Circulating Nucleic Acids in Plasma and Serum

Peter B. Gahan Editor

# Circulating Nucleic Acids in Plasma and Serum

Proceedings of the 6th International Conference on Circulating Nucleic Acids in Plasma and Serum Held on 9–11 November 2009 in Hong Kong



*Editor* Prof. Peter B. Gahan King's College London Anatomy & Human Sciences London Bridge SE1 1UL London United Kingdom pgahan@aol.com

ISBN 978-90-481-9381-3 e-ISBN 978-90-481-9382-0 DOI 10.1007/978-90-481-9382-0 Springer Dordrecht Heidelberg London New York

Library of Congress Control Number: 2010933647

© Springer Science+Business Media B.V. 2011

No part of this work may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying, microfilming, recording or otherwise, without written permission from the Publisher, with the exception of any material supplied specifically for the purpose of being entered and executed on a computer system, for exclusive use by the purchaser of the work.

Printed on acid-free paper

Springer is part of Springer Science+Business Media (www.springer.com)

# Contents

Part I Current Developments

1	Current Developments in Circulating Nucleic Acids in Plasma and Serum	3
Part	t II Nucleic Acids in Oncology – Diagnosis and Prognosis and Metastases	
2	Reflections on a Life of CNAPS: From Circulating DNAto the VirtosomeMaurice Stroun	15
3	Circulating Tumor-Related DNA Alterations as Prostate Cancer Biomarkers	21
4	Parallel Tagged 454 Sequencing for the Characterizationof Circulating DNAManiesh van der Vaart, Dmitry V. Semenov, Elena V. Kuligina,Vladimir A. Richter, and Piet J. Pretorius	29
5	Advanced Analysis of Human Plasma Circulating DNA Sequences Produced by Parallel Tagged Sequencing on the 454 Platform	35
6	Concentration and Distribution of Single-Copy β-ActinGene and LINE-1 Repetitive Elements in Blood of LungCancer PatientsCancer PatientsAnastasia A. Ponomaryova, Elena Y. Rykova,Nadezhda V. Cherdyntseva, Tatiana E. Skvortsova,Anna V. Cherepanova, Evgeniy S. Morozkin,Vladislav A. Mileiko, Nikolai V. Litvjakov,Alexey Y. Dobrodeev, Alexander A. Zav'yalov,Sergey A. Tuzikov, Elena D. Chikova, Valentin V. Vlassov,and Pavel P. Laktionov	41

Contents

7	Plasma DNA Methylation Analysis in PredictingShort-Term Recurrence of Surgical Patients with Non-smallCell Lung Cancer (NSCLC)Qingqing Ding, Yuan Mu, Shiyang Pan, Yongqian Shu,Shijiang Zhang, Bingfeng Zhang, Hong Wang, Li Gao,Wenying Xia, Jian Xu, Meijuan Zhang, Yuanyuan Zhang,Yan Cao, and Shan Lu	47
8	Blood Based Methylated DNA and Tumor-Specific Protein Analysis in Gastric Cancer Diagnostics Elena V. Elistratova, Petr I. Shelestyuk, Valentina I. Permyakova, Elena D. Chikova, Sergey A. Tuzikov, Valentin V. Vlassov, Pavel P. Laktionov, and Elena Y. Rykova	57
9	Increase in Circulating MicroRNA Levels in Blood of Ovarian Cancer Patients	63
10	The Course of Circulating Nucleosomes in Liver CancerPatients Undergoing Transarterial Chemoembolization TherapyNikolaus Kohles, Dorothea Nagel, Dietrich Jüngst,Jürgen Durner, Petra Stieber, and Stefan Holdenrieder	73
11	<b>Presence of Nucleosomes in Cerebrospinal Fluid</b> <b>of Glioblastoma Patients – Potential for Therapy Monitoring</b> Stefan Holdenrieder, Andreas Spuler, Michael Tischinger, Dorothea Nagel, and Petra Stieber	79
12	Circulating Nucleosomes and DNAse in Breast Cancer Patients During Neoadjuvant Chemotherapy	85
13	Circulating Nucleosomes in Cancer Patients with Liver Metastases Undergoing Selective Internal Radiation Therapy Using Yttrium-90 Labelled Microspheres	91
14	H3K9me3/H4K20me3 Ratio in Circulating Nucleosomes as Potential Biomarker for Colorectal Cancer	97

Contents
----------

15	Functionality of CNAPS in Cancer: The Theory of Genometastasis Dolores C. García-Olmo, Hector Guadalajara, Carolina Dominguez-Berzosa, María G. Picazo, Mariano García-Arranz, and Damián García-Olmo	105
Par	t III Nucleic Acids in Foetal Medicine	
16	Circulating Fetal DNA/RNA in Maternal Plasma for Aneuploidy Detection	111
17	A "Fluid-Agnostic" Approach to Analysis of Fetal and Neonatal Developmental Gene Expression	125
18	Non-invasive Prenatal Diagnosis: An Epigenetic Approach to the Detection of Common Fetal Chromosome Disorders by Analysis of Maternal Blood Samples	133
19	<b>Comparative Study of Extracellular DNA by FISH</b> Evgeniy S. Morozkin, Ekaterina M. Loseva, Vladislav A. Mileiko, Kira S. Zadesenets, Nikolay B. Rubtsov, Valentin V. Vlassov, and Pavel P. Laktionov	143
20	An Additional Pre-amplification Step for the EarlyDetermination of Fetal RHD from Maternal PlasmaTadeja Dovč-Drnovšek, Nataša Toplak, Irena Bricl,Tanja Blejec, Minka Kovač, and Primož Rožman	147
21	The Correlation of Circulating Cell-Free DNA, Cell-Free Fetal DNA and MicroRNA 325 Levels to Clinical Characteristics and Laboratory Parameters in Pre-eclampsia Levente Lázár, Bálint Nagy, Attila Morvarec, and János Rigó	153
Par	t IV Other Clinical Exploitation of CNAPS	
22	Comparison of Plasma Cell-Free DNA Levels with Gene Expression Profiles of Peripheral Blood Cells During Haemodialysis	159
23	Low-Molecular-Weight DNA of Blood Plasma as an Indicator of Pathological Processes	165

Contents
----------

24	The Clinical Significance of Plasma DNA Quantification   for Quake Trauma Patients	171
	Dan Chen, Shiyang Pan, Shijiang Zhang, Peijun Huang, Wenying Xia, Erfu Xie, Bing Gu, Fang Wang, Jian Xu, Ting Xu, Yachun Lu, Di Yang, and Shan Lu	
Par	t V The Biology of CNAPS	
25	Methylated Cell-Free DNA In Vitro and In Vivo	185
26	Circadian Rhythmicity and Clearance of Cell-Free DNA in Human Plasma	195
27	Fragments of Cell-Free DNA (cfDNA) Enhance Transcription Activity in Human Mesenchymal Stem Cells (hMSCs) and Inhibit Their In Vitro Differentiation Elena M. Malinovskaya, Svetlana V. Kostyuk, Aleksey V. Ermakov, Marina S. Konkova, Tatjana D. Smirnova, Larisa V. Kameneva, Liudmila V. Efremova, Anna Yu. Alekseeva, Liudmila N. Lyubchenko, and Natalya N. Veiko	199
28	Cell-Surface-Bound DNA Inhibits Poly(I:C)-Activated IL-6 and IL-8 Production in Human Primary Endothelial Cells and Fibroblasts	207
29	Accumulating Fragments of Extracellular DNA (ecDNA) Influence Rat Primary Cerebellum Granule Cell Culture Liudmila V. Efremova, Svetlana V. Kostyuk, Leonid G. Khaspekov, and Natalya N. Veiko	213
30	Cell Free DNA (cfDNA) Influences Nitric Oxide and ros Levels in Human Endothelial Cells	219
31	Development of the Adaptive Response and Bystander Effect Induced by Low-Dose Ionising Radiation in Human Mesenchymal Stem Cells	225

Contents

32	<b>Extracellular RNA as Regulators of Cellular Processes</b> Dmitry V. Semenov, Grigory A. Stepanov, Dmitry N. Baryakin, Olga A. Koval, Elena V. Kuligina, and Vladimir A. Richter	233
33	Microvesicles Circulating in Plasma of Rats Contain DNA: Are These Small Vesicles a Main Source of Cell-Free DNA in Plasma?	239
Par	t VI New Technologies for CNAPS	
34	Rapid Isolation and Detection of Cell Free CirculatingDNA and Other Disease Biomarkers Directly from Whole BloodRajaram Krishnan and Michael J. Heller	247
35	Yields of Viral and Circulating Cell-Free Nucleic Acids Using the QIAamp <sup>®</sup> Circulating Nucleic Acid Kit Martin Horlitz, Tanja Hartinger, Simone Graf, Annabelle Lucas, Annette Nocon, and Markus Sprenger-Haussels	259
36	Comparison of Nucleosomes and Quantitative PCR Using Diverse DNA Isolation Methods	269
37	MicroRNA Analysis in the Spinal Fluid of Alzheimer Patients: A Methodological Feasibility Study Argonde van Harten, Joyce Mulders, Cagla Çevik, Maartje Kester, Philip Scheltens, Wiesje van der Flier, and Cees Oudejans	275
Ind	ex	283

## Contributors

Elif Z. Akisik Department of Basic Oncology, Istanbul University Oncology Institute, Istanbul, Turkey, elifakisik@yahoo.com

**Ebru E. Akisik** Department of Basic Oncology, Istanbul University Oncology Institute, Istanbul, Turkey, ebruakisik@yahoo.com

Anna Yu. Alekseeva Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, ribgene@rambler.ru

**Dmitry N. Baryakin** Institute of Chemical Biology and Fundamental Medicine SB RAS, Novosibirsk, Russia, dimabiolog@mail.ru

**Diana W. Bianchi** Division of Genetics, Department of Pediatrics, Floating Hospital for Children at Tufts Medical Center, Boston, MA, USA; Division of Genetics, Department of Pediatrics, Floating Hospital for Children at Tufts Medical Center, Boston, MA, USA, DBianchi@Tuftsmedicalcenter.org

**Nikola Bila** Faculty of Medicine in Pilsen, Charles University in Prague, Pilsen, Czech Republic

Tanja Blejec Department of Perinatology, University Medical Centre, Ljubljana, Slovenia

Irena Bricl Blood Transfusion Centre of Slovenia, Ljubljana, Slovenia

**Olga E. Bryzgunova** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, olga.bryzgunova@niboch.nsc.ru

**Dursun Bugra** Department of Surgery, Istanbul Medical Faculty, Istanbul University, Istanbul, Turkey, dbugra@doruk.net.tr

Natalia V. Bulycheva Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, nbulicheva@mail.ru

Alexander V. Bushuev Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia Yan Cao Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, njmucaoyan@126.com

Nigel Carter Wellcome Trust Sanger Institute, Cambridge, UK

**Dalibor Cerny** 1st School of Medicine, Charles University, Prague, Czech Republic, Dalibor.Cerny@seznam.cz

**Cagla Çevik** Department Clinical Chemistry, VU University Medical Center, De Amsterdam, The Netherlands

**Dan Chen** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, lab.med@163.com

Nadezhda V. Cherdyntseva Siberian Division of the Russian Academy of Medical Sciences, Cancer Research Institute, Tomsk, Russia, nvch@oncology.tomsk.ru

**Anna V. Cherepanova** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, a\_cher@niboch.nsc.ru

**Elena D. Chikova** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, labor@cnmt.ru

**R.W.K. Chiu** Centre for Research into Circulating Fetal Nucleic Acids, Li Ka Shing Institute of Health Sciences, and Department of Chemical Pathology, Prince of Wales Hospital, The Chinese University of Hong Kong, Shatin, Hong Kong SAR, China, Rossachiu@cuhk.edu.hk

**Maurizio D'Esposito** Institute of Genetics and Biophysics 'A. Buzzati Traverso', CNR, Naples, Italy, desposit@igb.cnr.it

**Nejat Dalay** Department of Basic Oncology, Istanbul University Oncology Institute, Istanbul, Turkey, ndalay@yahoo.com

**Ugur Deligezer** Department of Basic Oncology, Istanbul University Oncology Institute, Istanbul, Turkey, ugur\_deligezer@yahoo.com

**Qingqing Ding** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China; Department of Oncology, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, helen.jsnj@yahoo.com.cn

Alexey Y. Dobrodeev Siberian Division of the Russian Academy of Medical Sciences, Cancer Research Institute, Tomsk, Russia, dobrodeev@sibmail.com

**Carolina Dominguez-Berzosa** IdiPAZ, "La Paz" University Hospital, Universidad Autónoma de Madrid, Madrid, Spain, cdominguez.hulp@salud.madrid.org

**Tadeja Dovč-Drnovšek** Blood Transfusion Centre of Slovenia, Ljubljana, Slovenia, tadeja.dovc-drnovsek@ztm.si

### Contributors

Jürgen Durner Institute of Clinical Chemistry, University-Hospital Munich-Grosshadern, Munich, Germany, juergen.durner@med.uni-muenchen.de

Liudmila V. Efremova Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, labmolbiol@gmail.com

**Elena V. Elistratova** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, alenakol@mail.ru

Aleksey V. Ermakov Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, avePlato@mail.ru

**Nilgün Erten** Department of Internal Medicine, Istanbul Medical Faculty, Istanbul University, Istanbul, Turkey, snilgunerten@yahoo.com

**Yvonne Fehr** Institute of Clinical Chemistry, University-Hospital Munich-Grosshadern, Munich, Germany, Yvonne.Fehr@med.uni-muenchen.de

**Debora M.I. Fersching** Institute of Clinical Chemistry, University Hospital Munich, Munich, Germany, Debora.Fersching@med.uni-muenchen.de

**Michael Fleischhacker** Medical Clinic – Oncology and Haematology, University Medicine Charité Berlin, Berlin, Germany, Michael.Fleischhacker@charite.de

**Peter B. Gahan** Anatomy and Human Sciences, King's College London, London, UK, pgahan@aol.com

Li Gao Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, glkorea@163.com

Mariano García-Arranz IdiPAZ, "La Paz" University Hospital, Universidad Autónoma de Madrid, Madrid, Spain, mgarciaa.hulp@salud.madrid.org

**Damián García-Olmo** IdiPAZ, "La Paz" University Hospital, Department of Surgery, Universidad Autónoma de Madrid, Madrid, Spain, damian.garcia@uam.es

**Dolores C. García-Olmo** Experimental Research Unit, General University Hospital of Albacete, Albacete, Spain, doloresg@sescam.jccm.es

Simone Graf R&D Department, QIAGEN GmbH, Hilden, Germany, simone.graf@qiagen.com

**Bing Gu** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, gb20031129@163.com

Hector Guadalajara IdiPAZ, "La Paz" University Hospital, Universidad Autónoma de Madrid, Madrid, Spain, hector.guadalab@salud.madrid.org

**Tanja Hartinger** R&D Department, QIAGEN GmbH, Hilden, Germany, tanja.hartinger@qiagen.com

**Michael J. Heller** Department of Bioengineering, Department of Nanoengineering, UCSD Moores Cancer Center, University of California San Diego, La Jolla, CA, USA, mheller@ucsd.edu

Martin Heubner Department of Gynaecology and Obstetrics, University of Duisburg-Essen, Essen, Germany, martin.heubner@uk-essen.de

**Ralf-Thorsten Hoffmann** Institute of Clinical Radiology, University-Hospital Munich-Grosshadern, Munich, Germany, rthoffma@med.uni-muenchen.de

**Stefan Holdenrieder** Institute of Clinical Chemistry, University-Hospital Munich-Grosshadern, Munich, Germany, Stefan.Holdenrieder@med.uni-muenchen.de

**Dave S.B. Hoon** Department of Molecular Oncology, John Wayne Cancer Institute, Santa Monica, CA, USA, hoon@jwci.org

Ales Horinek 1st School of Medicine, Charles University, Prague, Czech Republic; General Teaching Hospital, Prague, Czech Republic, ahor@lf1.cuni.cz

Martin Horlitz R&D Department, QIAGEN GmbH, Hilden, Germany, martin.horlitz@qiagen.com

**Peijun Huang** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, hpj63@163.com

**Maj A. Hultén** Warwick Medical School, University of Warwick, Coventry, UK, maj.hulten@warwick.ac.uk

**Tatyana V. Ivtchik** St-Petersburg State Medical University named after I. P. Pavlov, St-Petersburg, Russia, ivtchikt@mail.ru

**Tobias Jakobs** Institute of Clinical Radiology, University-Hospital Munich-Grosshadern, Munich, Germany, Tobias.Jakobs@med.uni-muenchen.de

**Dietrich Jüngst** Medical Clinic II, University-Hospital Munich-Grosshadern, Munich, Germany

Larisa V. Kameneva Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia

Sabine Kasimir-Bauer Department of Gynaecology and Obstetrics, University of Duisburg-Essen, Essen, Germany, sabine.kasimir-bauer@uk-essen.de

**Maartje Kester** Departments of Neurology, VU University Medical Center, Amsterdam, The Netherlands: Departments of Epidemiology/Biostatistics, VU University Medical Center, Amsterdam, The Netherlands

Leonid G. Khaspekov Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, labmolbiol@gmail.com, ribgene@rambler.ru

#### Contributors

Nikolaus Kohles Institute of Clinical Chemistry, University-Hospital Munich-Grosshadern, Munich, Germany, Nikolaus.Kohles@med.uni-muenchen.de

Marina S. Konkova Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, minozemceva@mail.ru

Marie Korabecna Department of Biology, Faculty of Medicine in Pilsen, Charles University in Prague, Pilsen, Czech Republic, marie.korabecna@lfp.cuni.cz

**Svetlana V. Kostyuk** Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, Svet-vk@yandex.ru

Minka Kovač Omega d.o.o., Ljubljana, Slovenia

**Olga A. Koval** Institute of Chemical Biology and Fundamental Medicine SB RAS, Novosibirsk, Russia, koval\_oa@niboch.nsc.ru

**Müge Kovancilar** Department of Basic Oncology, Istanbul University Oncology Institute, Istanbul, Turkey, mkvnclr@gmail.com

**Rajaram Krishnan** Department of Bioengineering, Department of Nanoengineering, UCSD Moores Cancer Center, University of California San Diego, La Jolla, CA, USA

**Elena V. Kuligina** Institute of Chemical Biology and Fundamental Medicine SB RAS, Novosibirsk, Russia, Kuligina@niboch.nsc.ru

**Pavel P. Laktionov** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, lakt@niboch.nsc.ru

**Levente Lázár** Department of Obstetrics and Gynecology, Semmelweis University, Budapest, Hungary, lazar\_levente@hotmail.com

Alena O. Lebedeva Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, alena.o.lebedeva@gmail.ru

Laurent Lessard Department of Molecular Oncology, John Wayne Cancer Institute, Santa Monica, CA, USA, LessardL@JWCI.ORG

**Gloria S. Leszinski** Institute of Clinical Chemistry, University Hospital Munich, Munich, Germany, Gloria.Leszinski@med.uni-muenchen.de

**Nikolai V. Litvjakov** Siberian Division of the Russian Academy of Medical Sciences, Cancer Research Institute, Tomsk, Russia, nvlitv72@sibmail.com

**Y.M.D. Lo** Centre for Research into Circulating Fetal Nucleic Acids, Li Ka Shing Institute of Health Sciences, and Department of Chemical Pathology, Prince of Wales Hospital, The Chinese University of Hong Kong, Shatin, Hong Kong SAR, China, loym@cuhk.edu.hk **Ekaterina M. Loseva** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, losevaem@mail.ru

Yachun Lu Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, luyachun121@yeah.net

Shan Lu Department of Medicine, University of Massachusetts Medical School, Worcester, MA, USA, Shan.Lu@umassmed.edu

Annabelle Lucas R&D Department, QIAGEN GmbH, Hilden, Germany, annabelle.lucas@qiagen.com

Liudmila N. Lyubchenko Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, admila@list.ru

Viktoria V. Mak Siberian Division of the Russian Academy of Sciences, Institute of Cytology and Genetics, Novosibirsk, Russia, mak@bionet.nsc.ru

**Elena M. Malinovskaya** Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, tigerilina@mail.ru

**Jill L. Maron** Division of Newborn Medicine, Department of Pediatrics, Floating Hospital for Children at Tufts Medical Center, Boston, MA, USA, jmaron@tuftsmedicalcenter.org

Vladislav A. Mileiko Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, mileiko@niboch.nsc.ru

Magdalena Mokrejsova General Teaching Hospital, Prague, Czech Republic; 1st School of Medicine, Charles University, Prague, Czech Republic, Magdalena.Mokrejsova@seznam.cz

**Evgeniy S. Morozkin** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, morozkin@niboch.nsc.ru

Attila Morvarec Department of Obstetrics and Gynecology, Semmelweis University, Budapest, Hungary

**Yuan Mu** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, muyuan629@163.com

**Joyce Mulders** Department Clinical Chemistry, VU University Medical Center, De Amsterdam, The Netherlands, j.mulders@vumc.nl

**Dorothea Nagel** Institute of Clinical Chemistry, University-Hospital Munich-Grosshadern, Munich, Germany, Dorothea.Nagel@med.uni-muenchen.de

**Bálint Nagy** Department of Obstetrics and Gynecology, Semmelweis University, Budapest, Hungary

Annette Nocon R&D Department, QIAGEN GmbH, Hilden, Germany, annette.nocon@qiagen.com

**Sylvie Opatrna** Faculty of Medicine in Pilsen, Charles University in Prague, Pilsen, Czech Republic, opatrna@fnplzen.cz

**Cees Oudejans** Department Clinical Chemistry, VU University Medical Center, De Amsterdam, The Netherlands, cbm.oudejans@vumc.nl

Shiyang Pan Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, sypan@njmu.edu.cn

Ales Panczak General Teaching Hospital, Prague, Czech Republic; 1st School of Medicine, Charles University, Prague, Czech Republic, apanc@lf1.cuni.cz

Klaus Pantel Institute of Tumour Biology, University Medical Centre Hamburg-Eppendorf, Hamburg, Germany, pantel@uke.uni-hamburg.de

**Elisavet A. Papageorgiou** Department of Cytogenetics and Genomics, The Cyprus Institute of Neurology and Genetics, Nicosia, Cyprus, eliza@cing.ac.cy

**Philippos C. Patsalis** Department of Cytogenetics and Genomics, The Cyprus Institute of Neurology and Genetics, Nicosia, Cyprus, patsalis@cing.ac.cy

Valentina I. Permyakova Siberian Division of the Russian Academy of Sciences, Central Clinical Hospital, Novosibirsk, Russia, v.i.permyakova@ngs.ru

María G. Picazo General University Hospital of Albacete, Albacete, Spain

Anastasia A. Ponomaryova Siberian Division of the Russian Academy of Medical Sciences, Cancer Research Institute, Tomsk, Russia, anastasia-ponomaryova@rambler.ru

**Piet J. Pretorius** Biochemistry Division, School for Physical and Chemical Sciences, North-West University, Potchefstroom, South Africa, Piet.pretorius@nwu.ac.za

**Floriana Della Ragione** Institute of Genetics and Biophysics 'A. Buzzati Traverso', CNR, Naples, Italy, dellarag@igb.cnr.it

**Vladimir A. Richter** Institute of Chemical Biology and Fundamental Medicine SB RAS, Novosibirsk, Russia, richter@niboch.nsc.ru

János Rigó Department of Obstetrics and Gynecology, Semmelweis University, Budapest, Hungary

**Katarina Rocinova** General Teaching Hospital, Prague, Czech Republic; 1st School of Medicine, Charles University, Prague, Czech Republic, rocinova@seznam.cz

**Carina Roth** Institute of Tumour Biology, University Medical Centre Hamburg-Eppendorf, Hamburg, Germany, c.roth@uke.uni-hamburg.de

Primož Rožman Blood Transfusion Centre of Slovenia, Ljubljana, Slovenia

**Nikolay B. Rubtsov** Siberian Division of the Russian Academy of Sciences, Institute of cytology and genetics, Novosibirsk, Russia, rubt@bionet.nsc.ru

**Elena Y. Rykova** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, rykova@niboch.nsc.ru

**Philip Scheltens** Departments of Neurology and Epidemiology/Biostatistics, VU University Medical Center, Amsterdam, The Netherlands

**Bernd Schmidt** Medical Clinic – Infectiology and Pulmonology, University Medicine Charité Berlin, Berlin, Germany, Bernd.Schmidt@medizin.uni-halle.de

**Heidi Schwarzenbach** Institute of Tumour Biology, University Medical Centre Hamburg-Eppendorf, Hamburg, Germany, hschwarz@uke.uni-hamburg.de

**Dmitry V. Semenov** Institute of Chemical Biology and Fundamental Medicine SB RAS, Novosibirsk, Russia, Semenov@niboch.nsc.ru

**Gemma Serrano-Heras** Experimental Research Unit, General University Hospital of Albacete, Albacete, Spain, gemmas@sescam.jccm.es

Petr I. Shelestyuk Novosibirsk Oncological Dispensary, Novosibirsk, Russia, pishelestuk@mail.ru

**Yongqian Shu** Department of Oncology, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, shuyongqian@csco.org.cn

**Barbara Siegele** Institute of Clinical Chemistry, University Hospital Munich, Munich, Germany, Barbara.Siegele@med.uni-muenchen.de

**Tatiana E. Skvortsova** Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, skvorts@niboch.nsc.ru

**Tatjana D. Smirnova** Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, ribgene@rambler.ru

Markus Sprenger-Haussels R&D Department, QIAGEN GmbH, Hilden, Germany, markus.sprenger-haussels@qiagen.com

Andreas Spuler Department of Neurosurgery, Helios Klinikum Berlin-Buch, Berlin, Germany, Andreas.Spuler@helios-kliniken.de

**Grigory A. Stepanov** Institute of Chemical Biology and Fundamental Medicine SB RAS, Novosibirsk, Russia, stepanov\_g@ngs.ru

**Petra Stieber** Institute of Clinical Chemistry, University-Hospital Munich-Grosshadern, Munich, Germany, Petra.Stieber@med.uni-muenchen.de

xviii

### Contributors

**Oliver J. Stoetzer** Hematology/Oncology Outpatient Specialty Center Munich, Munich, Germany, ojstoetzer@aol.com

Maurice Stroun OncoXL, Geneva, Switzerland, mauricestroun@bluewin.ch

**Eiji Sumami** Department of Molecular Oncology, John Wayne Cancer Institute, Santa Monica, CA, USA, Sunami-1su@h.u-tokyo.ac.jp

Klaus Tatsch Department of Nuclear Medicine, University-Hospital Munich-Grosshadern, Munich, Germany; Department of Nuclear Medicine, Municipal Hospital Karlsruhe, Karlsruhe, Germany, Klaus.Tatsch@klinikum-karlsruhe.de

**Vladimir Tesar** General Teaching Hospital, Prague, Czech Republic; 1st School of Medicine, Charles University, Prague, Czech Republic, Vladimir.Tesar@lf1.cuni.cz

Michael Tischinger Department of Psychiatry, University of Munich, Munich, Germany, m.tischinger@hochgrat-klinik.de

**Y.K. Tong** Centre for Research into Circulating Fetal Nucleic Acids, Li Ka Shing Institute of Health Sciences, and Department of Chemical Pathology, Prince of Wales Hospital, The Chinese University of Hong Kong, Shatin, Hong Kong SAR, China, yktong@cuhk.edu.hk

Nataša Toplak Omega d.o.o., Ljubljana, Slovenia

Sergey A. Tuzikov Siberian Division of the Russian Academy of Medical Sciences, Cancer Research Institute, Tomsk, Russia, tuzikovsa@oncology.tomsk.ru

**Wiesje van der Flier** Departments of Neurology, VU University Medical Center, Amsterdam, The Netherlands; Departments of Epidemiology/Biostatistics, VU University Medical Center, Amsterdam, The Netherlands; Alzheimer Center, VU University Medical Center, Amsterdam, The Netherlands, wm.vdflier@vumc.nl

Maniesh van der Vaart Biochemistry Division, School for Physical and Chemical Sciences, North-West University, Potchefstroom, South Africa, Manieshv@gmail.com

**Argonde van Harten** Departments of Neurology, VU University Medical Center, Amsterdam, The Netherlands; Departments of Epidemiology/Biostatistics, VU University Medical Center, Amsterdam, The Netherlands, a.vanharten@vumc.nl

Irina N. Vasilyeva St-Petersburg Scientific Research Institute of Phthisiopulmonology, St-Petersburg, Russia, nicolaivasiliev@hotmail.com

Natalya N. Veiko Research Centre for Medical Genetics, Russian Academy of Medical Sciences, Moscow, Russia, labmolbiol@gmail.com

Valentin V. Vlassov Siberian Division of the Russian Academy of Sciences, Institute of Chemical Biology and Fundamental Medicine, Novosibirsk, Russia, valentin.vlassov@niboch.nsc.ru **Igor A. Voznyuk** Military Medical Academy, St-Petersburg, Russia, voznjouk@yandex.ru

**Fang Wang** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, shywf74@sina.com

**Hong Wang** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, wanghong79@sohu.com

**Sabine Weickmann** Medical Clinic – Infectiology and Pulmonology, University Medicine Charité Berlin, Berlin, Germany, Sabine.Weickmann@charite.de

**Wenying Xia** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, xiawenying21106891@163.com

**Erfu Xie** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, xieerfu791010@163.com

**Jian Xu** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, xu\_jian79@163.com

**Ting Xu** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, xuting121@yeah.net

**Jian Xu** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China

**Di Yang** The Platform for Molecular Diagnosis and Biotherapy of Graveness Disease of Jiangsu Province, Nanjing, China, diyang@njmu.edu.cn

**Kira S. Zadesenets** Siberian Division of the Russian Academy of Sciences, Institute of cytology and genetics, Novosibirsk, Russia, kira\_z@bionet.nsc.ru

Alexander A. Zav'yalov Siberian Division of the Russian Academy of Medical Sciences, Cancer Research Institute, Tomsk, Russia, ZavyalovAA@oncology.tomsk.ru

**Shijiang Zhang** Department of Surgery, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, zhsj195177@yahoo.com.cn

**Bingfeng Zhang** Department of Surgery, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, bingfengzh2000@163.com

**Meijuan Zhang** Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, meijuan116@gmail.com

Yuanyuan Zhang Department of Laboratory Medicine, The First Affiliated Hospital of Nanjing Medical University, Nanjing, China, zhangyuanyuan121@163.com

# Part I Current Developments

### Chapter 1 Current Developments in Circulating Nucleic Acids in Plasma and Serum

Peter B. Gahan

**Abstract** DNA and RNA fractions have been isolated from the whole blood, serum, plasma, the surface of blood cells, urine, saliva and spinal fluid from both healthy individuals and patients. The ability to isolate, quantify, and analyze these molecules has led to the identification of specific nucleic acid fragments related to a variety of clinical disorders thereby permitting their early diagnosis and prognosis. This chapter summarizes the work reported in this volume.

Keywords Circulating nucleic acids  $\cdot$  Fetal medicine  $\cdot$  Oncology  $\cdot$  New technology  $\cdot$  Biology of CNAPS

### Introduction

The current volume concerns the meeting of the sixth international conference on circulating nucleic acids in plasma and serum (CNAPS) held in Hong Kong on 9-11 November 2009. The aim of the meeting was to bring together clinicians and scientists working in this field to present their latest findings on the basic biology, methodology and clinical applications of circulating nucleic acids in blood, urine, cerebro-spinal fluid and saliva.

Since the first publication by Mendel and Métais (1948) reporting the circulation of DNA in blood and its increase in amount in cancer patients, studies have evolved from just considering the amounts of DNA circulating in CNAPS during cancer and other clinical situations (Leon et al. 1977; Koeffler et al. 1973; Tan et al. 1966; Stroun et al. 1989) to more recent developments in the use of DNA, nucleosomes, mRNA and micro RNAs as both early markers and prognostic tools. Most work has concentrated upon the role of isolated DNA fractions and

P.B. Gahan (🖂)

Anatomy and Human Sciences, King's College London, London, UK e-mail: pgahan@aol.com

nucleosomes and to a lesser degree, mRNA. However, it is clear that the nucleic acids present in blood are derived from a number of sources including the breakdown of cells in the blood - both blood cells and circulating cancer cells, cell-surface bound DNA, the presence of bacteria and viruses, tissue necrosis, cell and tissue apoptosis, release of a newly synthesized DNA/RNA lipo-protein complex (the virtosome), exosomes, transposons and retrotransposons (Gahan and Stroun 2010). This medley of sources offers range of choice in the nucleic acid fraction to be assessed with respect to a particular disorder. In addition, the development of approaches involving the exploitation of epigenetic events such as methylation and hypermethylation, histone modifications in circulating nucleosomes, RNA-single nucleotide polymorphism, epigenetic allelic ratios and epigenetic-genetic chromosome dosage has offered more sensitive diagnostic methods that are applicable in the clinical environment. The following comments will highlight advances presented at the symposium and will raise questions as to the future developments needed.

# Nucleic Acids in Oncology – Diagnosis and Prognosis and Metastases

### **Diagnosis and Prognosis**

Two aspects of CNAPS in oncology have provided areas of development, namely, the clinical application in early diagnosis and prognosis and the other, a better understanding of the origins of metastases.

Nucleosomes have provided the basis for a number of studies in both early diagnosis and prognosis in cancer. Thus, trans-arterial chemo-embollization, the new loco-regional anticancer treatment option for advanced hepatocellular carcinoma patients, has been assessed in terms of serum nucleosome levels by Kohles et al. Although an initial decline was found in nucleosome levels shortly after treatment, by 24 hours there was a marked increase possibly due to the release of nucleosomes from the increased number of necrotic cells. Hence, this may provide a means of estimating the efficiency of the therapy. Likewise for the studies by Fehr et al. on nucleosome levels in patients after treatment by selective internal radiation therapy. This is a loco-regional anticancer treatment option for advanced cancer patients with liver metastases or liver cancer employing Yttrium-90 labelled microspheres.

A similar prognostic value might be available through the results of a preliminary investigation of nucleosome levels in both blood and cerebro-spinal fluid from patients with glioblastoma by Holdenreider et al. They showed that patients developing oedema after operation had a substantial increase in the nucleosome levels in the cerebo-spinal fluid. Additional studies may show this to be a useful marker of the development of post-operational complications.

As a variation on the nucleosome theme, Delizeger et al. examined a modification of the histone fraction of nucleosomes as a marker for colorectal cancer. Hence, an analysis of the trimethylation of histone H3 lysine 9 (H3K9me3) and histone H4 lysine 9 (H4K20me3), at the pericentric heterochromatin of the nucleosomes circulating in plasma, exploited the H3K9me3/H4K20me3 ratio for normalizing H3K9me3 concentrations. In this way, it was possible to distinguish patients with colorectal cancer (median 0.8) from the healthy group (median 3) and those with multiple myeloma (median 4.7).

An alternative approach in which allelic imbalance and DNA hypermethylation analyses have been combined, has been exploited by Lessard et al. who demonstrated a significantly improved sensitivity for the method to detect prostate cancer. This approach, using loss of heterozygosity combined with hypermethylation, has proved to be more sensitive than the currently used prostate specific antigen serum levels.

Another approach for the detection of prostate cancer was emplyed by Van der Vaart et al. using parallel tagged sequencing of circulating DNA on the GSFLX sequencer from 454 life sciences. A total of ~3600 unique sequences were analyzed and were seen to be distributed over the human genome with a slightly higher mutation rate being observed for DNA obtained from the cancer patients when compared to the control group. A further characterization of this array of sequences was performed by comparative analysis of chromosome distribution, repeat content and epigenetic characteristics of plasma DNA. Satellite repeats attributed to chromosome 12 were elevated in plasma of prostate cancer patients.

Although the average concentration of circulating DNA, measured as LINE-1 repetitive elements, in plasma was shown to be similar in healthy individuals and non-small cell lung cancer patients, Ponomaryova et al. also found that the concentration of cell-surface-bound circulating DNA was significantly low. This correlated with a poor disease prognosis. The ratio of the  $\beta$ -actin gene to LINE-1 was found to be elevated in the cell-surface-bound DNA of the non-small cell lung cancer patients compared to healthy individuals,. Hence, these results indicate a possible role for  $\beta$ -actin gene and LINE-1 fragments circulating in non-small cell lung cancer patients in both tumour detection and prognosis. Ding et al. also found that the quantification of methylated *RASSF1A* after operation provided a useful prognostic biomarker for predicting the recurrence in non-small cell lung cancer patients after curative-intent surgery.

Epigenetic effects in the shape of promotor methylation rates of three tumour suppressor genes from both plasma DNA and cell-surface-bound DNA from gastric cancer patients were considered by Elistratova et al. Methylated forms of p15, MGMT and hMLH1 genes were detected with high rates at stages II, III and IV of gastric cancer. However, no significant correlation was found between epigenetic and protein markers so indicating their independent development in gastric tumor pathogenesis.

A different approach has been taken by Roth et al. in studies on ovarian cancer patients. The concentrations of four circulating microRNAs (miRNA10b, miRNA34a, miRNA141 and miRNA155) were measured in the serum of 59 patients with ovarian cancer and 29 healthy individuals. The levels of total RNA, miRNA10b, miRNA34a, miRNA141, and miRNA155 in ovarian cancer patients,

were significantly higher than those from the healthy controls. A significant correlation was also recorded of increasing amounts of miRNA34a with lymph node metastases.

### Metastases

Amongst the DNAs circulating in cancer patients will be DNAs released from tumour cells either by necrosis or apoptosis or as newly synthesised virtosomes (Adams et al. 1997; Anker et al. 1994; Garcia-Olmo et al. 2010; Stroun et al. 1989). CNAPS DNAs can readily enter cells and in some cases be expressed in a way that modifies the biology of the recipients cell (Adams et al. 1997; Anker and Stroun 1972; Anker et al. 1980, 1994; Bulicheva et al. 2008; Ermakov et al. 2008; Garcia-Olmo et al. 2010; Ottolenghi and Hotchkiss 1960; Skvortsova et al. 2008). Thus, it is not only possible for cancer cells circulating in the blood to result in metastases, but also for the DNA released from tumour cells to do likewise.

One of the most common alterations of tumour related DNA found in CNAPS DNA from cancer patients is its hypermethylation. Thus, methylated fragments of the RAR2 gene from CNAPS have been shown to be taken up by HeLa and human umbilical vein endothelial cells twice as efficiently as unmethylated fragments. Since the methylated RAR 2 gene fragments are more prevalent than the unmethylated fragments in intracellular traffic, they would appear to pose a higher transformation potential (Skvortsova et al. 2008).

It has been shown that the SW 480 cell line, originating from a human colon carcinoma and containing a point mutation of the K-ras gene on both alleles, can be released in the form of the newly synthesised, virtosomal DNA/RNA-lipoprotein complex containing the mutated K-ras gene. Culturing NIH/3T3 cells in the presence of the non-purified SW 480 cell supernatant containing the virtosome complex resulted in the appearance of transformed foci. The presence of a mutated ras gene in the transfected foci of the 3T3 cells was confirmed by hybridization after PCR and by sequencing the PCR product (Anker et al. 1994). In a similar fashion, the virtosomes released from mouse tumour cell lines J774 cells (leukemia) and P497 cells (glial tumour) entered non-stimulated lymphocytes and resulted in their stimulation to synthesize DNA for cell division (Adams et al. 1997). Therefore, it comes as no surprise that Garcia-Olmo et al. (2010) have proposed the Genometastases concept in which the DNA released from tumour cells into the blood moves to other cell sites - possibly stem cells - which are transformed into secondary tumours (Garcia-Olmo et al. 1999). Experimental evidence comes from studies by Garcia-Olmo et al. (2010) in which cultures of NIH-3T3 cells were supplemented with samples of plasma from patients with either K-ras-mutated colorectal tumours or from healthy subjects. This was made by either direct addition of plasma to cultures in standard plates or avoiding plasma-cell contact by placing membranes with  $0.4 \,\mu$ m pores between the plasma and the cultured cells to act as a filter and so avoid the involvement of any free host cancer cells. Human gene transfer occurred in most

7

cultures of NIH-3T3 cells, as shown by the presence of human K-ras sequences, p53 sequences and  $\beta$ -globin encoding sequences. Furthermore, the NIH-3T3 cells were shown to be oncogenically transformed after being cultured with plasma from colon cancer patients by the development of carcinomas in NOD-SCID mice injected with the transformed NIH-3T3 cells. Cultures with an artificial membrane containing 0.4  $\mu$ m diameter pores placed between the NIH-3T3 cells and the plasma gave similar results showing that the transforming factor was smaller than 0.4  $\mu$ m (Garcia-Olmo et al. 2010). The presence of small vesicle-like structures was confirmed by Serrano-Heras et al. through the demonstration of an increased release of DNA-containing vesicles in the bloodstream of tumour bearing, compared to normal, rats. The DNA was shown to contain K-ras sequences and, hence, may be the source of the transforming DNA in the bloodstream. This is strong confirmation to the idea that circulating DNA released from tumour cells can be the direct cause of metastases (Garcia-Olmo et al. 1999).

### **Nucleic Acids in Foetal Medicine**

Pregnant women often opt for prenatal diagnosis to test for foetal chromosomal aneuploidies, the most common aneuploidies including trisomy 21, trisomy 18, trisomy 13 (Savva et al. 2010) and monosomy X in females (Ranke and Saenger 2001). This usually involves the invasive procedures of chorionic villus sampling and amniocentesis in order to obtain foetal genetic material for analyses, such procedures, at times, resulting in the loss of the foetus (Tabor et al. 1986).

The discovery that cell-free foetal DNA contributes a mean of 3–6% of the total maternal plasma DNA (Lo et al. 1998b). has permitted the development of some methods that have already been translated into clinical use e.g. the non-invasive determination of fetal rhesus D status (Lo et al. 1998a; Daniels et al. 2009) and the exclusion of sex-linked disorders (Costa et al. 2002). The early approaches focussed on the detection of foetal-specific RNA/DNA molecules for chromosome dosage determination involving RNA-single nucleotide polymorphism (SNP), epigenetic allelic ratios and epigenetic-genetic chromosome dosage. Tong et al. presented a highly sensitive polymorphism-independent approach using a very precise digital polymerase chain reaction platform together with a single molecule counting technology and a parallel sequencing platform for the direct detection of foetal chromosomal aneuploidies from maternal plasma.

An alternative analyses has been considered by Hultén et al. in which methylated DNA immunoprecipitation in combination with high resolution oligonucleotide microarray analysis has permitted the identification of chromosomal DNA methylation patterns using a high-throughput approach. The methylation patterns of chromosomes 13, 18, 21 and the sex chromosomes in female peripheral blood, CVS and placental DNA will form the basis of non/minimally-invasive prenatal analysis. Morozkin et al. have employed fluorescent *in situ* hybridization to examine extracellular DNA versus genomic or apoptotic DNA from culture medium and bound to the cell surface of human primary endotheliocytes, human primary fibroblasts

and HeLa cells. An over-representation was found for chromosome 9 fragments and the regions of the short arms of chromosomes 13, 14, 15, 21, 22 in DNA isolated from the culture medium of primary fibroblasts. These finding offer DNA targets for diagnostic purposes.

While the above approaches are important in identifying some chromosomal abnormalities, foetal sex and Rhesus factors in the first trimester, only a relatively small fraction of foetuses are affected with trisomy 21. There are many infants and children with a variety of developmental disorders that are not due to aneuploidy and who could benefit from a real-time genomic approach to better understand foetal development and to identify key genes involved in the pathogenesis of disorders such as a means of targeting for therapy. Working with neonatal mRNA rather than DNA, Maron and Bianchi in a "fluid agnostic" approach, have concentrated on mRNA fractions from maternal and neonatal whole blood, amniotic fluid, and neonatal saliva as potential sources of genomic information that could assist an understanding of foetal development, pathology, and diagnosis. Working with mRNA will give a better chance to study differentially regulated genes and so expand the range of developmental and pathological targets.

### **Other Clinical Exploitation of CNAPS**

The level of circulating DNA has been shown to increase in patients presenting with injury, the concentration relating to the severity of the injury (Lan et al. 2003). DNA measurement on admission could be used to predict the outcome in terms of organ failure, acute lung injury, acute respiratory syndrome and death. Similarly, ß-globulin DNA concentration was found to be higher in patients presenting with stroke and could be used as a predictor of death (Rainer and Lam 2006) as were nucleosomes (Geiger et al. 2007). A new duplex real-time PCR assay with internal control developed by Chen et al. was used by them to study circulating plasma DNA levels in trauma patients from the Wenchuan, China earthquake in 2009. During the early stage of injury, the median plasma DNA level of patients was more than five times that of the healthy controls and a statistically significant difference of plasma DNA concentration between patients with and without organ injury was determined.

Cerebrovascular accidents are also characterized by the increase in low molecular weight DNA concentration in the course of 3 days after acuity with a maximum after 3 hours in the case of hemorrhage and after 24 hours in the case of ischemia. Recent analysis by Vasilyeva et al. of such low molecular weight DNA from the spinal fluid from patients with severe cerebral vascular circulatory problems showed a sharp increase within 3 h from the start of the attack, similar to that seen with the DNA fraction from blood. However, since the spinal fluid contains no blood cells during the first 24 h after the attack, the DNA is likely to have the brain lesion as its source.

Horinek et al. have found that plasma DNA levels increase sharply in patients undergoing dialysis and although the levels drop subsequently, they do not return to the control values. The increased DNA levels could be due to apoptosis as shown by over-expression of the pro-apoptotic genes *BAX* and CASP8.

### The Biology of CNAPS

As has already been mentioned, there are a variety of sources of CNAPS and there are many questions relating to the biology of these nucleic acid fractions to be answered. In some instances CNAPS have been demonstrated to be actively released from cells, readily taken up by other cell populations and biologically active (reviewed Gahan and Stroun 2010). The mechanisms controlling the production and release of both the DNA and RNA fractions, the mechanisms of release and uptake and the way in which they can modify the recipient cell's biology are still to be clarified.

Methylated DNA enters cells more easily than non-methylated DNA as shown by the uptake of methylated fragments of RAR2 gene into HeLa and human umbilical vein endothelial cells being twice as efficient as that of unmethylated fragments. A common alteration of tumour related DNA found in CNAPS concerns the hypermethylation of DNA from cancer patients. Since the methylated RAR 2 gene fragments are more prevalent than the unmethylated fragments in intracellular traffic, they would appear to pose a higher transformation potential (Skvortsova et al. 2008). Skortsova et al. have gone on to show that when human CNAPS is injected into mice methylated DNA was degraded less quickly than the unmethylated form. In addition, a quantitative study of RARbeta2 gene methylation in cell-free DNA and genomic DNA of primary and transformed cells showed an over-representation of methylated DNA sequences in the circulating DNA of primary cells.

From the results of Korabecna et al., it would appear that plasma DNAase II makes only a minor contribution to the degradation of circulating DNA.

A number of studies have reinforced the concept that the circulating nucleic acids can enter host cells and modify the biology of those cells. Thus, Malinovskaya et al. have shown that CpG-enriched rDNA accumulating in human cfDNA significantly stimulates gene transcription in mesenchymal stem cells by activating TLR9 and MyD88-dependent signaling pathways and inhibiting differentiation of mesenchymal stem cells into adipocytes. Inhibition of Poly(I:C)-activated IL-6 and IL-8 Production in Human Primary Endothelial Cells and Fibroblasts was also demonstrated by Cherepanova et al.

The accumulating CpG-rich ribosomal repeat was demonstrated by Efremova et al. to influence brain cell function in pathology and injury being accompanied by intensive DNA liberation from cells as the result of apoptosis or necrosis. pBRTRRR significantly up-regulated iNOS gene expression, being more effective in low concentrations as was the case for iNOS gene expression. Similarly, Alexseeva et al. also showed that cell free DNA could influence the elaboration of NO and ROS depending upon the sample concentration and the content of CG-DNA marker with cell free DNA isolated from blood from patients with cardiovascular diseases influencing ROS synthesis more efficiently than did cell free DNA from healthy donors.

Ermakov et al. have previously shown that low-dose ionizing radiation induced in human  $G_0$ -lymphocytes the development of an adaptive response that was accompanied by transposition of homologous-chromosomes loci within the cell nucleus

and activation of the chromosomal nucleolar-forming regions. Such reactions were transmitted to unirradiated lymphocytes via the bystander-effect mechanism (Ermakov et al. 2008). Similar results have been obtained with monolayer mesenchymal stem cell cultures after the development of the radiation induced bystander effect.

Few studies have been made on the effects of circulating RNA to enter cells though RNA has previously been shown to be capable of transforming cells (Skog et al. 2008). Semenov et al have developed a number of analogues of both singleand double-stranded RNAs that have readily entered cells and produced a variety of biological effects in the recipient cells.

### **New Technology**

Although the use of massively parallel sequencing (Rogers and Ventner 2005) has facilitated the development of the analyses of CNAPS such as for diagnosis of trisomy 21 (Chiu et al. 2010), the inclusion of CNAPS as a major player in predictive and preventive medicine will depend upon the reliability of the way that the withdrawn blood is handled prior to nucleic acid extraction and the mechanism employed for the extraction of the nucleic acids as well as the development of rigorous and repeatable techniques linking a particular nucleic acid fragment to a specific clinical disorder.

A way forward designed by Krishnan and Heller allows the nucleic acids to be directly removed from whole blood, even immediately after withdrawal. Using a microarray dielectrophoretic system, high molecular weight DNA can be both detected and rapidly isolated directly from whole blood. Levels of < 260 ng per ml DNA are detectable. The method can also be applied for the isolation of nanoparticles at <  $9.5 \times 10^9$  particles per ml.

Given the variability between available commercial methods for the extraction of nucleic acids from plasma and serum, Fleischhacker et al. have made a comparison of three kits in an effort to establish which kit yields the most DNA isolated versus the immunological quantification of circulating nucleosomes using the Cell Death Detection ELISA plus. The study was performed simultaneously in two separate laboratories. Comparable results were obtained with large differences being recorded between the different procedures and with the MagNA-Pure isolation system giving the highest DNA yield.

The isolation of nucleic acids from plasma, serum and urine has been improved by the development of a new QIAamp<sup>®</sup> Circulating Nucleic Acid Kit by Horlitz et al. This large volume kit yields 7–9 times as much as that derived with the the QIAamp Blood Mini as well as offering improved recovery of short DNA fragments. It would appear that the QIAamp Circulating Nucleic Acid Kit can serve as a sample preparation solution for processing up to 5 ml cell-free body fluid and can extract and concentrate circulating nucleic acids, including microRNA, and viral nucleic acids up to 250-fold. One of the newer developments in CNAPS has been the introduction of microRNAs as early markers for diagnosis and prognosis. One area in which this has been applied concerns the study of Alzheimer disease by van Harten et al. Using the Megaplex protocol with Taqman Array MicroRNA cards on small RNA isolated with the MirVana Paris kit it was possible to isolate all 667 currently known microRNAs from the spinal fluid of Alzheimer patients.

### Conclusions

There is still a long way to go before CNAPS will become fully integrated into predictive and preventive medicine. However, a strong beginning has been established in both the technology available and the identification of the relevant nucleic acid fragments linked to specific clinical disorders. The trialling of foetal diagnostic methods in national health programmes in some countries is good evidence for this. Nevertheless, it is clear that quality assured methodologies will be needed for three important areas, namely, whole blood handling prior to nucleic acid extraction, the mechanisms employed for the nucleic acid extraction and the development of rigorous and repeatable techniques linking a particular nucleic acid fragment to a specific clinical disorder for either prediction or prognosis.

### References

- Adams DH, Diaz N, Gahan PB (1997) In vitro stimulation by tumour cell mediaof [3H]thymidine incorporation by mouse spleen lymphocytes. Cell Biochem Funct 15:1191–1126
- Anker P, Stroun M (1972) Bacterial ribonucleic acid in the frog brain after a bacterial peritoneal infection. Science 178:621–623
- Anker P, Jachertz D, Stroun M, Brogger R, Lederrey C, Henri J, Maurice P (1980) The role of extracellular DNA in the transfer of information from T to B human lymphocytes in the course of an immune response. J Immunogenet 6:475–481
- Anker P, Lyautey J, Lefort F, Lederrey C, Stroun M (1994) Transformation of NIH/3T3 cells and SW 480 cells displaying K-ras mutation. CR Acad Sci III 10:869–874
- Bulicheva N, Fidelina O, Krtumova MN, Neverova M, Bogush A, Bogush M, Roginko O, Veiko N (2008) Effect of cell-free DNA of patients with cardiomyopathy and rDNA on the frequency of contraction of electrically paced neonatal rat ventricular myocytes in culture. Ann NY Acad Sci 1137:273–277
- Chiu RWK, Sun H, Akolekar R, et al (2010) Maternal plasma DNA analysis with massively parallel sequencing by ligation for noninvasive prenatal diagnosis of trisomy 21. Clin Chem 56:459–463
- Costa JM, Benachi A, Gautier E (2002) New strategy for prenatal diagnosis of X-linked disorders. N Engl J Med 346:1502
- Daniels G, Finning K, Martin P, et al (2009) Noninvasive prenatal diagnosis of fetal blood group phenotypes: current practice and future prospects. Prenat Diagn 29:101–107
- Ermakov AV, Kostyuk SV, Konkova MS, Egolina NA, Malinovskaya EM, Natalya N, Veiko NN (2008) Extracellular DNA fragments: factors of stress signalling between X-irradiated and unirradiated human lymphocytes. Proc NY Acad Sci 1137:41–46