

Childhood Acute Lymphoblastic Leukemia

Ajay Vora
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Ajay Vora
Department of Paediatric Haematology
Sheffield Children's Hospital NHS Trust
University of Sheffield
Sheffield
South Yorkshire
UK

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Contributors

Persis Amrolia Department of Bone Marrow Transplant, Great Ormond St Children's Hospital, London, UK

Molecular and Cellular Immunology Section, UCL Institute of Child Health, London, UK

Deepa Bhojwani Children's Center for Cancer and Blood Diseases, Children's Hospital Los Angeles, Los Angeles, USA

Keck School of Medicine, University of Southern California, Los Angeles, CA, USA

Denise Bonney Paediatric Haematology, Royal Manchester Children's Hospital, Central Manchester University Hospitals NHS Trust, Manchester, UK

Children's Cancer Group, Paterson Building, Institute of Cancer, University of Manchester, Manchester, UK

M. Brüggemann Department of Hematology, University Hospital Schleswig Holstein, Campus Kiel (UNIKIEL), Kiel, DE, Germany

Christiane Chen-Santel Department of Pediatric Oncology/Hematology/BMT, Charité University Medicine Berlin, Berlin, Germany

Rachel Dommett Bristol Royal Hospital for Children, Bristol, UK

Sara Ghorashian Molecular and Cellular Immunology Section, UCL Great Ormond Street Institute of Child Health, London, UK

Christine Harrison Leukaemia Research Cytogenetics Group, Newcastle University, Newcastle upon Tyne, UK

Stephen P. Hunger Department of Pediatrics and the Center for Childhood Cancer Research, Children's Hospital of Philadelphia and the Perelman School of Medicine at the University of Pennsylvania, Philadelphia, PA, USA

Ilaria Iacobucci St Jude's Hospital, Memphis, USA

Julie Irving Northern Institute for Cancer Research, Newcastle University, Newcastle upon Tyne, UK

Su Han Lum Paediatric Haematology, Royal Manchester Children's Hospital, Central Manchester University Hospitals NHS Trust, Manchester, UK

Anthony Moorman Leukaemia Research Cytogenetics Group, Newcastle University, Newcastle upon Tyne, UK

John Moppett Bristol Royal Hospital for Children, Bristol, UK

Charles Mullighan St Jude's Hospital, Memphis, USA

A. Orfao Department of Medicine, Cancer Research Center (IBMCC-CSIC-USAL) and Cytometry Service (NUCLEUS), University of Salamanca (USAL) and IBSAL, Salamanca, ES, Spain

Etan Orgel Children's Center for Cancer and Blood Diseases, Children's Hospital Los Angeles, Los Angeles, USA

Keck School of Medicine, University of Southern California, Los Angeles, CA, USA

Vaskar Saha Paediatric Haematology, Royal Manchester Children's Hospital, Central Manchester University Hospitals NHS Trust, Manchester, UK

Children's Cancer Group, Paterson Building, Institute of Cancer, University of Manchester, Manchester, UK

Tata Translational Cancer Research Centre, Tata Medical Center, Kolkata, India

Kjeld Schmiegelow, MD, DrMedSci Department of Pediatrics and Adolescent Medicine, University Hospital Rigshospitalet, Copenhagen, Denmark

Institute of Clinical Medicine, University of Copenhagen, Copenhagen, Denmark

Claire Schwab Leukaemia Research Cytogenetics Group, Newcastle University, Newcastle upon Tyne, UK

Ulrik Kristoffer Stoltze Department of Pediatrics and Adolescent Medicine, University Hospital Rigshospitalet, Copenhagen, Denmark

Sarah K. Tasian Department of Pediatrics and the Center for Childhood Cancer Research, Children's Hospital of Philadelphia and the Perelman School of Medicine at the University of Pennsylvania, Philadelphia, PA, USA

Morten Tulstrup Department of Pediatrics and Adolescent Medicine, University Hospital Rigshospitalet, Copenhagen, Denmark

Jun J. Yang, PhD Department of Pharmaceutical Sciences, Hematological Malignancies Program, St. Jude Children's Research Hospital, Memphis, TN, USA

Inge van der Sluis, MD, PhD Department of Pediatric Hematology-Oncology, Erasmus Medical Center – Sophia Children's Hospital, Rotterdam, The Netherlands

V.H.J. van der Velden Department of Immunology, Erasmus MC, University Medical Center Rotterdam, Rotterdam, Netherlands

J.J.M. van Dongen, MD, PhD Department of Immunohematology and Blood Transfusion, Room E-03-50, Leiden University Medical Center (LUMC), ZA, Leiden, The Netherlands

Department of Immunology, Erasmus MC, University Medical Center Rotterdam, Rotterdam, The Netherlands

Ajay Vora University College London, London, UK

Allen Yeoh Viva-University Children's Medical Institute, Singapore, Singapore
Khoo Teck Puat-National University Children's Medical Centre, National University Hospital, Singapore, Singapore

Yong Loo Lin School of Medicine, National University of Singapore, National University Health System, Singapore, Singapore

Chapter 1

Epidemiology and Etiology of Childhood ALL

Morten Tulstrup, Ulrik Kristoffer Stoltze, Kjeld Schmiegelow,
and Jun J. Yang

1.1 Introduction

Despite extensive research, the etiology of childhood acute lymphoblastic leukemia (ALL) remains largely unknown. There is growing evidence that this cancer may arise from in utero chromosomal abnormalities that can lead to clonal expansion of pre-leukemic precursor cells. The risk factors for ALL in children are multiple, most notably common germline polymorphisms and rare genetic syndromes that directly influence hematopoiesis and cell cycling, as well as possibly infection-related aberrant DNA editing.

1.2 General Epidemiology

The incidence of ALL varies by age, ethnicity, geographic region, and also differs by immunologic and molecular subtypes. In both the United States and the Nordic countries, the overall incidence rate is 3.9 per 100,000/year before the age of

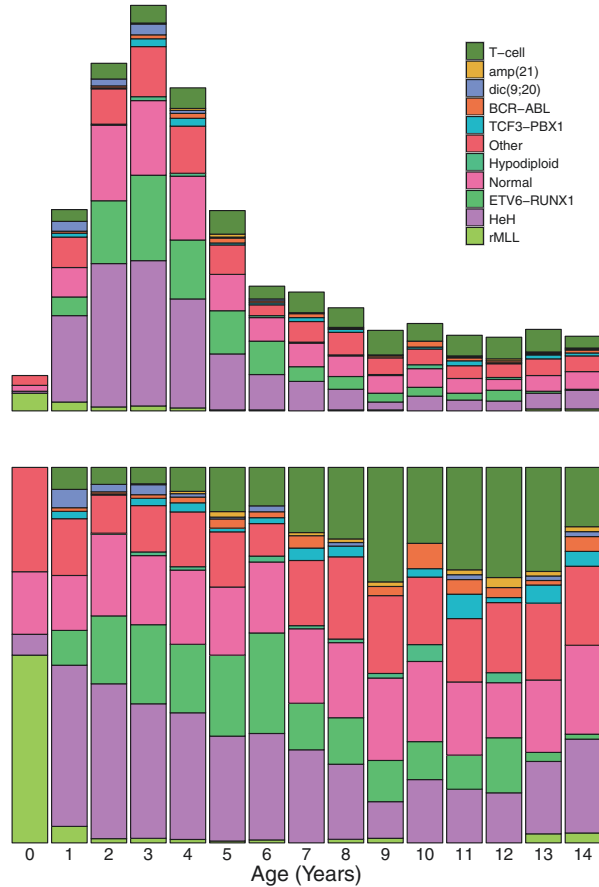
M. Tulstrup • U.K. Stoltze
Department of Pediatrics and Adolescent Medicine, University Hospital Rigshospitalet,
Copenhagen, Denmark

K. Schmiegelow
Institute of Clinical Medicine, University of Copenhagen, Copenhagen, Denmark

Department of Pediatrics and Adolescent Medicine, University Hospital Rigshospitalet,
Copenhagen, Denmark

J.J. Yang (✉)
Hematological Malignancies Program, St. Jude Children's Research Hospital,
Memphis, TN, USA
e-mail: jun.yang@stjude.org

Fig. 1.1 Age distribution of childhood ALL cases by immunologic and molecular subtypes. Numbers represented are all children diagnosed with ALL in Denmark, Sweden, Norway, Finland, and Iceland between 1992–2007. Upper panel: bar heights represent the number of cases in each age group relative to the total number of cases between 0 and 14 years. Lower panel: relative distribution of subtypes within each age group. The testing for t(12;21) [*ETV6-RUNX1*] by *fluorescence in situ hybridization* was gradually introduced during this period, and accordingly some amp(21) patients have been missed. *Ph+*, Philadelphia chromosome-positive; *HeH*, high-hyperdiploid



15 years [1, 2]. The incidence is higher in Hispanic Americans (4.1 per 100,000/year), and is lower in African American children (2.1 per 100,000/year) [3, 4]. In general, low-income countries have lower incidences of ALL than high-income countries, with a few exceptions such as Costa Rica (4.6 per 100,000/year), however these differences may be the result of incomplete registration [4–7]. The incidence of ALL shows a characteristic peak between 2 and 5 years after birth [2, 7], but age-related ALL risk differs substantially by cytogenetic subtype (Fig. 1.1). ALL in infants (<1 year) is in most cases characterized by *MLL* gene rearrangements (*rMLL*), which are rare in older children [8–10]. Between 2 and 5-year olds, ALL is dominated by high-hyperdiploid (HeH, modal chromosome number >50) and t(12;21)[*ETV6-RUNX1*] karyotypes, while T-cell ALL has a less pronounced peak around 4–9 years [2, 10, 11]. In low-income countries, the 2–5 year age peak is much less obvious, with a higher proportion of T-ALL [5–7, 12–15]. Interestingly, some studies noted incremental increase in ALL incidences specifically in this age range as a function of economic growth and improving living conditions [16–19].

Taken together, these observations (1) suggest that different ALL subtypes may have distinctive etiological mechanisms and (2) point toward possible effects of economic development-related environmental factors on ALL risk.

1.3 Natural History

Monozygotic twins have a 10–20% concordance rate for ALL, and concordant cases have been shown to harbor identical and clonotypic molecular signatures (e.g. *ETV6-RUNX1* fusion sequence, or T-cell receptor (*TCR*), immunoglobulin (*IGH*) gene and *MLL* rearrangements) possibly because a leukemic or preleukemic clone arose prenatally in one twin and spread to the other through placental vascular anastomoses [20–25]. Further evidence for a prenatal initiation is provided by studies backtracking disease-specific molecular markers in both twin and singleton leukemias in dried blood spot samples (DBSS) from birth (Table 1.1).

For infant *rMLL* ALL, rearrangement has been identified in DBSS in the vast majority of cases, suggesting that this disease almost always arises prenatally. Older patients with *rMLL* are usually DBSS negative, but the translocation has been successfully backtracked in one case diagnosed at 6 years. Similar findings have been reported for *ETV6-RUNX1* ALL. This translocation causes a fusion of the *ETV6* and *RUNX1* genes, and the resulting chimeric protein has been shown to promote cell survival in mice and human cells [44–46]. Three studies on concordant (monozygotic) twins revealed identical *ETV6-RUNX1* fusion sequences in both twins and clonal expansion of fusion-positive precursors at a minimum level of 10^{-4} preleukemic cells at birth. Prenatally initiated *ETV6-RUNX1*⁺ cases have had a latency of up to 14 years before overt leukemia occurred [47]. Furthermore, two ALL-discordant twin pairs have been described in which the healthy twin also harbored an *ETV6-RUNX1*⁺ clone at birth [30] or at age 3 [44], suggesting that the translocation in itself is insufficient for leukemia development. Leukemic *ETV6-RUNX1*⁺ cells harbor a variable number of additional mutations; often a deletion of the wildtype *ETV6* allele or other genes involved in B-lymphocyte development and differentiation [48–51]. Molecular studies of concordant monozygotic twins showed that these mutations are unique to each twin and thus occur as secondary postnatal events [51, 52]. An often cited study found that 1% of all healthy newborns harbored *ETV6-RUNX1* at birth (i.e. 100-fold of the incidence of *ETV6-RUNX1* ALL) [53], but subsequent validation studies have raised questions about the reliability of the initial finding [54–58]. Thus, while healthy children in general may harbor *ETV6-RUNX1*⁺ cells without developing ALL, the exact prevalence of such an event has yet to be determined.

HeH ALL cases frequently have detectable clonotypic *IGH* rearrangements in neonatal blood spots (17 of 29) and the hyperdiploidy in itself can also arise prenatally [38]. Importantly, a hyperdiploid clone has been found in a healthy twin sibling of a child with HeH ALL [59]. Recently, a whole-genome sequencing approach has further supported the notion that gross chromosomal gains occur early

Table 1.1 Backtracking studies and their findings

Paper	Subtype	Marker	N	N pos.	Comments
Gale <i>PNAS</i> 1997 [26]	rMLL	MLL-AF4	3	3	
Wiemels <i>Lancet</i> 1999 [27]	ETV6-RUNX1	ETV6-RUNX1	11	8	One twin pair, both positive
Fasching <i>Blood</i> 2000 [28]		IGH	1	1	
	rMLL	IGH + MLL-AF4/AF4-MLL	2	2	
	T-ALL	TCR	2	2	
Yagi <i>Blood</i> 2000 [29]	B-ALL	IGH + TCRD	4	1	IGH and TCRD positive
	HeH	IGH + TCRD	1	1	IGH and TCRD positive
	rMLL	IGH + TCRD	2	2	Both IGH positive, only one TCRD positive
Maia <i>Blood</i> 2001 [30]	ETV6-RUNX1	ETV6-RUNX1	3	3	Triples, one healthy
Panzer-Grümayer <i>Blood</i> 2002 [31]	HeH*	IGH	1	1	
Taub <i>Blood</i> 2002 [32]	B-ALL	IGH	10	5	1 Down syndrome, TCF3-PBX1, 1 rMLL
	HeH	IGH	6	6	
	ETV6-RUNX1	IGH	1	1	
Hjalgrim <i>Br J Cancer</i> 2002 [33]	ETV6-RUNX1	ETV6-RUNX1	9	3	
Wiemels <i>PNAS</i> 2002 [34]	TCF3-PBX1	IGH and TCF3-PBX1	15	0	
McHale <i>Genes Chrom Cancer</i> 2003 [35]	ETV6-RUNX1	ETV6-RUNX1	14	7	
Teuffel <i>Leukemia</i> 2004 [36]	ETV6-RUNX1	ETV6-RUNX1 + IGH	2	2	Twins. Shared ETV6-RUNX1 sequence, different IGH
Maia <i>Genes Chrom Cancer</i> 2004 [37]	rMLL	MLL-AF4	4	1	Specifically chose children with higher age a diagnosis
	ETV6-RUNX1	ETV6-RUNX1	7	3	
Maia <i>Genes Chrom Cancer</i> 2004 [2] [38]	HeH	IGH	11	1	DBSS
	HeH	Trisomy 15 + 17	1	1	Cord blood – this patient was DBSS-IGH negative
Fischer <i>Blood</i> 2007 [39]	T-ALL	TCR	16	1	Other markers: <i>TALI</i> , <i>Notch1</i> , and <i>TCRD-LMO</i> .
Gruhn <i>Leukemia</i> 2008 [40]	B-ALL	IGH	17	11	
	HeH	IGH	6	5	
	ETV6-RUNX1	IGH	6	3	
	BCR-ABL	IGH	2	0	
	rMLL	IGH	1	0	

Table 1.1 (continued)

Paper	Subtype	Marker	N	N pos.	Comments
Wiemels <i>Blood Cell Mol Dis</i> 2010 [41]	HeH	<i>RAS mutations</i>	14	0	
	HeH	<i>IGH</i>	4	3	All four were also tested for <i>RAS</i> mutations
Eguchi-Ishimae <i>Blood</i> 2011 [42]	T-ALL	<i>NOTCH1</i>	3	1	
Mansur <i>Br J Haematol</i> 2015 [43]	T-ALL	<i>PTEN</i>	4	3	Infant T-ALL

High-hyperdiploidy (HeH): >50 chromosomes

^aChromosome number or DNA index not specified

in life as the sentinel event and that additional postnatal events are also necessary for leukemia development [60].

Clonal development of T-ALL is much less understood and leukemic genomic aberration is rarely detected at birth in children with this ALL subtype, suggesting an entirely different etiology compared to B-ALL.

In summary, *rMLL*, *ETV6-RUNX1*⁺, and HeH ALL show the most convincing evidence of prenatal initiation, while other subtypes such as T-ALL, *BCR-ABL* and *TCF3-PBX1* are less frequently or never prenatally initiated.

1.4 Environmental Risk Factors

1.4.1 Infectious Disease and Immune Stimulation

It has long been hypothesized that infectious disease plays a role in the development of ALL. In 1988, Leo Kinlen postulated that mixing of previously isolated populations could cause epidemics of an unidentified pathogen to which leukemia was a rare response [61]. This hypothesis was based on observed spatial and temporal clustering of ALL cases, which occurs at an exceedingly rare frequency [62]. The same year, Mel Greaves suggested that children with little early-life immune stimulation can develop leukemia as an aberrant response to a delayed exposure to common infections [63]. This ‘delayed-infection hypothesis’, which in many ways is similar to the ‘hygiene hypothesis’ concerning allergies and atopic disease, is particularly relevant to ALL risk in the 2–5 year age peak [64–67]. In these cases, the prenatal formation of a preleukemic clone may constitute a commonly occurring ‘first hit’, and an aberrant immune response due to delayed immune maturation and subsequent uncontrolled proliferative stress on exposure to a common childhood infection occur subsequently will in rare cases cause a second hit and initiate malignant transformation [64, 65].

A substantial body of evidence has been gathered in support of an association between infections and ALL risk. Since the actual number of childhood infections is difficult to measure, proxy measures such as daycare-attendance (children in

daycare are more exposed to common infections early in life) are typically examined [68]. A meta-analysis from 2010 by Urayama et al. included 14 studies and a total of 6108 cases and found a significantly reduced risk of ALL among children in daycare (OR = 0.76; 95% CI: 0.67–0.87) [69]. A recent study confirmed this finding and furthermore indicated that the protective effect of daycare is even stronger with earlier start of attendance [70]. Another measure of early immune stimulation is breastfeeding, for which two meta-analyses consistently found an association with a reduced risk of ALL; subsequently a large case-control study with 7,399 ALL cases and 11,181 controls also reported an OR of 0.86 (95% CI: 0.79–0.94) for children breastfed for 6 months or more [70–72]. Other proxies for immune stimulation include birth order and vaccinations, but epidemiological findings on these exposures are inconsistent [70, 73–79]. More direct attempts at measuring actual number of infections during early childhood have included patient registries [80–84], questionnaires [70, 85, 86], and interviews [87–89]. Generally, studies using parentally reported measures found an inverse or no association between infections and ALL risk, while the patient registry-based methods, which have the strength of eliminating recall bias, found either positive or null associations. Interpreting data from these studies is difficult for a number of reasons, most notably the heterogeneity of exposure definitions and the timing of infections in relation to ALL diagnosis. According to the delayed infection hypothesis, children prone to ALL-development should have fewer infections in early life and subsequently start developing aberrant responses to common infections, most likely resulting in symptomatic infectious disease. However, in the months leading up to ALL diagnosis the disease itself also becomes a risk factor for infections, and thus the expected direction of causality between infection and leukemia becomes difficult to identify in such epidemiologic studies [84].

Recent molecular studies have shed new lights on the role of infection in ALL development. Whole-genome sequencing of *ETV6-RUNX1* ALL cells revealed that most of the somatic deletions commonly seen in this subtype are mediated by the RAG enzymes, the main function of which is V(D)J recombination in normal pre-B cells [90], potentially as a result of infection-related hyperactivation of RAG. Subsequently, Swaminathan et al. showed that premature activation of the AID enzyme (which normally mediates somatic hypermutation and class-switch recombination in mature B-cells) resulting in inappropriate, synchronous activation of AID and RAG increases genetic instability in pre-B cells, especially those with the *ETV6-RUNX1* fusion [91]. The authors furthermore showed that while infectious stimuli (mimicked by lipopolysaccharide) could induce leukemic transformation of *ETV6-RUNX1*⁺ cells, this development was delayed or prevented in mice without functional *AID* or *RAG*, respectively. Another example highlighting a molecular mechanism involved in infection-mediated ALL development is *PAX5*, a gene commonly mutated in B-ALL. A recent study showed that *PAX5* heterozygous mice were prone to develop ALL, but only if they were exposed to common infections [92]. It is important to note that these molecular studies show that infections are likely involved in ALL development, but provide no direct evidence of how early vs. late infection alters the risk of ALL during childhood.

Table 1.2 Non-infectious environmental risk factors

Risk factor	Certainty ^a	Comments
<i>In utero</i> diagnostic radiation [94, 95]	Inconclusive	
Background ionizing radiation [96–98]	Inconclusive	Uncertain association, but if true may account for 8–30% of all cases
Radon [99–101]	Inconclusive	
Extremely low-frequency electromagnetic fields [102–104]	Inconclusive	If true, this could account for 2–3% of cases
Radio frequency electromagnetic fields [105, 106]	Unlikely	
Birth weight [93]	Confirmed	OR = 1.26 (95% CI: 1.17–1.37) for children ≥ 4000 g
Maternal age [107, 108]	Inconclusive	
Alcohol [108–111]	Unlikely	
Maternal smoking [108, 109]	Inconclusive	OR = 1.10 (95% CI: 1.02–1.19)
Paternal smoking [112]	Inconclusive	OR = 1.11 (95% CI: 1.05–1.18)
Prenatal folic acid [113–115]	Inconclusive	If anything, folate intake during pregnancy is protective
Postnatal vitamin K [116, 117]	Unlikely	
Pesticides [118–120]	Inconclusive	
Dietary topoisomerase II-inhibitors [121, 122]	Unlikely	

^a“Confirmed” indicates factors with consistent association in meta-analyses, “inconclusive” denotes factors with some evidence of association but also inconsistent results from different studies, whereas “unlikely” is for those with no reliable evidence of association

1.4.2 Other Risk Factors

Despite a large number of epidemiological studies and meta-analyses, most findings regarding proposed environmental risk factors remain inconclusive. The only confirmed association is high birth weight, although the underlying mechanism is unknown [93]. Other factors such as ionizing radiation, electromagnetic fields and maternal smoking during pregnancy remain uncertain (Table 1.2). A common limitation is that the vast majority of these studies address ALL as a single disease entity and thus may have missed associations with specific ALL subtypes.

1.5 Heritability of ALL

Studies addressing the risk of leukemia among offspring of childhood leukemia survivors have been hampered by small sample sizes [123–126]. More reliable estimates of ALL heritability come from studies on risk in siblings of affected children. These studies have two important limitations: first, because of preleukemic cells’ ability to spread *in utero*, twins with leukemia need to be excluded

before estimating disease heritability, and secondly it is difficult to distinguish genetic effects from shared environmental risk factors between siblings. A recent Nordic population-based study reported a standardized incidence ratio (SIR) of 3.2 for ALL risk among siblings [127]. Furthermore, one study investigating 54 sibships with two or more cases of ALL found an unexpectedly high subtype concordance, pointing to a genetic basis of ALL etiology [128]. On the basis of genome-wide SNP data, it was estimated that inherited genetic polymorphisms account for at least 24% (95% CI: 6–42%) of variation in ALL risk [129]. In conclusion, these reports provide evidence for a genetic component in disease susceptibility, although reliable quantitative estimates of genetic contribution to ALL risk are not available.

1.6 High-Penetrance Genetic Predisposition

Out of more than 125 known cancer predisposition genes (CPGs), only 27 genes (associated with 9 rare syndromes and two non-heritable congenital disorders) are convincingly linked to childhood ALL (Table 1.3 and Fig. 1.2) [130, 131].

In a 2015 a registry study of 4939 childhood ALL cases, only 29 subjects were diagnosed with non-Down syndrome (DS) predisposition syndromes (0.6%) [161]. However, a recent comprehensive study of whole genome or whole exome sequencing in 588 non-DS childhood leukemia cases found germline mutations in known CPGs in 26 cases (4.4%) [161, 162]. This suggests that high-penetrance Mendelian genetics, discussed in detail below, may play a larger role in ALL etiology than previously appreciated.

1.6.1 Syndromes Where ALL Is a Dominant Cancer Phenotype

DS is one of the most common congenital abnormalities (1 in 691 live births) and also the most recognizable ALL-predisposition syndrome [140, 163]. ALL and AML risk is significantly increased, with SIR before 30 years of 24.4 and 20.3, respectively. Interestingly, individuals with DS have significantly lower incidence of solid cancers than the background population [142, 164]. DS-associated ALL is more likely to have somatic rearrangements involving the CRLF2 gene and almost always has B-cell immunophenotype. DS patients represent the only known group where ALL is the most common malignancy at any age. Taken together DS-ALL constitutes 2–3% of ALL [131, 165].

While the driver of leukemogenesis remains uncertain for DS it is likely that chromosome 21 is involved, as an acquired extra copy of chromosome 21 is also seen in hyperdiploid ALL and the intrachromosomal amplification of chromosome 21 seen in the iAMP21-ALL subtype [139].

Table 1.3 ALL syndromes

Syndrome	Heritability and involved genes	Association study/studies (ages at ALL diagnosis)	Approx. ALL relative risk	Common clinical features	Assoc. ALL Subtype	Commonly associated cancers	Estimated birth incidence ^b
Ataxia-telangiectasia (A-T)	AR, <i>ATM</i>	14 leukemias (all ages, presumably ALL) in 263 A-T patients [132]	OR = ~71 [132, 133]	Cerebellar ataxia, oculom. apraxia, telangiectasia	T-ALL	NHL, BC, OV, GC, STS	1 in 40,000–100,000 [134]
Bloom syndrome (BSyn)	AR, <i>BLM</i>	12 ALLs (mean age 19.4 (range: 5–40)) in 272 BSyn patients [135]	OR = ~92 [135] ^a	Short stature, skin lesions, infections, male infertility		Epithelial, AML, NHL	Unknown, 272 cases documented [135]
Constitutional Mismatch Repair Deficiency (CMMRD)	AR, <i>PMS2</i> (58%) <i>MLH1</i> / <i>MSH2</i> (22%) <i>MSH6</i> (20%) [136]	9 ALLs (median age 6 (range: 2–21)) in 146 CMMRD patients [136]	OR = ~131 [136] ^a	CALM, hyperpigmentation, NF1 symptoms,		Gliomas, NHL, GI, PNET, MB	1 in 775,000 [137]
Constitutional Robertsonian Translocation rob(15;21)(q10;q10) ^c	rob(15;21) ^c	None	OR (iAMP21-ALL only) = 2700 [138]	None	iAMP21-ALL	None	1 in 100,000–200,000 [139]

(continued)

Table 1.3 (continued)

Syndrome	Heritability and involved genes	Association study/studies (ages at ALL diagnosis)	Approx. ALL relative risk	Common clinical features	Assoc. ALL Subtype	Commonly associated cancers	Estimated birth incidence ^b
Down Syndrome (DS)	Trisomy 21 (95%) [140]	20 ALLs (ages 0–29) in 2814 DS patients [142]	SIR (95% CI) = 24.4 (14.9–37.6) [142]	Extensive; flat facial profile, CHD, hearing/vision loss,	HeH	AML, TC	1 in 691 [140]
	Rob. or other (4%) mosaic DS (1%) [141]						
ETV6 deficiency syndrome	AD, <i>ETV6</i>	16 ALLs in >35 <i>ETV6</i> mutation carriers [143–146]	Unknown, likely very high [144]	Thrombocytopenia	B-ALL	Likely ST	Unknown, 9 families documented [143–146]
Fanconi Anemia (FA)	AR, <i>FANCA</i> (60–70%) <i>FANCC</i> (14%) <i>FANCG</i> (10%) <i>BRCA2</i> (3%)	7 childhood ALLs in 1300 FA patients [148]	OR = ~11 [148] ^a	Short stature, CALM. None in 25–40% [149]		AML, MDS, GU, head/neck ST	1 in 100,000–360,000 [149, 150]
	+13 more (3–13%) [147]						
Li-Fraumeni Syndrome (LFS)	AD, <i>TP53</i>	12 leukemias (mean age 11 (range: 2–17), presumably ALL) in 415 LFS patients [151]	OR = ~60 [151] ^a	None	Hypodiploid ALL	OS, ACC, CNS, STS	1 in 5000 [152]–20,000 [153]

Neurofibromatosis 1 (NF1)	AD, <i>NF1</i>	12 NF1 patients (ages 1.25–14.5) in 5725 ALL cases [154]	OR(95% CI) = 5.4 (2.8–9.4) [154]	CALM, freckling, neurofibromas, learning disabilities		MPNST, JMML, MDS, GIST	1 in 2712 [155]- 3000 [156]
Nijmegen Breakage Syndrome (NBS)	AR, <i>NBN</i>	6 vs. 0 NBS cases in a ALL cases (389) vs. controls (7653) study [157]	OR(95%CI) = 1325 (860–2168) [157]	Microcephaly, short stature, pulm. Infections	T-ALL	NHL	Unknown, 170 cases documented [158]
PAX5 deficiency syndrome	AD, <i>PAX5</i>	11 B-ALLs in 21 PAX5 mutation carriers [159, 160]	Unknown, likely very high [159]	None	B-ALL	None	Unknown, 3 families documented [159, 160]

AD autosomal dominant, AR autosomal recessive, OR odds ratio, SIR standardized incidence ratio, *iAMP21* intrachromosomal amplification of chromosome 21, DS Down Syndrome, CI95%, 95% confidence interval, CALM café-au-lait macules, NHL non-Hodgkin lymphoma, BC breast cancer, OV ovarian cancer, STS soft tissue sarcoma, AML acute myeloid leukemia, GI gastrointestinal (cancers), PNET primitive neuroectodermal tumor, MB medullablastoma, TC testicular cancer, MDS myelodysplastic syndrome, genitourinary (cancers), ST solid tumors, OS osteosarcoma, ACC adrenocortical carcinoma, CNS central nervous system (tumors), MPNST malignant peripheral nerve sheath tumors, JMML juvenile myelomonocytic leukemia, GIST gastrointestinal stromal tumors

^aOR was estimated as the ratio of the reported ALL incidence in individuals with genetic syndrome with genetic syndrome of interest vs. incidence in the general population (i.e., 1 in 2000 between ages 0 and 15). For example, the frequency of ALL is ~5 per 1000 individuals with FA (7 in 1300 subjects) and thus ~11-fold higher than the expected ALL incidence in general populations

^bIncidences shown for ARs are for homozygote/biallelic status

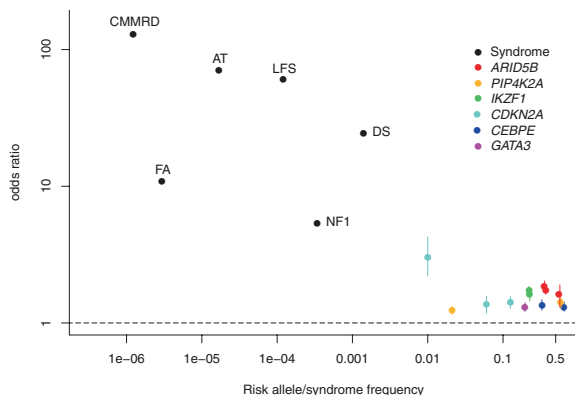


Fig. 1.2 Effect sizes and frequencies for known ALL genetic risk factors. Syndrome risks and frequencies are based on best available evidence as described in Table 1.3; SNP odds ratios are based on references in Table 1.4, and SNP risk allele frequencies are based on worldwide populations from the 1000 Genomes Project. CMMRD, constitutional mismatch repair-deficiency; AT ataxia-telangiectasia, LFS Li-Fraumeni syndrome, DS Down syndrome, FA Fanconi anemia, NF neurofibromatosis type 1

In fact, iAMP21-ALL has recently been found to be more frequent in individuals with the germline translocation $rob(15;21)(q10;q10)c$, a rare constitutional genetic abnormality. Amplification of the genes involved in the translocation duplicates the entire abnormal chromosome and confers an estimated 2,700-fold increased risk of iAMP21-ALL [138]. However, considering the rarity of both iAMP21-ALL and $rob(15;21)c$, <1 in 1,500 ALL cases are likely to be related to $rob(15;21)c$ associated.

PAX5 is known to be somatically mutated or deleted in approximately 30% of B-ALL cases [166]. In 2013, one germline *PAX5* mutation was found in three kindreds of familial ALL [159, 160]. The 3 families had 18 documented and 3 obligate mutation carriers with 11 cases of B-ALL, with another 2 ALLs in untested children. These *PAX5* mutations may be exclusively related to ALL risk, but further study is warranted.

ETV6, like *PAX5*, is known to be recurrently mutated or translocated in leukemic cells [166, 167]. In 2015, three studies independently reported nine families with *ETV6* germline mutations, all having a dominantly heritable thrombocytopenia and high incidence of ALL among mutation carriers [143, 144, 146]. Collectively, 35 documented and 4 obligate carriers have developed a total of 14 leukemias (mostly ALL), with another 2 occurring in untested children. One systematic sequencing study targeting germline *ETV6* in 4,405 ALL cases, identified 31 *ETV6* variants potentially related to 35 ALL cases, with carriers found to be significantly older than non-carriers (mean age 10.2 vs. 4.7) [144]. Thus, *ETV6* mutations may be present in nearly 1% of all ALL cases, and perhaps higher in patients over 5 years of age.

1.6.2 Syndromes Where ALL Is Part of a Mixed Cancer Phenotype

Li-Fraumeni Syndrome (LFS) is a rare cancer predisposition syndrome, in which germline *TP53* mutation confers a ~90% lifetime risk of developing cancer in a spectrum of tissues with one third being diagnosed before 18 years of age. The increased ALL risk is largely restricted to cases with low hypodiploid leukemia karyotype (underlying *TP53* mutation present in 43.3% of low hypodiploid ALL) [168].

Ataxia-telangiectasia (A-T) is a rare syndrome caused by recessive mutations in the *ATM* gene and typically presents with progressive cerebellar ataxia before 4 years of age [134]. A-T patients have a high risk of leukemias (especially T-cell ALL) and lymphomas, as well as hypersensitivity to ionizing radiation and chemotherapy related to the role of ATM in DNA repair [132, 134].

Bloom Syndrome is characterized by pre- and postnatal growth deficiency (stature typically <1.5 m), skin lesions and high risk of ALL, AML, lymphoma, and epithelial carcinomas [135]. Twelve ALLs were found in less than the 300 cases registered world-wide and in at least two cases ALL preceded Bloom Syndrome diagnosis [169, 170].

Nijmegen Breakage Syndrome (NBS) is another very rare recessive syndrome, which mainly occurs in Slavic populations [157] (a Slavic founder deletion of five bases in the *NBN* gene is found in >90% of NBS cases), yet NBS has also been described in >8 other countries with private mutations [157, 158, 171, 172]. Patients display microcephaly, intrauterine growth retardation with short stature, recurrent sinopulmonary infections and increased risk of cancers, especially lymphoma and leukemia [157, 173].

Fanconi Anemia (FA) is a rare recessive syndrome with a high risk of AML, MDS and other hematological diseases set at ~10%/year [147, 174]. In a registry with 1300 FA patients only 7 ALLs were reported and FA-leukemias are predominantly myeloid (96%) [147, 148]. While skeletal deformations and classic hematological findings often lead to diagnosis early in life, malignancies including ALL can be the presenting feature [175, 176].

There is a long string of genetic syndromes for which sporadic reports described ALL as a possible cancer manifestation, although the matter has not been systematically examined. The most common are RASopathies (e.g. NF1) [177–179], where 6 ALLs were seen among 1176 mutation carriers in 1 study [180]. Others include: Bruton's Agammaglobulinemia [181], Familial Platelet Disorder with Associated Myeloid Malignancies [182, 183], Weaver syndrome [184], Sotos syndrome [185], Rubinstein-Taybi syndrome [186], Börjeson-Forssman-Lehmann Syndrome [187] and SH2B3 deficiency [188].

It should be noted that ALL predisposition syndrome may not be symptomatic prior to leukemia diagnosis with only non-specific clinical features such as growth failure and microcephaly. Family history needs to be carefully examined to identify possible underlying genetic causes in a pediatric oncology setting.

1.7 Low-Penetrance Genetic Predisposition

Emerging from the ‘common disease—common variant’ hypothesis, the past two decades have seen the application of first candidate gene-driven and later genome-wide association studies in ALL etiology research [189, 190].

Single nucleotide polymorphism (SNP)-based candidate gene studies (CGSs) have explored ALL etiology by focusing on genes involved in carcinogen metabolism, folate metabolism, and DNA repair pathways. A 2010 systematic review identified 47 CGSs on 25 variations in 16 genes all tested for association with ALL, showing pooled significance ($P < 0.05$) in only 8 variants (OR range; 0.73–1.78) with an apparent false positive report probabilities of at least 20% [191]. Other studies have focused on human leukocyte antigen (HLA) genes, particularly class II loci *HLA-DR* and *HLA-DP*, with the latter showing evidence of significantly different associations between ALL subtypes as well as interactions with proxies for immune stimulation [192, 193]. However, a larger study has cast doubt on the validity of these findings [194].

2009 saw the first two genome-wide association studies (GWAS) independently demonstrating associations between ALL susceptibility and SNPs in *ARID5*, *IKZF1* and *CEBPE* [195, 196]. Subsequently, SNPs in four other genes have been found to be associated with either overall ALL risk or subtype-specific risk, with a total of 13 SNPs in 6 genes having been widely validated thus far (Table 1.4) [197–201].

Table 1.4 GWAS results

Gene	rsid(reference)	OR(95% CI)	Associated subtype, OR(95% CI)	RAF
<i>ARID5B</i>	rs7089424 [202]	1.65 (1.54–1.76)	HeH, 2.17 (1.5–3.1)	0.37
	rs10821936 [196]	1.91 (1.6–2.2)		0.36
	rs10994982 [197]	1.86 (1.71–2.03)		0.57
<i>CDKN2A</i>	rs17756311 [197]	1.36 (1.18–1.56)		0.06
	rs3731217 [203]	0.71 (0.64–0.78)		0.13
	rs3731249 [204]	2.99 (2.21–4.26)		0.01
<i>CEBPE</i>	rs2239633 [202]	1.31 (1.22–1.42)		0.64
	rs4982731 [197]	1.36 (1.24–1.48)		0.33
<i>GATA3</i>	rs3824662 [198]	1.31 (1.21–1.41)	Ph-like, 3.85 (2.7–5.4)	0.20
<i>IKZF1</i>	rs4132601 [202]	1.71 (1.58–1.85)		0.22
	rs11978267 [197]	1.59 (1.45–1.74)		0.23
<i>PIP4K2A</i>	rs7088318 [197]	1.40 (1.28–1.53)	HeH	0.59
	rs10828317 [198]	1.23 (1.15–1.32)		0.02

Selected SNPs associated with ALL risk. Risk allele frequencies are global frequencies from the 1000 Genomes Project. *SNP* single nucleotide polymorphism, *HeH* high-hyperdiploid, *OR* odds ratio, 95% CI, 95% confidence interval, *RAF* risk allele frequency, *Ph-like* Philadelphia-like ALL

The heterogeneity of ALL is reflected in the GWAS findings, with some SNPs showing a stronger association with specific subtypes. *ARID5B*, for instance, is most strongly associated with HeH ALL. SNPs in *TP63* and *GATA3*, on the other hand, show isolated associations with *ETV6-RUNX1* ALL and Ph-like ALL, respectively [196, 199, 201, 204, 205].

While there is little doubt that the GWAS findings identified genuine inherited risk factors for ALL, there is a paucity of studies describing the molecular mechanisms underlying these associations. Somatic deletions in both *CDKN2A* and *IKZF1* are frequent in ALL, and these two genes play important roles in tumor suppression and lymphocyte development, respectively [200, 206]. In one recent study, 35 tumors from *CDKN2A* risk variant rs3731217 carriers preferentially retained the risk allele, suggesting that the SNP is advantageous during tumor growth [204]. *ARID5B* is also involved in lymphocyte differentiation, but its mechanism in ALL development is poorly understood.

Within the validated risk variants, no significant gene–gene interactions have been reported [195, 196, 203]. The effects of these risk alleles are relatively stable across ethnicities, and risk allele frequencies correlate well with population differences in ALL incidence [197]. One pathway-based GWAS on ALL risk was recently described but these results have yet to be reproduced [207]. Inspired by the observations that ALL subtypes differ across both environmental and genetic risk factors, other researchers have attempted to identify interactions between the two by combining genotypes with data on various environmental exposures [208–211]. These studies, however, have so far failed to reliably identify gene–environment interactions.

Studies on childhood ALL etiology will improve knowledge of the pathogenesis, predict disease risk, and provide new targets for treatment.

The low-penetrance genetic predispositions discussed above (e.g., risk alleles identified by GWAS) constitute a minor increase in the absolute risk of developing ALL, e.g. from 1 in 2,000 to 1 in 1,500. While the effects of these variants individually are modest with limited clinical implication, their cumulative impact can be comparable to those of the highly penetrant genetic predisposition syndromes. However, it is debatable whether early diagnosis of an aggressive cancer like ALL can lead to improved outcome [212]. Hence, clinical surveillance aimed at early diagnosis of ALL may not necessarily benefit at-risk subjects and may in fact lead to uncertainty and anxiety for the families [213].

Still, many of the genetic syndromes discussed above may modify health conditions other than the risk of developing ALL. Preemptive surveillance for non-ALL cancers (e.g. *TP53* carriers) and/or treatment modification (e.g. avoidance of radiation therapy in cases with A-T) can lead to lower mortality and morbidity for the children and their at-risk family members [214–218]. For this reason, recognition and diagnosis of predisposition syndromes in pediatric oncology is crucial. In fact, it has been suggested that pediatric cancer patients under the age of five should be evaluated for A-T before starting chemotherapy and/or radiotherapy because of potentially fatal adverse effects of conventional doses due to defective DNA repair in these cases [134, 219].

1.8 Future Directions

While substantial progress has been made in identifying risk factors for ALL (especially the role of inherited genetic variants), our understanding of ALL disease etiology is far from complete. An important field of research in the coming years will be to identify gene-gene and gene-environment interactions that contribute to ALL leukemogenesis, and whether approaches can be developed to target these processes and reduce disease risk and burden in genetically predisposed children.

References

1. SEER Cancer Statistics review 1975–2012 [Internet]. National Cancer Institute; [cited 2016 Feb 29]. Available from: http://seer.cancer.gov/csr/1975_2012/browse_csr.php.
2. Hjalgrim LL, Rostgaard K, Schmiegelow K, Soderhall S, Kolmannskog S, Vetterranta K, et al. Age- and sex-specific incidence of childhood leukemia by immunophenotype in the nordic countries. *JNCI J Natl Cancer Inst.* 2003;95(20):1539–44.
3. Linabery AM, Ross JA. Trends in childhood cancer incidence in the U.S. (1992–2004). *Cancer.* 2008;112(2):416–32.
4. Parkin DM, Kramárová E, Draper GJ, Masuyer E, et al. International incidence of childhood cancer. Lyon: IARC Scientific Publications; 1998.
5. Stiller CA, Parkin DM. Geographic and ethnic variations in the incidence of childhood cancer. *Br Med Bull.* 1996;52(4):682–703.
6. Schmiegelow K, Vestergaard T, Nielsen SM, Hjalgrim H. Etiology of common childhood acute lymphoblastic leukemia: the adrenal hypothesis. *Leukemia.* 2008;22(12):2137–41.
7. Greaves MF, Colman SM, Beard ME, Bradstock K, Cabrera ME, Chen PM, et al. Geographical distribution of acute lymphoblastic leukaemia subtypes: second report of the collaborative group study. *Leukemia.* 1993;7(1):27–34.
8. Biondi A, Cimino G, Pieters R, Pui CH. Biological and therapeutic aspects of infant leukemia. *Blood.* 2000;96(1):24–33.
9. Pieters R, Schrappe M, De Paola L, Hann I, De Giulio R, Felice M, et al. A treatment protocol for infants younger than 1 year with acute lymphoblastic leukemia (Interfant-99): an observational study and a multicenter randomized trial. *Lancet.* 2007;370(9583):240–50.
10. Szczepanski T, Harrison CJ, van Dongen JJM. Genetic aberrations in paediatric acute leukaemias and implications for management of patients. *Lancet Oncol Elsevier Ltd.* 2010;11(9):880–889.
11. Forestier E, Schmiegelow K. The incidence peaks of the childhood acute leukemias reflect specific cytogenetic aberrations. *J Pediatr Hematol Oncol.* 2006;28(8):486–95.
12. Rajalekshmy KR, Abitha AR, Pramila R, Gnanasagar T, Shanta V. Immunophenotypic analysis of T-cell acute lymphoblastic leukaemia in Madras, India. *Leuk Res.* 1997; 21(2):119–24.
13. Gmidène A, Sennana H, Elghezal H, Ziraoui S, Youssef YB, Elloumi M, Issaoui L, Harrabi I, Raynaud SSA. Cytogenetic analysis of 298 newly diagnosed cases of acute lymphoblastic leukaemia in Tunisia. *Hematol Oncol.* 2008;26:91–7.
14. Williams CK. Childhood leukemia and lymphoma: African experience supports a role for environmental factors. *Cancer Res.* 2012;72(8 Suppl):5484.
15. Liang D-C, Shih L-Y, Yang C-P, Hung I-J, Liu H-C, Jaing T-H, et al. Frequencies of ETV6–RUNX1 fusion and hyperdiploidy in pediatric acute lymphoblastic leukemia are lower in far East than West. *Pediatr Blood Cancer.* 2010;55:430–3.

16. McNally RJQ, Birch JM, Taylor GM, Eden OB. Incidence of childhood precursor B-cell acute lymphoblastic leukaemia in north-west England. *Lancet*. 2000;356:484–5.
17. Hrusak O, Trka J, Zuna J, Polouckova A, Kalina T, Stary J. Acute lymphoblastic leukemia incidence during socioeconomic transition: selective increase in children from 1 to 4 years. *Leukemia*. 2002;16:720–5.
18. Fraumeni JF, Miller RW. Epidemiology of human leukemia. *J Natl Cancer Inst*. 1967;38(4):593–603.
19. Spix C, Eletr D, Blettner M, Kaatsch P. Temporal trends in the incidence rate of childhood cancer in Germany 1987–2004. *Int J Cancer*. 2008;122(8):1859–67.
20. Greaves MF, Maia AT, Wiemels JL, Ford AM. Leukemia in twins: lessons in natural history. *Blood*. 2013;102(7):2321–33.
21. Ford AM, Ridge SA, Cabrera ME, Mahmoud H, Steel CM, Chan LC, et al. In utero rearrangements in the trithorax-related oncogene in infant leukaemias. *Nature*. 1993;363(6427):358–60.
22. Gill Super HJ, Rothberg PG, Kobayashi H, Freeman AI, Diaz MO, Rowley JD. Clonal, non-constitutional rearrangements of the MLL gene in infant twins with acute lymphoblastic leukemia: in utero chromosome rearrangement of 11q23. *Blood*. 1994;83(3):641–4.
23. Maia AT, van der Velden VHJ, Harrison CJ, Szczepanski T, Williams MD, Griffiths MJ, et al. Prenatal origin of hyperdiploid acute lymphoblastic leukemia in identical twins. *Leukemia*. 2003;17(11):2202–6.
24. Kadan-Lottick NS, Kawashima T, Tomlinson G, Friedman DL, Yasui Y, Mertens AC, et al. The risk of cancer in twins: a report from the childhood cancer survivor study. *Pediatr Blood Cancer*. 2006;46(5):476–81.
25. Ford AM, Pombo-de-Oliveira MS, McCarthy KP, MacLean JM, Carrico KC, Vincent RF, et al. Monoclonal origin of concordant T-cell malignancy in identical twins. *Blood*. 1997;89(1):281–5.
26. Gale KB, Ford AM, Repp R, Borkhardt A, Keller C, Eden OB, et al. Backtracking leukemia to birth: identification of clonotypic gene fusion sequences in neonatal blood spots. *Proc Natl Acad Sci U S A*. 1997;94(25):13950–4.
27. Wiemels JL, Cazzaniga G, Daniotti M, Eden OB, Addison GM, Masera G, et al. Prenatal origin of acute lymphoblastic leukemia in children. *Lancet*. 1999;354(9189):1499–503.
28. Fasching K, Panzer S, Haas OA, Marschalek R, Gadner H, Panzer-Grümayer ER. Presence of clone-specific antigen receptor gene rearrangements at birth indicates an in utero origin of diverse types of early childhood acute lymphoblastic leukemia. *Blood*. 2000;95(8):2722–4.
29. Yagi T, Hibi S, Tabata Y, Kuriyama K, Teramura T, Hashida T, et al. Detection of clonotypic IGH and TCR rearrangements in the neonatal blood spots of infants and children with B-cell precursor acute lymphoblastic leukemia. *Blood*. 2000;96(1):264–8.
30. Maia AT, Ford AM, Reza Jalali G, Harrison CJ, Malcolm Taylor G, Eden OB, et al. Molecular tracking of leukemogenesis in a triplet pregnancy. *Blood*. 2001;98(2):478–82.
31. Renate Panzer-Grümayer E, Fasching K, Panzer S, Hettinger K, Schmitt K, Stöckler-Ipsiroglu S, et al. Nondisjunction of chromosomes leading to hyperdiploid childhood B-cell precursor acute lymphoblastic leukemia is an early event during leukemogenesis. *Blood*. 2002;100(1):347–9.
32. Taub JW, Konrad MA, Ge Y, Naber JM, Scott JS, Matherly LH, et al. High frequency of leukemic clones in newborn screening blood samples of children with B-precursor acute lymphoblastic leukemia. *Blood*. 2002;99(8):2992–6.
33. Hjalgrim LL, Madsen HO, Melbye M, Jørgensen P, Christiansen M, Andersen MT, et al. Presence of clone-specific markers at birth in children with acute lymphoblastic leukaemia. *Br J Cancer*. 2002;87(9):994–9.
34. Wiemels JL, Leonard BC, Wang Y, Segal MR, Hunger SP, Smith MT, et al. Site-specific translocation and evidence of postnatal origin of the t(1;19) E2A-PBX1 fusion in childhood acute lymphoblastic leukemia. *Proc Natl Acad Sci U S A*. 2002;99(23):15101–6.
35. McHale CM, Wiemels JL, Zhang L, Ma X, Buffler PA, Guo W, et al. Prenatal origin of TEL-AML1-positive acute lymphoblastic leukemia in children born in California. *Genes Chromosom Cancer*. 2003;37(1):36–43.

36. Teuffel O, Betts DR, Dettling M, Schaub R, Schäfer BW, Niggli FK. Prenatal origin of separate evolution of leukemia in identical twins. *Leukemia*. 2004;18(10):1624–9.
37. Maia AT, Koechling J, Corbett R, Metzler M, Wiemels JL, Greaves M. Protracted postnatal natural histories in childhood leukemia. *Genes Chromosom Cancer*. 2004;39(4):335–40.
38. Maia AT, Tussiwand R, Cazzaniga G, Rebutta P, Colman S, Biondi A, et al. Identification of preleukemic precursors of hyperdiploid acute lymphoblastic leukemia in cord blood. *Genes Chromosom Cancer*. 2004;40(1):38–43.
39. Fischer S, Mann G, Konrad M, Metzler M, Ebetsberger G, Jones N, et al. Screening for leukemia- and clone-specific markers at birth in children with T-cell precursor ALL suggests a predominantly postnatal origin. *Blood*. 2007;110(8):3036–8.
40. Gruhn B, Taub JW, Ge Y, Beck JF, Zell R, Häfer R, et al. Prenatal origin of childhood acute lymphoblastic leukemia, association with birth weight and hyperdiploidy. *Leukemia*. 2008;22(9):1692–7.
41. Wiemels JL, Kang M, Chang JS, Zheng L, Kouyoumji C, Zhang L, et al. Backtracking RAS mutations in high hyperdiploid childhood acute lymphoblastic leukemia. *Blood Cells Mol Dis*. 2010;45(3):186–91.
42. Eguchi-ishimae M, Eguchi M, Kempinski H, Greaves M, De W. NOTCH 1 mutation can be an early, prenatal genetic event in T-ALL brief report NOTCH1 mutation can be an early, prenatal genetic event in T-ALL. *Blood*. 2011;111(1):376–8.
43. Mansur MB, van Delft FW, Colman SM, Furness CL, Gibson J, Emerenciano M, et al. Distinctive genotypes in infants with T-cell acute lymphoblastic leukaemia. *Br J Haematol*. 2015;171(4):574–84.
44. Hong D, Gupta R, Ancliff P, Atzberger A, Brown J, Soneji S, Green J, Colman S, Piacibello W, Buckle V, Tsuzuki S, Mel Greaves TE. Initiating and cancer-propagating cells in TEL-AML1-associated childhood leukemia. *Science* (80-). 2008;339:1095–9.
45. Tsuzuki S, Seto M, Greaves M, Enver T. Modeling first-hit functions of the t(12;21) TEL-AML1 translocation in mice. *Proc Natl Acad Sci U S A*. 2004;101(22):8443–8.
46. Fischer M, Schwieger M, Horn S, Niebuhr B, Ford A, Roscher S, et al. Defining the oncogenic function of the TEL/AML1 (ETV6/RUNX1) fusion protein in a mouse model. *Oncogene*. 2005;24(51):7579–91.
47. Wiemels JL, Ford AM, Van Wering ER, Postma A, Greaves M. Protracted and variable latency of acute lymphoblastic leukemia after TEL-AML1 gene fusion in utero. *Blood*. 1999;94(3):1057–62.
48. Lilljebjorn H, Heidenblad M, Nilsson B, Lassen C, Horvat A, Heldrup J, et al. Combined high-resolution array-based comparative genomic hybridization and expression profiling of ETV6/RUNX1-positive acute lymphoblastic leukemias reveal a high incidence of cryptic Xq duplications and identify several putative target genes within the c. *Leukemia*. 2007;21(10):2137–44.
49. Tsuzuki S, Karnan S, Horibe K, Matsumoto K, Kato K, Inukai T, et al. Genetic abnormalities involved in t(12;21) TEL-AML1 acute lymphoblastic leukemia: analysis by means of array-based comparative genomic hybridization. *Cancer Sci*. 2007;98(5):698–706.
50. Mullighan CG, Goorha S, Radtke I, Miller CB, Coustan-Smith E, Dalton JD, et al. Genome-wide analysis of genetic alterations in acute lymphoblastic leukaemia. *Nature*. 2007;446(7137):758–64.
51. Bateman CM, Colman SM, Chaplin T, Young BD, Eden TO, Bhakta M, et al. Acquisition of genome-wide copy number alterations in monozygotic twins with acute lymphoblastic leukemia. *Leukemia*. 2010;115(17):3553–8.
52. Ma Y, Dobbins SE, Sherborne AL, Chubb D, Galbiati M, Cazzaniga G, et al. Developmental timing of mutations revealed by whole-genome sequencing of twins with acute lymphoblastic leukemia. *Proc Natl Acad Sci U S A*. 2013;110(18):7429–33.
53. Mori H, Colman SM, Xiao Z, Ford AM, Healy LE, Donaldson C, et al. Chromosome translocations and covert leukemic clones are generated during normal fetal development. *Proc Natl Acad Sci U S A*. 2002;99(12):8242–7.

54. Greaves M, Colman SM, Kearney LFA. To the editor: fusion genes in cord blood. *Blood*. 2011;117(1):369–70.
55. Schmiegelow K, Lausten-Thomsen U, Madsen HO, Nersting J, Hjalgrim H. Challenges and pitfalls in the mapping of the natural history of t(12;21)-positive childhood ALL. *Blood*. 2011;117(1):370–2.
56. Kusk MS, Lausten-Thomsen U, Andersen MK, Olsen M, Hjalgrim H, Schmiegelow K. False positivity of ETV6/RUNX1 detected by FISH in healthy newborns and adults. *Pediatr Blood Cancer*. 2014;61:1704–6.
57. Lausten-thomsen U, Madsen HO, Vestergaard TR, Hjalgrim H, De W, Nersting J. Prevalence of t(12;21)[ETV6-RUNX1]-positive cells in healthy neonates. *Blood*. 2012;117(1):186–9.
58. Zuna J, Madzo J, Krejci O, Zemanova Z, Kalinova M, Muzikova K, Zapotocky M, Starkova J, Hrusak O, Horak J, Trka J. ETV6/RUNX1 (TEL/AML1) is a frequent prenatal first hit in childhood leukemia. *Blood*. 2011;117(1):369–70.
59. Bateman CM, Alpar D, Ford AM, Colman SM, Wren D, Morgan M, et al. Evolutionary trajectories of hyperdiploid ALL in monozygotic twins. *Leukemia*. Nature Publishing Group. 2014;29(1):58–65.
60. Paulsson K, Lilljebjörn H, Biloglav A, Olsson L, Rissler M, Castor A, et al. The genomic landscape of high hyperdiploid childhood acute lymphoblastic leukemia. *Nat Genet* Nature Publishing Group. 2015;47(6):672–677.
61. Kinlen L. Evidence for an infective cause of childhood leukaemia: comparison of a Scottish new town with nuclear reprocessing sites in Britain. *Lancet*. 1988;332(8624):1323–7.
62. Kreis C, Grotzer M, Hengartner H, Spycher BD, Spycher BD. Space-time clustering of childhood cancers in Switzerland: a nationwide study. *Int J Cancer*. 2016;2135:2127.
63. Greaves MF. Speculations on the cause of childhood acute lymphoblastic leukemia. *Leukemia*. 1988;2(2):120–5.
64. Greaves M. Infection, immune responses and the aetiology of childhood leukaemia. *Nat Rev Cancer*. 2006;6(3):193–203.
65. Wiemels J. Perspectives on the causes of childhood leukemia. *Chem Biol Interact*. Elsevier Ireland Ltd. 2012;196(3):59–67.
66. Strachan DP. Hay fever, hygiene, and household size. *Br Med J*. 1989;299(November):1259–60.
67. Greaves M. The “delayed infection” (aka “hygiene”) hypothesis for childhood leukaemia. In: *The hygiene hypothesis and darwinian medicine*. Birkhauser Verlag Basel, Switzerland. 2009. p. 239–55.
68. Osterholm MT. Infectious disease in child day care: an overview. *Pediatrics*. 1994; 94:987–90.
69. Urayama KY, Buffler PA, Gallagher ER, Ayoob JM, Ma X. A meta-analysis of the association between day-care attendance and childhood acute lymphoblastic leukaemia. *Int J Epidemiol*. 2010;39(3):718–32.
70. Rudant J, Lightfoot T, Urayama KY, Petridou E, Dockerty JD, Magnani C, et al. Childhood acute lymphoblastic leukemia and indicators of early immune stimulation: a childhood leukemia international consortium study. *Am J Epidemiol*. 2015;181(8):549–62.
71. Kwan ML, Buffler PA, Abrams B, Kiley VA. Breastfeeding and the risk of childhood leukemia: a meta-analysis. *Public Health Rep*. 2004;119(6):521–35.
72. Martin RM, Gunnell D, Owen CG, Smith GD. Breast-feeding and childhood cancer: a systematic review with metaanalysis. *Int J Cancer*. 2005;117(6):1020–31.
73. Altieri A, Castro F, Bermejo JL, Hemminki K. Number of siblings and the risk of lymphoma, leukemia, and myeloma by histopathology. *Cancer Epidemiol Biomark Prev*. 2006;15(7): 1281–6.
74. Von Behren J, Spector LG, Mueller BA, Carozza SE, Chow EJ, Fox EE, et al. Birth order and risk of childhood cancer: a pooled analysis from five US States. *Int J Cancer*. 2011;128(11): 2709–16.
75. Ma X, Does MB, Metayer C, Russo C, Wong A, Buffler PA. Vaccination history and risk of childhood leukaemia. *Int J Epidemiol*. 2005;34(5):1100–9.

76. Auvinen A, Hakulinen T, Groves F. Haemophilus influenzae type B vaccination and risk of childhood leukaemia in a vaccine trial in Finland. *Br J Cancer*. 2000;83(7):956–8.
77. Groves FD, Gridley G, Wacholder S, Shu XO, Robison LL, Neglia JP, et al. Infant vaccinations and risk of childhood acute lymphoblastic leukaemia in the USA. *Br J Cancer*. 1999;81(1):175–8.
78. MacArthur AC, McBride ML, Spinelli JJ, Tamaro S, Gallagher RP, Theriault GP. Risk of childhood leukemia associated with vaccination, infection, and medication use in childhood: the Cross-Canada childhood leukemia study. *Am J Epidemiol*. 2008;167(5):598–606.
79. Mallol-Mesnard N, Menegaux F, Auvignon A, Auclerc MF, Bertrand Y, Nelken B, et al. Vaccination and the risk of childhood acute leukaemia: the ESCALE study (SFCE). *Int J Epidemiol*. 2007;36(1):110–6.
80. Chang JS, Tsai CR, Tsai YW, Wiemels JL. Medically diagnosed infections and risk of childhood leukaemia: a population-based case-control study. *Int J Epidemiol*. 2012;41(4):1050–9.
81. Cardwell CR, McKinney PA, Patterson CC, Murray LJ. Infections in early life and childhood leukaemia risk: a UK case-control study of general practitioner records. *Br J Cancer*. 2008;99(9):1529–33.
82. Roman E, Simpson J, Ansell P, Kinsey S, Mitchell CD, McKinney PA, et al. Childhood acute lymphoblastic leukemia and infections in the first year of life: a report from the United Kingdom Childhood Cancer Study. *Am J Epidemiol*. 2007;165(5):496–504.
83. Vestergaard T, Rostgaard K, Grau K, Schmiegelow K, Hjalgrim H. Hospitalisation for infection prior to diagnosis of acute lymphoblastic leukaemia in children. *Pediatr Blood Cancer*. 2013;60:428–32.
84. Crouch S, Lightfoot T, Simpson J, Smith A, Ansell P, Roman E. Infectious illness in children subsequently diagnosed with acute lymphoblastic leukemia: modeling the trends from birth to diagnosis. *Am J Epidemiol*. 2012;176(5):402–8.
85. Jourdan-Da Silva N, Perel Y, Méchinaud F, Plouvier E, Gandemer V, Lutz P, et al. Infectious diseases in the first year of life, perinatal characteristics and childhood acute leukaemia. *Br J Cancer*. 2004;90(1):139–45.
86. Rosenbaum PF, Buck GM, Brecher ML. Allergy and infectious disease histories and the risk of childhood acute lymphoblastic leukaemia. *Paediatr Perinat Epidemiol*. 2005;19(2):152–64.
87. Neglia JP, Linet MS, Shu XO, Severson RK, Potter JD, Mertens AC, et al. Patterns of infection and day care utilization and risk of childhood acute lymphoblastic leukaemia. *Br J Cancer*. 2000;82(1):234–40.
88. Perillat F, Clavel J, Auclerc MF, Baruchel A, Leverger G, Nelken B, Philippe N, Schaison G, Sommelet D, Vilmer EHD. Day-care, early common infections and childhood acute leukaemia: a multicentre French case-control study. *Br J Cancer*. 2002;86(7):1064–9.
89. Urayama KY, Ma X, Selvin S, Metayer C, Chokkalingam AP, Wiemels JL, et al. Early life exposure to infections and risk of childhood acute lymphoblastic leukemia. *Int J Cancer*. 2011;128(7):1632–43.
90. Papaemmanuil E, Rapado I, Li Y, Potter NE, Wedge DC, Tubio J, et al. RAG-mediated recombination is the predominant driver of oncogenic rearrangement in ETV6-RUNX1 acute lymphoblastic leukemia. *Nat Genet* Nature Publishing Group. 2014;46(2):116–125.
91. Swaminathan S, Klemm L, Park E, Papaemmanuil E, Ford A, Kweon S-M, et al. Mechanisms of clonal evolution in childhood acute lymphoblastic leukemia. *Nat Immunol*. 2015;16(7):766–74.
92. Martin-Lorenzo A, Hauer J, Vicente-Duenas C, Auer F, Gonzalez-Herrero I, Garcia-Ramirez I, et al. Infection exposure is a causal factor in B-precursor acute lymphoblastic leukemia as a result of Pax5 inherited susceptibility. *Cancer Discov*. 2015;5:1328–43.
93. Hjalgrim LL, Westergaard T, Rostgaard K, Schmiegelow K, Melbye M, Hjalgrim H, et al. Birth weight as a risk factor for childhood leukemia: a meta-analysis of 18 epidemiologic studies. *Am J Epidemiol*. 2003;158(8):724–35.

94. Doll R, Wakeford R. Risk of childhood cancer from fetal irradiation. *Br J Radiol. The British Institute of Radiology.* 1997;70(830):130–139.
95. Mathews JD, Forsythe AV, Brady Z, Butler MW, Goergen SK, Byrnes GB, et al. Cancer risk in 680,000 people exposed to computed tomography scans in childhood or adolescence: data linkage study of 11 million Australians. *BMJ.* 2013;346:f2360.
96. Wakeford R. The cancer epidemiology of radiation. *Oncogene.* 2004;23(38):6404–28.
97. Brenner DJ, Doll R, Goodhead DT, Hall EJ, Land CE, Little JB, et al. Cancer risks attributable to low doses of ionizing radiation: assessing what we really know. *Proc Natl Acad Sci U S A.* 2003;100(24):13761–6.
98. Wakeford R, Kendall GM, Little MP. The proportion of childhood leukaemia incidence in Great Britain that may be caused by natural background ionizing radiation. *Leukemia Nature Publishing Group.* 2009;23(4):770–776.
99. Raaschou-Nielsen O, Andersen CE, Andersen HP, Gravesen P. Domestic radon and childhood cancer in Denmark. *Epidemiology.* 2008;19(4):536–43.
100. Tong J, Qin L, Cao Y, Li J, Zhang J, Nie J, et al. Environmental radon exposure and childhood leukemia. *J Toxic Environ Health B.* 2012;15(5):332–47.
101. Hauri D, Spycher B, Huss A, Zimmermann F, Grotzer M, von der Weid N, Weber D, Spoerri A, Kuehni CRM. Domestic radon exposure and risk of childhood cancer: a prospective census-based cohort study. *Environ Health Perspect.* 2013;121(10):1239–44.
102. Schüz J, Ahlbom A. Exposure to electromagnetic fields and the risk of childhood leukaemia: a review. *Radiat Prot Dosim.* 2008;132(2):202–11.
103. Schüz J. Exposure to extremely low-frequency magnetic fields and the risk of childhood cancer: update of the epidemiological evidence. *Prog Biophys Mol Biol.* 2011;107(3):339–42.
104. Schüz J, Dasenbrock C, Ravazzani P, Rössli M, Schär P, Bounds P, et al. Extremely low-frequency magnetic fields and the risk of childhood leukemia: a risk assessment by the ARIMMORA consortium. *Environ Health Perspect.* 2015;37:1–7.
105. Merzenich H, Schmiedel S, Bennack S, Brüggemeyer H, Philipp J, Blettner M, et al. Childhood leukemia in relation to radio frequency electromagnetic fields in the vicinity of TV and radio broadcast transmitters. *Am J Epidemiol.* 2008;168(10):1169–78.
106. Elliott P, Toledano MB, Bennett J, Beale L, de Hoogh K, Best N, et al. Mobile phone base stations and early childhood cancers: case-control study. *BMJ.* 2010;340:c3077.
107. Sergentanis TN, Thomopoulos TP, Gialamas SP, Karalexi MA, Biniaris-Georgallis SI, Kontogeorgi E, et al. Risk for childhood leukemia associated with maternal and paternal age. *Eur J Epidemiol. Springer Netherlands.* 2015;30(12):1229–1261.
108. Yan K, Xu X, Xu X, Liu X, Wang X, Hua S, et al. The associations between maternal factors during pregnancy and the risk of childhood acute lymphoblastic leukemia: a meta-analysis. *Pediatr Blood Cancer.* 2015;14(5):1526–31.
109. Orsi L, Rudant J, Ajrouche R, Leverger G, Baruchel A, Nelken B, et al. Parental smoking, maternal alcohol, coffee and tea consumption during pregnancy, and childhood acute leukemia: the ESTELLE study. *Cancer Causes Control Springer International Publishing.* 2015;26(7):1003–1017.
110. Ferreira JD, Couto AC, Emerenciano M, Pombo-de-Oliveira MS, Koifman S. Maternal alcohol consumption during pregnancy and early age leukemia risk in Brazil. *Biomed Res Int.* 2015;2015:732495.
111. Milne E, Greenop KR, Scott RJ, de Klerk NH, Bower C, Ashton LJ, et al. Parental alcohol consumption and risk of childhood acute lymphoblastic leukemia and brain tumors. *Cancer Causes Control.* 2013;24(2):391–402.
112. Liu R, Zhang L, McHale CM, Hammond SK. Paternal smoking and risk of childhood acute lymphoblastic leukemia: systematic review and meta-analysis. *J Oncol.* 2011;2011:16.
113. Dockerty JD, Herbison P, Skegg DCG, Elwood M. Vitamin and mineral supplements in pregnancy and the risk of childhood acute lymphoblastic leukaemia: a case-control study. *BMC Public Health.* 2007;7:136.