molecules follows certain physiological and physicochemical principles that are outlined below (see Fig. 1.1a).

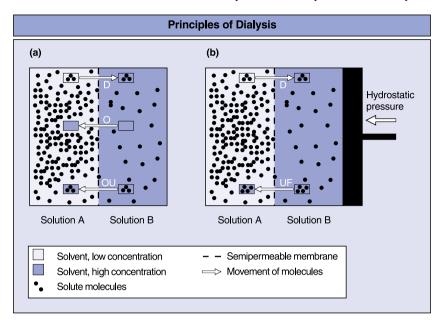


Fig. 1.1 a Diffusion, osmosis, and osmotic ultrafiltration by osmotic pressure. b Hydrostatic ultrafiltration. D diffusion, O osmosis, OU osmotic ultrafiltration, UF ultrafiltration by hydrostatic pressure, C convection

1.2.1 Diffusion

If two solutions of different concentrations are separated by a semipermeable membrane, solute will move from the side of higher to the side of lower solute concentration. This process of solute movement on a concentration gradient is called *diffusion* and is caused by the random movement of the solute molecules striking and moving across the membrane. Several factors influence this random movement and thus the rate of diffusion. The transport of any solute or solvent molecule is dependent on the *physical size of the molecule* relative to the size of the pores in the membrane. Any molecules larger than the pores of the membrane cannot pass through. Similarly, the *electrical charge* and the *shape* of the molecule also determine the rate of transport across the membrane. If the membrane has a negative charge, particles with a like charge will have limited transport as compared with those with a positive or a neutral charge.

1.2.2 Ultrafiltration

A solvent such as water can be forced across a semipermeable membrane on a pressure gradient, from higher to lower pressures (see Fig. 1.1). The pressure could be a result of osmotic force (see below) or of mechanical hydrostatic pressure. The

1.3 Clearance 5

solvent carries with it the dissolved solute molecules small enough to pass through the membrane pores (see below). This movement of molecules across a semipermeable membrane, caused by a pressure difference, is called *ultrafiltration* (UF). If the pressure is hydrostatic, the process is called "hydrostatic UF." Conversely, the UF caused by osmotic pressure is called "osmotic UF."

1.2.3 Osmosis

As solute concentration increases, solvent concentration correspondingly decreases and vice versa. If a semipermeable membrane separates solutions of different concentrations, solvent along with dissolved small solutes will flow from the side with the higher solvent concentration to the side with the lower solvent concentration. This process is called *osmosis* (see Fig. 1.1).

1.2.4 Convection

As solvent molecules move on a pressure gradient, the dissolved solute molecules are dragged along (solvent drag); this process of solute movement is called *convection*. The ease with which the solute is dragged along is determined by the size of the solute molecule relative to the size of the membrane pores. Smaller solutes are transported easily and the entire solution can sieve across the membrane without any change in concentration. In contrast, larger solutes move more slowly and the rate of convective transport is slower. Thus, the convective transport of a solute depends on the porosity of the membrane. This porosity, known as the "sieving coefficient of the membrane," can be calculated by dividing the concentration of solute on side A by the concentration on side B.

1.3 Clearance

In a clinical setting, the removal of a solute is measured in terms of clearance, the term being defined as the volume of blood or plasma from which the solute is completely removed in unit time. Let us assume that the blood urea concentration across a hemodialyzer drops from 100 mg/dl at the inlet to 10 mg/dl at the outlet. This 90% decline represents the diffusion of urea from blood into the dialysate and depends largely on the concentration gradient between these fluids. However, the magnitude of the "cleaning" of blood also depends on blood flow rates (Qb). Thus, in the above example, a blood flow rate of 100 ml/min means that 90 ml of the blood was cleared of urea. However, for a blood flow of 200 ml/min, 180 ml of blood is cleared of urea each minute (see the example below for a more accurate calculation). Clearance measures the magnitude of blood cleaning, independent of the concentration of the solute entering the dialyzer.

1.3.1 Blood vs Plasma Clearance

During transit across the dialyzer, most solutes are removed from plasma water (about 93% of blood volume, depending on plasma protein concentration). If the solute is not in the blood cells or if the movement of solute out of these cells is slow, the clearance of the solute decreases as the hematocrit increases (since the plasma volume decreases). Urea is often used as a solute to measure dialysis efficiency (it is present in plasma water as well as in erythrocytes), and the flux of urea across the erythrocyte membrane is reasonably fast. This means that urea is cleared from whole blood during dialysis and is not affected greatly by the hematocrit. The following example clarifies these concepts:

Example

 $Qb = 200 \,\text{ml/min}$, hematocrit = 35%

Plasma flow rate = $200 \text{ml/min} \times (1 - 0.35) = 130 \text{ml/min}$

Plasma water flow rate = $130 \,\text{ml/min} \times 0.93 \,(93\% \,\text{of plasma is water}) =$

121 ml/min

Erythrocyte flow rate = $200 \,\text{ml/min} - 130 \,\text{ml/min} = 70 \,\text{ml/min}$

Erythrocyte water flow rate = $70 \,\text{ml/min} \times 0.80$ (about 80% of erythrocyte volume is water [containing diffusible urea]) = $56 \,\text{ml/min}$

Thus, the whole blood water flow rate effective for urea clearance = $121 \,\text{ml/min} + 56 \,\text{ml/min} = 177 \,\text{ml/min}$

If the blood water concentration of urea = $100 \,\text{mg/dl}$ at dialyzer inlet and $10 \,\text{mg/dl}$ at outlet, the urea clearance of whole blood = $177 \,\text{ml/min} \times \{1 - [(10 \,\text{mg/dl})/(100 \,\text{mg/dl})]\} = 159 \,\text{ml/min}$

This means that 159 ml of blood is cleared of urea each minute.

1.3.2 Clinical Factors Influencing Dialysis Urea Clearance

The three major determinants of urea clearance during hemodialysis are:

Blood flow rate (Qb)

Dialysate flow rate (Qd)

Membrane (dialyzer/peritoneal membrane) efficiency

Reference

 Ahmad S, Gallagher N, Shen F. Intermittent peritoneal dialysis: status reassessed. Trans Am Soc Artif Intern Organs 1979, 25:86–89.

Chapter 2 Hemodialysis Technique

As discussed in the previous chapter, the clearance of a solute is dependent on the Qb, Qd, and membrane efficiency. The dialyzer membranes have different pore sizes that are variably distributed, larger pores being fewer than smaller pores. Small solutes like urea can be transported through all pore sizes whereas the larger molecules such as vitamin B12 or beta-2-microglobulin can only pass through the larger pores. Thus the clearance of the larger solutes, unlike urea, is more influenced by the membrane and less by Qb and Qd.

2.1 Blood Flow Rate

Because clearance is calculated using Qb, it would be understandable to mistakenly assume that the relationship between urea clearance and Qb is linear. However, although urea clearance increases steadily as Qb is increased from zero, at faster flow rates, the dialyzer is unable to continue to transport urea with the same efficiency and the urea concentration at the dialyzer outlet increases. In other words, the urea removed as a percentage of urea inflow into the dialyzer decreases and (as clearance is Qb multiplied by the fractional decline in urea) the clearance curve plateaus (see Fig. 2.1).

2.2 Dialysate Flow Rate

An increase in Qd generally increases the urea clearance. This effect is negligible, however, as long as Qd is $150-250 \,\text{ml/min}$ faster than Qb. With high-efficiency dialyzers, there is little (<10%) increase in urea clearance if Qd is increased from $500 \,\text{ml/min}$, provided that Qb remains $350 \,\text{ml/min}$.