

Ratan Kumar Choudhary  
Shanti Choudhary *Editors*

# Stem Cells in Veterinary Science

 Springer

---

# Stem Cells in Veterinary Science

---

Ratan Kumar Choudhary •  
Shanti Choudhary  
Editors

# Stem Cells in Veterinary Science

 Springer

*Editors*

Ratan Kumar Choudhary  
Department of Bioinformatics  
College of Animal Biotechnology  
Ludhiana, Punjab, India

Shanti Choudhary  
Department of Microbial and Environmental  
Biotechnology  
College of Animal Biotechnology  
Ludhiana, Punjab, India

ISBN 978-981-16-3463-5

ISBN 978-981-16-3464-2 (eBook)

<https://doi.org/10.1007/978-981-16-3464-2>

© The Editor(s) (if applicable) and The Author(s), under exclusive license to Springer Nature Singapore Pte Ltd. 2021

This work is subject to copyright. All rights are solely and exclusively licensed by the Publisher, whether the whole or part of the material is concerned, specifically the rights of translation, reprinting, reuse of illustrations, recitation, broadcasting, reproduction on microfilms or in any other physical way, and transmission or information storage and retrieval, electronic adaptation, computer software, or by similar or dissimilar methodology now known or hereafter developed.

The use of general descriptive names, registered names, trademarks, service marks, etc. in this publication does not imply, even in the absence of a specific statement, that such names are exempt from the relevant protective laws and regulations and therefore free for general use.

The publisher, the authors, and the editors are safe to assume that the advice and information in this book are believed to be true and accurate at the date of publication. Neither the publisher nor the authors or the editors give a warranty, expressed or implied, with respect to the material contained herein or for any errors or omissions that may have been made. The publisher remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This Springer imprint is published by the registered company Springer Nature Singapore Pte Ltd. The registered company address is: 152 Beach Road, #21-01/04 Gateway East, Singapore 189721, Singapore

---

## Preface

Stem cells have high hopes for regenerative medicine in human health and treating animal diseases. In recent years, it has gained significant momentum. Stem cells are unspecialized cells with profound self-renewal capability and differentiation ability into cells with specialized functions. Therefore, stem cell therapies are used to repair the body's inability to regenerate damaged tissues after acute or chronic ailments. Among the various stem cell types, mesenchymal stem cells hold promising therapeutic applications in treating many diseases because of the simplicity of isolation from various tissue sources and the lack of ethical concern regarding their usage. With the paucity of universal stem cell markers and the high plasticity of stem cells, biomarker-based identification may not precisely quantify stem cells. Administering incorrect doses of stem cells often results in unsuccessful outcomes both in human and veterinary medicine. This book provides the best and most updated information to students, clinicians, young budding veterinary scientists, professionals, and researchers. Reaching out current information to the young mind is critical for scientific research and teaching. Merits of stem cell therapy for the animal disease are naïve but poorly understood; therefore, they have not been practiced widely. The importance of stem cell research should not be underestimated. This book reviews the principle, practices, and up-to-date knowledge of animal stem cells therapeutic applications in veterinary medicine. We hope that our efforts will provide readers updated information about the application of stem cells in regenerative medicine of the animal diseases and techniques of characterizing them. It may provide a reference book of stem cell applications in veterinary and animal sciences.

Ludhiana, Punjab, India  
Ludhiana, Punjab, India  
2021

Ratan Kumar Choudhary  
Shanti Choudhary

---

# Contents

## Part I Overview and Introduction

<b>1</b>	<b>Overview of Stem Cells and Their Applications in Veterinary Medicine . . . . .</b>	<b>3</b>
	Ratan K. Choudhary	
<b>2</b>	<b>Introduction to Mammary Gland and Its Cell Types . . . . .</b>	<b>25</b>
	Tajeshwar Preet Kaur, Ramneek Verma, and Ratan K. Choudhary	
<b>3</b>	<b>Mammary Stem Cells: How Much Do We Know? . . . . .</b>	<b>39</b>
	Alok Kumar and Satish Kumar	
<b>4</b>	<b>Methods of Identification and Characterization of Stem Cells . . . . .</b>	<b>51</b>
	Shanti Choudhary	
<b>5</b>	<b>Potential of Stem Cell Therapy to Combat Mastitis in Dairy Animals . . . . .</b>	<b>63</b>
	Neelesh Sharma, Sapna Devi, and Goran Bacic	

## Part II Stem Cells and Veterinary Research

<b>6</b>	<b>Fatty Liver Disease and Utility of Stem Cells in Developing the Disease Model . . . . .</b>	<b>79</b>
	Shanti Choudhary, Michelle LaCasse, Donald C. Beitz, and Eric D. Testroet	
<b>7</b>	<b>Mammary Epithelial Cells: A Potential Cellular Model to Understand the Impact of Heat Stress on Mammary Gland and Milk Production in Dairy Animals . . . . .</b>	<b>97</b>
	Manishi Mukesh, Nampher Masharing, Preeti Verma, Manish Tiwari, Prince Vivek, and Monika Sodhi	
<b>8</b>	<b>Milk and Milk-Derived Stem Cells . . . . .</b>	<b>111</b>
	Ratan K. Choudhary	

- 9 Cryopreservation of Testicular Stem Cells and Its Application in Veterinary Science . . . . . 125**  
Tanushree Patra, Rakesh Bhaskar, and Mukesh Kumar Gupta
- 10 Testicular Stem Cell Niche . . . . . 161**  
Devendra Pathak, Kritima Kapoor, and Mukesh Kumar Gupta
- 11 Proteomics of Mammary Gland and Mammary Stem Cells . . . . . 183**  
Sudarshan Kumar, Nikunj Tyagi, Ashok Kumar Mohanty, and Jai Kumar Kaushik

### **Part III Therapeutic Applications**

- 12 Advancing Quantitative Stem Cell Dosing for Veterinary Stem Cell Medicine . . . . . 207**  
Samuel R. Boutin and James L. Sherley
- 13 Mesenchymal Stem Cells: A Novel Therapy for the Treatment of Bovine Mastitis . . . . . 223**  
Oscar A. Peralta
- 14 Therapeutic Applications of Mesenchymal Stem Cells in Canine Diseases . . . . . 241**  
Mudasir Bashir Gugjoo, Amarपाल, A. C. Saxena, Rohit Kumar, P. Kinjavdekar, A. M. Pawde, and G. Taru Sharma
- 15 Biomaterials and Scaffolds in Stem Cell Therapy . . . . . 255**  
Mukesh Kumar Bharti, Vikash Chandra, and G. Taru Sharma
- 16 Prospects of Mesenchymal Stem Cell Secretome in Veterinary Regenerative Therapy . . . . . 271**  
Vikash Chandra and G. Taru Sharma
- 17 Reprogramming and Induced Pluripotent Stem Cells in Porcine . . . . . 289**  
Sujoy K. Dhara, Basavaraj K. Sajjanar, and Jyotirmoy Ghosh
- 18 CRISPR/Cas System and Stem Cell Editing: Prospects and Possibilities in Veterinary Sciences . . . . . 323**  
Md Saddam Hussain and Manish Kumar

### **Part IV Issues and Perspectives**

- 19 Identification of Species-Specific Stem Cells and Challenges . . . . . 357**  
Ratan Kumar Choudhary
- 20 Regulations of Animal Cell-Based Drugs in Veterinary Regenerative Medicine . . . . . 367**  
Ratan Kumar Choudhary and Shanti Choudhary

---

## Editors and Contributors

---

### About the Editors

**Ratan Kumar Choudhary** is an Assistant Professor at the College of Animal Biotechnology, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab, India. He holds a master's degree in veterinary sciences and a doctoral degree in animal science with a specialization in bovine mammary stem cells and their characterization. His research chiefly focuses on the development of stem cell therapy for the dry period and mastitis in dairy animals. He worked at the United States Department of Agriculture (USDA) for six years (2007–2012), followed by two post-doctoral trainings at the University of Kentucky (2012–2013) and the University of Vermont (2018–2020), USA. He received many awards and fellowships during his Ph.D. including the Dean's Fellowship, R. F. Davis Scholarship and Jacob K. Goldhaber Travel Award, Beltsville Agricultural Research Center, USDA poster awards, first prize winner of the National Milk Producers Federation, Graduate Student Paper Presentation Award in Dairy Production, and Novus International Inc. Travel Award. He is currently serving as a reviewer of the international journals. He has published more than 40 research articles in peer-reviewed international journals, attended/presented research papers at more than 30 conferences, and authored one book. He is a member of various international scientific societies and organizations, e.g., the American Dairy Science Association, the European Association for Animal Production (EAAP), the Indian Society for the Study of Reproduction and Fertility, and the Veterinary Council of India.

**Shanti Choudhary** served at the University of Vermont, Burlington, the USA, as a Lab Manager and Lab Safety Officer for more than a year. She has extensive research experience in many aspects of molecular biology, including mammary stem cell physiology. She worked at the USDA in Beltsville, USA, as a visiting scientist for more than two years, where her focus was on parasitic diseases in animals. She has also worked as a Research Associate at Guru Angad Dev Veterinary and Animal Sciences University. She received a National Post-doctoral Fellowship (N-PDF; SERB-DST, India) for a project on developing an in vitro



model for the characterization and manipulation of buffalo mammary stem cells. She has published more than 30 research articles in peer-reviewed international journals and is a member of several scientific societies, e.g., the American Society for Biochemistry and Molecular Biology and the American Dairy Science Association. Currently, she is engaged in teaching at the Students College of Animal Biotechnology, GADVASU, Ludhiana.

---

## Contributors

**Amarpal** Veterinary Surgery Division, Indian Veterinary Research Institute, Bareilly, UP, India

**Goran Bacic** Clinic for Reproduction and Theriogenology, Faculty of Veterinary Medicine, University of Zagreb, Zagreb, Croatia

**Donald C. Beitz** Department of Animal Sciences, Iowa State University, Ames, IA, USA

**Mukesh Kumar Bharti** Department of Veterinary Physiology & Biochemistry, Faculty of Veterinary and Animal Sciences, BHU, RGSC, Mirzapur, UP, India

**Rakesh Bhaskar** Department of Biotechnology and Medical Engineering, National Institute of Technology, Rourkela, Odisha, India

**Samuel R. Boutin** Asymmetrex, LLC, Boston, MA, USA

**Vikash Chandra** Physiology and Climatology Division, Indian Veterinary Research Institute, Bareilly, UP, India

**Ratan Kumar Choudhary** College of Animal Biotechnology, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab, India

**Shanti Choudhary** Department of Animal and Veterinary Sciences, University of Vermont, Burlington, VT, USA

**Sapna Devi** Laboratory of Animal Stem Cells, Division of Veterinary Medicine, Faculty of Veterinary Science & Animal Husbandry, Sher-e-Kashmir University of Agricultural Sciences & Technology of Jammu, Jammu, India

**Sujoy K. Dhara** Veterinary Biotechnology Division, Indian Veterinary Research Institute, Bareilly, UP, India

**Jyotirmoy Ghosh** Molecular Biology Laboratory, Division of Physiology, ICAR - National Institute of Animal Nutrition and Physiology, Bangalore, Karnataka, India

**Mudasir Bashir Gugjoo** Division of Veterinary Clinical Complex, FVSc & AH, SKUAST, Jammu, India

**Mukesh Kumar Gupta** National Institute of Technology, Rourkela, Odisha, India

**Md Saddam Hussain** Department of Biosciences and Bioengineering, Indian Institute of Technology Guwahati, Guwahati, Assam, India

**Kritima Kapoor** Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab, India

**Tajeshwar Preet Kaur** College of Animal Biotechnology, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab, India

**Jai Kumar Kaushik** Animal Biotechnology Centre, National Dairy Research Institute, Karnal, India

**P. Kinjavdekar** Division of Surgery, Indian Veterinary Research Institute, Izatnagar, UP, India

**Alok Kumar** Department of Oncology, School of Medicine, Johns Hopkins University, Baltimore, MD, USA

**Manish Kumar** Department of Bioscience and Bioengineering, Indian Institute of Technology, Guwahati, India

**Rohit Kumar** Division of Surgery, Indian Veterinary Research Institute, Izatnagar, UP, India

**Satish Kumar** Central University of Haryana, Jant-Pali, India

**Sudarshan Kumar** Animal Biotechnology Centre, National Dairy Research Institute, Karnal, India

**Michelle LaCasse** Department of Animal and Veterinary Sciences, University of Vermont, Burlington, VT, USA

**Nampher Masharing** ICAR-National Bureau of Animal Genetic Resources, Karnal, Haryana, India

**Ashok Kumar Mohanty** Animal Biotechnology Centre, National Dairy Research Institute, Karnal, India

**Manishi Mukesh** ICAR-National Bureau of Animal Genetic Resources, Karnal, Haryana, India

**Devendra Pathak** College of Veterinary Sciences, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab, India

**Tanushree Patra** Department of Biotechnology and Medical Engineering, National Institute of Technology, Rourkela, Odisha, India

**A. M. Pawde** Division of Surgery, Indian Veterinary Research Institute, Izatnagar, UP, India

**Oscar A. Peralta** University of Chile, Región Metropolitana, Chile

**Basavaraj K. Sajjanar** Stem Cell Laboratory, Division of Veterinary Biotechnology, ICAR - Indian Veterinary Research Institute, Bareilly, UP, India

**A. C. Saxena** Division of Surgery, Indian Veterinary Research Institute, Izatnagar, UP, India

**G. Taru Sharma** Physiology and Climatology Division, Indian Veterinary Research Institute, Bareilly, UP, India

**Neelesh Sharma** Division of Veterinary Medicine, Sher-e-Kashmir University of Agricultural Sciences and Technology, Jammu, India

**James L. Sherley** Asymmetrex LLC, Boston, MA, USA

**Monika Sodhi** ICAR-National Bureau of Animal Genetic Resources, Karnal, Haryana, India

**Eric D. Testroet** Department of Animal and Veterinary Sciences, University of Vermont, Burlington, VT, USA

**Manish Tiwari** ICAR-National Bureau of Animal Genetic Resources, Karnal, Haryana, India

**Nikunj Tyagi** Animal Biotechnology Centre, National Dairy Research Institute, Karnal, India

**Preeti Verma** ICAR-National Bureau of Animal Genetic Resources, Karnal, Haryana, India

**Ramneek Verma** College of Animal Biotechnology, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab, India

**Prince Vivek** ICAR-National Bureau of Animal Genetic Resources, Karnal, Haryana, India

---


## Part I

# Overview and Introduction



# Overview of Stem Cells and Their Applications in Veterinary Medicine

# 1

Ratan K. Choudhary 

## Abstract

Stem cell therapy has high hopes, and several veterinary diseases may be treated using stem cells. Recent advances in stem cell isolation, characterization, and studies parallel in human medicine have advanced this field. Stem cells possess self-renewal ability and infinite capacity to divide and differentiated into functional cells. The therapeutic potential of either autologous or allogeneic transplantation—the regeneration potential of stem cells gained through immunomodulatory functions by secreting growth factors and many cytokines. Many studies demonstrated in various animal models that stem cell transplantation is beneficial, especially in chronic and problematic diseases. This chapter describes different types of animal stem cells, mechanisms of their action, and some of the diseases in which treatment has shown promising effects in curing the ailments. The application of mesenchymal stem cells, derived from variety of tissues, in diseases of livestock species and pet animals is noteworthy.

## Keywords

Stem cells · Types · Application · Mechanism · Veterinary medicine

## 1.1 History of Stem Cells

Stem cells (SCs) are blank or non-specialized cells capable of differentiating into various types of cells of the body. Self-replication, the capacity to differentiate into several cell types, and unlimited proliferation are the specialized properties of a stem

---

R. K. Choudhary (✉)

Department of Bioinformatics, College of Animal Biotechnology, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, Punjab, India

© The Author(s), under exclusive license to Springer Nature Singapore Pte Ltd. 2021

R. K. Choudhary, S. Choudhary (eds.), *Stem Cells in Veterinary Science*, [https://doi.org/10.1007/978-981-16-3464-2\\_1](https://doi.org/10.1007/978-981-16-3464-2_1)

3

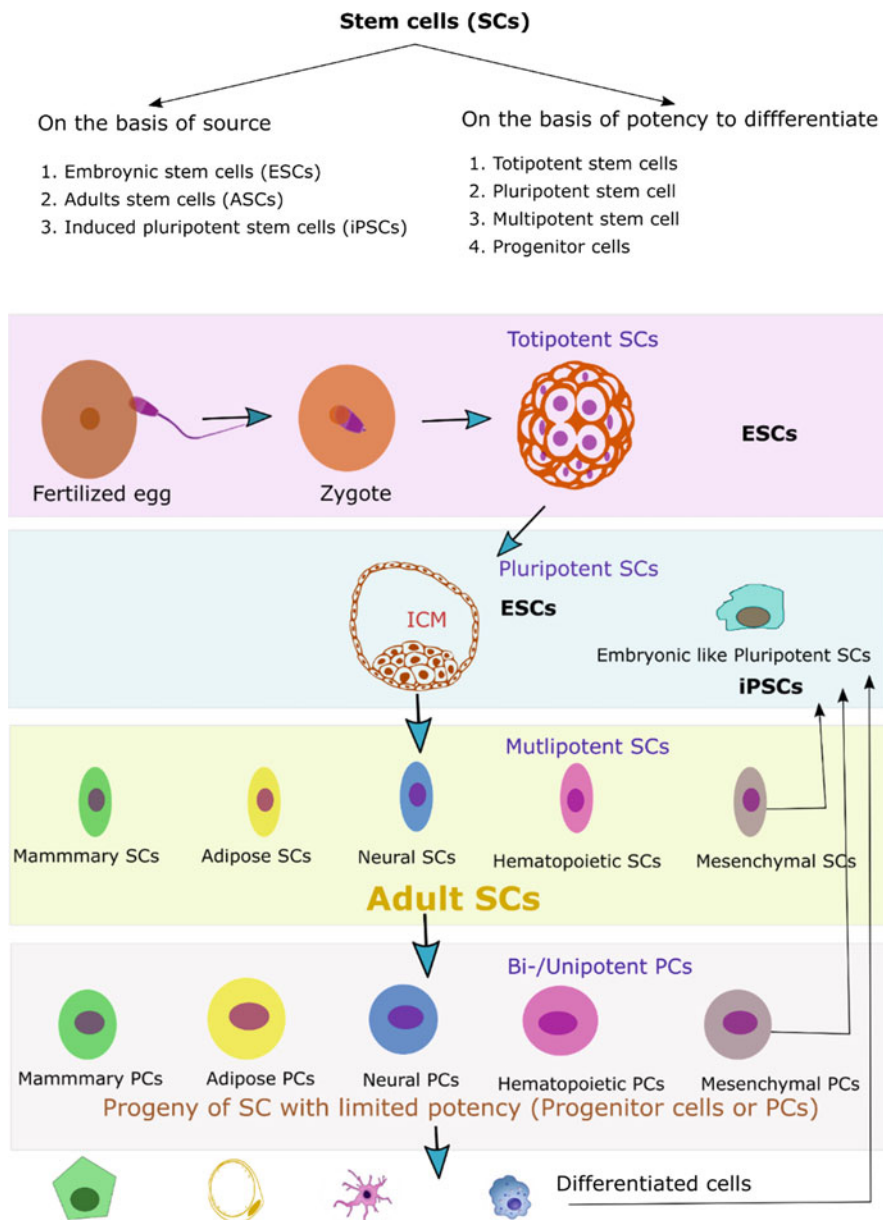
cell. The existence of stem cells was shown long back in 1959 when DeOme and his colleagues characterized hyperplastic nodular lesion of mouse mammary tumour virus (MMTV) in mouse mammary gland (DeOme et al. 1959). By that time, investigators had developed the technology of removing endogenously developing epithelial cells from fat pad (called “cleared pad”) under the mammary gland of mice and the ability to develop epithelium from exogenously infused cells in the cleared pad. There were many other investigators during 1960s who discovered similar regeneration properties of specialized cells in spleen and other organs. Later, slow proliferating clonogenic precursor cells of hemopoietic tissue were discovered where the successive passage of initial colonies of precursors cells retained the capacity to form stromal tissue like that of their hemopoietic organs (Friedenstein et al. 1974). Discovery of the pluripotent embryonic stem-like cells from inner cell mass (ICM) (Martin 1981; Evans and Kaufman 1981), where cells could form descents of all the tissue layers of the adult, has set a new promises of stem cell research. In the year 2007, the discovery of induced pluripotent stem cells (iPSCs) independently by two scientists namely, Shinya Yamanaka from Kyoto University, Japan (Takahashi and Yamanaka 2006) and James Thomson of University of Wisconsin-Madison, USA (Yu et al. 2007) led the revolution of stem cell research for therapeutic applications.

Though, it is hard to pinpoint when, where, and who discovered the foundation work of stem cells but the first blood transfusion attempted soon after the discovery of blood circulation system in 1628 by William Harvey. Key properties of stem cells defined by McCulloch and till in 1960s in mice spleen appears to be the one of the earliest discoveries of blood forming cells, hematopoietic stem cells or HSCs (Becker et al. 1963). Thus, the foundation of stem cell research lies with adult stem cells (HSCs) but not with the embryonic stem cells. Finally, the discovery of iPSCs has fuelled tremendous impetus to the regenerative medicine.

---

## 1.2 Types of Stem Cells

Stem cells, based on the source, has been classified into three types: (1) embryonic stem cells (ESCs), (2) adult stem cells (ASCs), and (3) artificially induced pluripotent stem cells (iPSCs). Another classification based on potency and differentiation capacity includes (1) totipotent, (2) pluripotent, (3) multipotent, (4) unipotent, and (5) progeny of stem cells called progenitor cells (Fig. 1.1). Cells derived from inner cell mass (ICM) of blastocysts are the source of ESCs. Somatic or adult stem cells are the undifferentiated cells present in various organs after its development. The key role of ASCs is the healing, growth, and repair of tissue in the event of damage. Among the various types of ASCs, the name of these cells depends on the tissue from where cells have been derived. They are mesenchymal stem cells (MSCs) of bone marrow, neural stem cell (NSCs), hematopoietic stem cells, skin stem cells, mammary stem cells, and so on. ESCs and ASCs are obtained from the body, but iPSCs are created or manmade stem cells that are induced from differentiated cells by rewiring various pluripotency transcription factors (OCT4, SOX2, c-Myc, and KLF4) of the genome of differentiated cells to behave like embryonic stem cells.



**Fig. 1.1** Various types of stem cells and their differentiation

Capacity of totipotent stem cells includes the generation of embryonic (embryo proper) plus extra-embryonic tissues such as placenta. Cells of zygote and up to morula stage (solid balls of 16–32 celled stage) are totipotent. Cells derived from

inner cell mass (ICM) of blastocysts are pluripotent and are the source of ESCs. Pluripotent stem cells classified into naïve and primed ESC based on their in vitro growth characteristics and potency to give rise to all somatic cell lineages. In mouse embryo proper, naïve ESC are the cells' pre-implanted blastocysts whereas primed ESCs are of post-implanted blastocysts (Takahashi et al. 2018). In comparison to toti- and multi-potent stem cells, proliferative potential and differentiation ability of multi-potent stem cells are limited and produces only few varieties of cells. Generally, variety of cells are derived from closely related lineages. Adult stem cells (ASCs) are the good examples of multi-potent stem cells. Unipotent stem cells have very limited differentiation potential but have a unique repeated divisional capacity (Zakrzewski et al. 2019). Features of repeated divisional capacity make unipotent stem cell a good candidate for therapeutic application. Stem cell of skin that divides and produces dermatocytes is the good example of unipotent stem cell.

---

### 1.3 Stem Cells of Veterinary Importance

**Embryonic Stem Cells** Isolation of human ESCs in 1998 provided the first evidence of pluripotency and given rise to more than 200 cell types (Thomson 1998). Identification of pluripotency transcription factors namely OCT4, SOX2, NANOG, c-Myc, and KLF4 in ESCs provided the core internal circuit of pluripotency. Later, it was demonstrated how interplay between these pluripotency transcription factors determines differentiation of cells during transition phase in progenitor cell. For example, transcriptional circuit dynamic analysis showed OCT4 and SOX2 proteins maintain ESC identity by suppressing neural ectodermal differentiation while SOX2 promotes neural differentiation by suppressing mesodermal differentiation in progenitor cells (Thomson et al. 2011).

Use of ESCs in therapeutic applications is still controversial due to ethical concerns especially with human ESCs. In animals, in addition to ethical concern, the use of animal ESCs also involves religious issues. Therefore, the exploration of animal ESCs is yet to be explored fully. Moreover, lack of specific marker to identify ESCs in various animal species, the vulnerability of ESCs for teratoma formation (Solter 2006), and genomic instability of ESCs occurring during in vitro passaging (Tosca et al. 2015) caused to explore the alternate source of creating embryonic-like stem cells called induced pluripotent stem cells.

---

### 1.4 Adult Stem Cells

**Mesenchymal Stem Cells (MSCs)** MSCs are the alternative source of stem cells present in adult tissue. Though the origin of MSCs is not fully understood and hypothesized to derive from pericytes (Crisan et al. 2008), perivascular cells of microvasculature present in every organ and thus MSCs are harvested from bone marrow, adipose tissue, umbilical cords, dental pulp, and other organs including tonsils. MSCs are not only available in plenty but also lack ethical issues of tissue



harvest and least immunogenic upon autologous and heterologous application. MSCs are multipotent and give rise to bone, cartilage, tendon, ligaments, fat cells, skin, muscle, and connective tissues. Thus, the application of MSCs received the greatest attention in regenerative medicine. In companion animals, dogs, cats, and horse MSCs were harvested from (1) bone marrow, (2) adipose tissues, (3) placenta, (4) Wharton's jelly, (5) synovial fluids, (6) peripheral blood, (7) muscle and periosteum, (8) umbilical cord blood, and (9) muscle (reviewed in Voga et al. 2020). With the graded success of MSC therapy, various animals were depended on the source of stem cells and the species. Conventionally, MSCs are collected from either bone marrow or from adipose tissues. Cells from bone marrow are aspirated from femur of dogs and cats and from sternum of horse for the prospective isolation of MSCs (Marx et al. 2015). Bone marrow-derived MSCs are brought to the lab in special condition and proceed with either direct applications (after centrifugation to collect mononuclear cells) into the patients or culture for further purification and expansion and later applied to the patients. While the direct application of bone marrow-derived MSCs is not the pure population of MSCs but the mixtures of other cell types, growth factors and hormones and hence may provide quicker and better results. However, the culture and expansion of MSCs in the laboratory provides enriched population and abundant quantity for heterologous transplantations of cells into more patients. After fourth passages, MSCs are usually ready for clinical use and up to tenth passage cells can be harvested (Lee et al. 2014; Markoski 2016). In the later passage, MSCs loose in vivo characteristics and tends to differentiate, evidenced by the morphological characteristics and gene expression studies.

Applications of adipose-derived MSC for the treatment of inflammation of mammary gland, mastitis are noteworthy. Intra-mammary infusion of fetal adipose tissue-derived MSC into *S. aureus*-induced mastitis reduced somatic cell counts in bovine milk, indicating the possibility of allogenic transplantation of MSCs in the cow is safe for therapeutic application (Peralta et al. 2020). Fetal bovine MSC secretes anti-proliferative factors and antibacterial peptides. In vitro studies indicated conditioned media of MSC decreased about 30% of bacterial population (Cahuascanco et al. 2019) and contained proangiogenic factors that supports neovascularization (Tao et al. 2016). One report on the application of adipose tissue-derived MSCs to treat goat mastitis indicated the efficacy of somatic tissue-derived stem cells in morpho-functional differentiation of cells into milk-producing epithelial cells (Costa et al. 2019). Such studies are indicating promising capabilities of adult stem cell's multipotentiality and immunomodulation properties as an alternative to the treatment of mastitis. Biotechnological and clinical potential of MSCs in livestock species and prospectus of its therapeutic applications has been described here (Hill et al. 2019).

**Hematopoietic Stem Cells (HSCs)** Similar to MSCs, HSCs are also available in plenty of amount from the bone mammary of developed (adult) tissue. Like that of MSCs, HSCs are multipotent stem cells which can give rise to different cells of immune systems, red and white blood corpuscles (RBCs and WBCs), and platelets.

**Mammary Stem Cells (MaSCs)** Application of MaSC for the use of treatment of mastitis is yet to be explored. Based on the work of secretome of MSC that enabled identification of healing processes by participating in inflammation, cell proliferation, remodelling of tissue repair and bacterial clearance (Krasnodembskaya et al. 2010; Cortés-Araya et al. 2018), MaSC-derived mammosphere has led the identification of factors and molecules that promoted angiogenesis, cell migration, and enhance cellular defence in the secretome (Ledet et al. 2018).

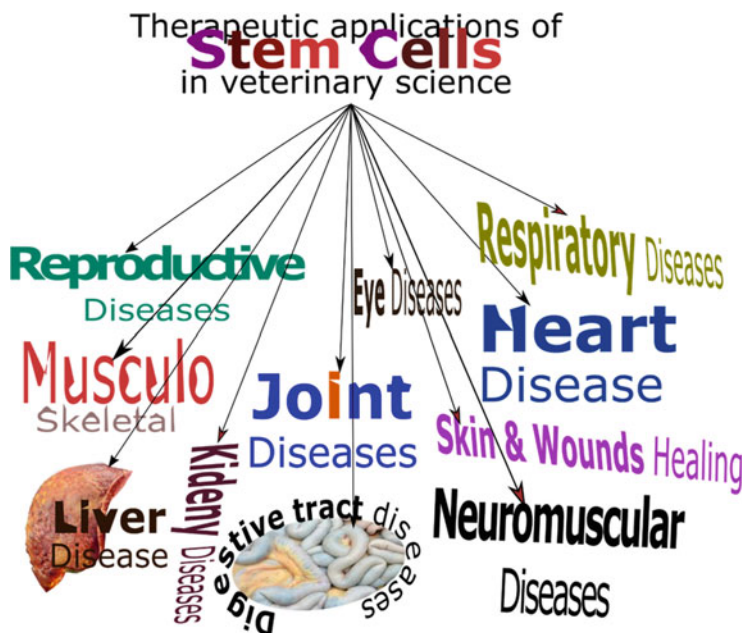
**iPSCs** The possibilities of use of iPSCs in veterinary science may bring great possibilities. In comparison to adult stem cells, use of iPSCs involves more safety challenges as all the induced cells are pluripotent and hence liable to uncontrolled division and tumour formation. Additionally, embryonic-like stem cells divide symmetrically (2 identical daughter cells) rather than asymmetrical (2 non-identical daughter cells, say one differentiated and the other stem cell). For the safety and tissue regeneration point of view, asymmetrical cells division is advantageous than the symmetrical division because it repairs the tissue while maintaining the stem cell pool. Moreover, upon receiving the differentiation signal, iPSCs tends to differentiate on all and none principle meaning all iPSCs will either terminally differentiated or none of them will differentiate. Adult stem cells have default asymmetrical divisional capacity in which they divide, differentiate, and maintain the stem cell pool in the tissues.

Remarkable works have been done on exploring the possibilities of ESCs and later iPSCs in pigs and other ungulates by Bhanu and his team. Difficulty in the identification of ICM (the source of ESCs) due to lack of ICM cell marker in ungulates hampered the research in those biomedically important species (Telugu et al. 2010). However, later the group successfully developed iPSCs from pig and other ungulates and showed capabilities of iPSCs to produce differentiated cells (Zhao et al. 2016). Potential of ESCs and iPSCs for committed progenitor cells and finally terminally differentiated cells for tissue or organ development is yet to come. In conclusions, a number of diseases can be treated with the applications of ESCs and ASCs (Fig. 1.2).

---

## 1.5 Mechanisms of Stem Cell Actions

Regeneration potentials of stem cells are achieved through re-epithelization, immunomodulations, remodelling of extracellular matrix and proangiogenic factors. Stem cells secrete growth factors, cytokines, growth regulators, anti-inflammatory factors, and some antibacterial peptides (Wangler et al. 2021). Initially, it was thought that stem cells, upon proliferation, provides (1) differentiated cells at the site of injection, or (2) migrate to distantly located tissue sites when injected into the circulation. Under the hypoxic condition, which is the microenvironment where stem cells maintain 'stemness' and the cell releases angiogenic growth factors like SDF-1, VEGF, FGF-2, angiopoietin-1, growth suppressor like TGF-alpha and



**Fig. 1.2** Applications of stem cells for therapeutic purposes in various diseases of animals

TGF-beta and many cytokines like IL-1, IL-6, IL-10 (Spees et al. 2016) and even miRNAs loaded in extracellular vesicles (Asgarpour et al. 2020).

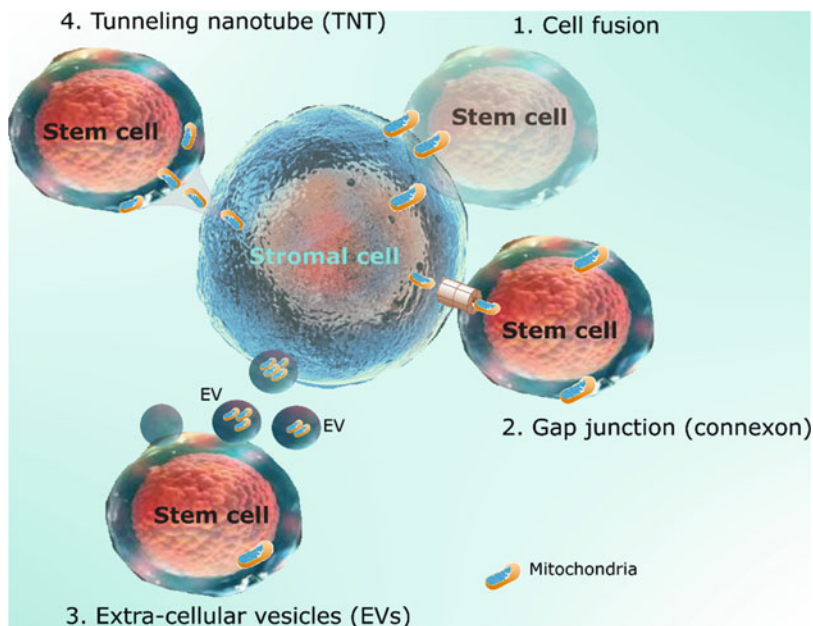
**Transfer of Extracellular Vesicles (EVs)** EVs include microvesicles, exosomes, and ectosomes that are generated either inward or outward budding of cellular plasma membrane. EVs are the lipid bilayer structures loaded with the cargos like proteins, fats, lipids, metabolites, nuclei acid (miRNA, mRNA), and other molecules has emerged as a significant cellular communicator (Raposo and Stoorvogel 2013). These EVs travel long distances inside the body and release from the healthy and diseased cells. Thus, stem cells also release EVs and transfer cargos to the other cell as a means of cellular communication. EVs was shown to increase the concentration of cytokine IL-10 (Park et al. 2019), IL-22, IL-23 (Hyvarinen et al. 2018), and TFG-beta (Crain et al. 2019). Communications between MSCs and cancer cells induces hybrid cell formation, indirectly evidenced by the presence of exosomes and micro-vesicles, which either promote or inhibit tumour (Melzer et al. 2018). Roles of EVs in stem cell biology has been discussed in detail elsewhere (Hur et al. 2020).

**Paracrine Signalling** Initial efforts to use stem cell therapy for the purpose of tissue regeneration was centred on the directed differentiation of these cells to the intended cell types. More recently, however, it is becoming apparent stem cells secrete factors which mediate action via paracrine fashion, rather than stem cell differentiation and repopulation. Paracrine mode of signalling is the cell-to-cell

communication method where, signalling molecule is released by one cell and the changes are induced in neighbouring cells. A resultant shift in research and the emergence of studies aiming to elucidate the paracrine mechanisms of stem cell action underlies tissue repair and regeneration mechanisms. Release of TGF-beta causes cell proliferation, differentiation, wound healing and angiogenesis. Appropriate regulation of TGFβ by the osteocytes in the tissue homeostasis has been emphasized for the management of bone diseases (Xu et al. 2018). This was based on the hypothesis that the eroded bone matrix of diseased patient releases TGFβ that has mitogenic and pro-survival activities, thus contributing to tumorigenesis (Portela et al. 2014). In addition, cardiac levels of IL1β and TNFα, factors implicated in angiogenesis, following bone marrow-MSc transplantation into ischemic myocardium showed optimal therapeutic potential of stem cell (Kamihata et al. 2001).

**Immunomodulation** There are number of evidences showed that MSCs interact with innate and adaptive immune cells and bring immunomodulatory activities. MSCs inhibited CD<sup>4+</sup> Th1 and Th2 cells, CD<sup>8+</sup> T cells and NK cells proliferation and functions. Inhibition of T-cell was mediated by soluble cytokines and factors including TGFβ and human growth factors (Zhao et al. 2016). By understanding mechanisms by which MSCs modulate inflammation and contribute to healing will benefit the investigation of cellular therapy of animals. Animal models like pigs for cardiovascular diseases (Murphy et al. 2003) and goats for osteoarthritis (Shake et al. 2002) are currently used in understanding human diseases. It is a well-known fact that MSCs are hypoallergic due to low expression of HLA class I and no expression of HLA class II molecules and hence devoid of allogenic tissue rejection (Ryan et al. 2005). In addition to the release of soluble cytokines by MSCs, apoptotic and metabolically inactive MSCs also possess immunomodulation properties (Song et al. 2020), indicating cytokine independent mechanism. Apoptotic tissues-derived MSCs reduced rat mortality after sepsis induction, resulting in significantly low levels of circulating TNFα and T-cells (Chang et al. 2012). The current status of cellular and molecular mechanisms of MSC-mediated immunomodulation by live MSCs, dead MSCs and apoptotic MSCs has been discussed in the literature (Weiss and Dahlke 2019).

**Mitochondrial Transfer** Dysfunction of mitochondria has been associated with ageing and certain diseases like Alzheimer's disease, Parkinson's disease, and type 2 diabetes. Mitochondrial dysfunction contributes to reactive oxygen species (ROS) production which ultimately disturbs homeostatic mechanism. Apart from the therapeutics mechanisms of stem cells, directly by cell differentiation, or indirectly *via* paracrine signalling, EVs, and immunomodulation, stem cells have also been shown to transfer mitochondria to the diseased cells. Mitochondrial transfer mediated in bone (Guo et al. 2020) and renal proximal tubular epithelial cells (Konari et al. 2019) enhanced cell proliferation, migration, and cellular differentiation in the tissue. Co-culture experiments of PC12 cells (neuronal cell line) with bone marrow-derived MSCs showed protective effects of neuronal cells under ischemic condition (Sarmah et al. 2020). Increasing evidences suggest that stem cells can directly donate



**Fig. 1.3** Patterns of mitochondrial transfer from stem cell to stromal cell or diseased cell. These mechanisms include, (1) fusion of stem cell with the stromal cell, (2) transfer of mitochondria through gap junction (connexons, a transmembrane tunnel protein), (3) via extracellular vesicles (EVs) and (4) intracellular tunnelling nanotube (TNT) formation. Formation of TNT is regulated by  $TNF\alpha/NFK\beta$  and  $TNF\alpha ip2$  signalling pathway (Jiang et al. 2016)

mitochondria in order to recover from cell injury, thus cells rescue mitochondrial damage-provoked tissue degeneration (Li et al. 2019). Mitochondrial transfer by stem cells thus could represent an emerging therapeutic approach for tissue regeneration of treating animal diseases. Various possible patterns of mitochondrial transfer from stem cell to somatic or diseased cells are presented (Fig. 1.3).

**Homing Mechanisms of Stem Cells** Homing is the process where stem cells migrate to specific tissue sites in response to chemoattractant gradients, released by the damaged tissue. Chemotactic factors like cytokines released from the damaged tissue. Understanding homing of stem cells is essential to enhance and refine its clinical significance in therapeutic settings. Chemo-attractive signals are perceived by the receptors present on the surface of stem cells. For example, stromal cell-derived factor 1 (SDF-1) and osteopontin (OPN) are the chemokine released by the damaged tissue sends chemoattractive signals to the cells expressing CXCR4 (Wynn et al. 2004) and integrin- $\beta 1$  (Chen et al. 2014), respectively. There are many other factors like hepatocyte growth factor, insulin-like growth factor, TGF- $\beta 1$ , dimers of vascular cell adhesion molecule 1 and very late antigen 4 (VCAM 1-CLA 4), matrix metalloproteinases 2 (MMP-2) and tissue inhibitor of metalloproteinases 3 (TIMP-1), PDGFR, and others (Becker et al. 1963).

Homing of stem cells depends on the route of administration of the cell. Local injection of stem cells is preferred but an invasive method like intraperitoneal injection. Intravenous injection of stem cells is the least invasive but suffers with many limitations. The limitations of intravenous injections of stem cells are (Voga et al. 2020): (1) stem cells have to exit circulation in order to reach the site of injury, (2) cells are squeezed in dimension to the size of narrowest capillary, particularly in lungs where the average diameter of capillary is 14  $\mu\text{m}$  in comparison to the cell's diameter (25–30  $\mu\text{m}$ ), (3) high expression of integrin-beta 1 by lung epithelium attracts more number MSCs which blocks the availability of MSCs by cell entrapment, (4) systemic injection of stem cells may require high dose of stem cells to have a sufficient number of stem cells reaching to the site of injury, and (5) circulation of stem cells in the blood for a longer period (more than 24 h) may not provide viable MSCs to the damaged tissue. However, long-term effects of MSCs have been reported which likely to be mediated through non-viable and apoptotic MSCs.

---

## 1.6 Clinical Applications of Stem Cells in Regenerative Veterinary Medicine

Worldwide, investigators and veterinary clinicians are working on applications of stem cell for therapeutic potential in autologous and allogenic fashions either freshly isolated or cultured and expanded stem cell in the laboratory for the treatment of various diseases of animals. This indicates that the stem cell therapy in veterinary science is a reality. Existing literatures support that the stem cell transplantation is safe and beneficial to the animal health.

***Diseases of Reproductive System*** Reproductive diseases in animals causes great loss to farmer's profit. Inflammation of uterus (endometritis), poorly developed ovarian follicles in female, and testicular degenerative disease in males are some of the reproductive disorders. In mares, endometritis is the main cause of reproductive failure and an incurable disease. Application of bone marrow-derived MSC did not restore ovarian functions in aged mares (Grady et al. 2019). However, in human MSCs therapy appears to restore ovarian functions (Yoon 2019). One may argue that the failure of MSC therapy in mare could be (1) due to old age of mare as fertility is naturally reduced in aged animals, (2) source of stem cells as MSCs can be derived from various tissue namely adipose, bone marrow, umbilical cord, menstrual blood in human, amniotic fluid, and (3) insufficient dose of MSCs. Study showed that, in human, multiple doses of MSCs ( $1-5 \times 10^6$  cells/kg body weight) improved ovarian functions in primary ovarian insufficiency (Gadkari et al. 2014), whereas in mice  $1 \times 10^6$  to  $5 \times 10^8$  cells (Kalhori et al. 2018) were used for clinical applications. Mechanisms of stem cell action in restoring ovarian functions have been shown to help in folliculogenesis, prevention of granulosa cell apoptosis, regulation of reproductive hormones and vasculogenesis.

Stem cells treatment could offer solutions to infertility related disease. Testicular degeneration in dogs often leads to the production of defective sperms and affects fertility. Viability and fertility of frozen-thawed canine sperm were improved using adipose tissue-derived MSCs (Qamar et al. 2020) and exosomes derived from conditioned media of MSCs (Qamar et al. 2019). These results showed that like the availability of frozen semen from cattle, we may use freeze pure breed canine sperm for artificial insemination of female dogs. It is well known that during cryopreservation, cellular injury to organelles like plasma membrane, mitochondria, and nucleus occurs that affects post-thaw sperm motility and hence impaired fertility. The possible mechanisms of improved fertility of canine were due to the secretion of proteins like annexin, dysferlin, and fibronectin by MSCs (Qamar et al. 2020). These proteins are involved in the repairing process at a cellular level at many ways like prevention of cells from apoptosis and improvement of membrane integrity. Other possible mechanisms of restoring canine fertility are also possible like regulating glucose metabolism of sperm (Hsiao et al. 2019) that provides more energy to the cell for enhanced motility.

**Musculoskeletal Disease** Treatment of horse musculoskeletal disease using MSCs has been fairly a good success. Many reports with varying successful results have been reported since more than a decade (Wilke et al. 2007; Broeckx et al. 2014, 2019). Intra-articular administration of adipose tissue-derived MSCs in horse resulted no lameness after 3 months of treatment (Nicpon et al. 2013). Properties, cultural characteristic, and potential applications of MSCs in the management of equine diseases have been described (Gugjoo et al. 2019a, b). Ready-to-use stem cell containing products (like Arti-Cell Forte from Boehringer Ingelheim and Investigational Veterinary Product or IVP from Dechra Limited, UK) are also available for the clinically applications in treating horse's degenerative disease.

Autologous stem cell therapy using adipose tissue-derived MSCs treated chronic osteoarthritis in canine (Black et al. 2007). MSCs derived from umbilical cord (UMSCs) have also shown promising results in treating osteoarthritis in dogs. No adverse reactions were reported in using UMSCs (Kim et al. 2019). Interestingly, allogenic transplantation of MSCs appears to be safe in dogs for osteoarthritis (Shah et al. 2018). In vivo studies summarizing the clinical effects of MSCs in canine osteoarthritis are given in Table 1.1. Majority of the results of stem cell applications for canine disease therapy are positive; however, investigators consider the results inconclusive (Gugjoo et al. 2019a, b), suggesting more basic research are needed to understand the biology of MSCs or other stem cell types for positive and conclusive results.

**Liver Diseases** Liver diseases (acute and chronic) are common in canines. Viral or bacterial infection, cancer, trauma, ingestion of toxic substances, and endocrine disturbances can cause liver disease in dogs. Many studies have tried the administration of autologous and allogeneic MSCs treatment with varied success. Mechanisms by which MSCs repair liver tissue is the generation of different cell

**Table 1.1** Various studies and their results of canine mesenchymal stem cell therapy for osteoarthritis (table adapted from Sasaki et al. 2019)

Source	Injection/trans-plantation	Combination use	Cell number (million)	Model	Evaluation method
Bone marrow	Single intraarticular, autologous	–	7–8	Partial thickness articular cartilage defect	Histology
Adipose tissue	Single intraarticular, autologous	–	7–8	Partial thickness articular cartilage defect	Morphology, histology, fluorescence analysis
Adipose tissue	Four times intraarticular, allogeneic	–	5 (three times), 66 (once)	Intact	Pain and lameness scoring, immunohistochemistry
Bone marrow	Transplantation with scaffold, allogeneic	Scaffold	0.01	Full-thickness cartilage defect	Histology, immunohistochemistry, micro-CT
Synovium	Single intraarticular, autologous	Hyaluronic acid	0.05, 5 or 50	Partial thickness articular cartilage defect	Histology
Adipose tissue	Single intraarticular, allogeneic	Platelet-rich plasma	10	Cranial cruciate ligament transection	Lameness scoring, focal compression strength, extracellular matrix composition, histopathology, real-time PCR
Bone marrow	Single intraarticular, allogeneic	Hyaluronic acid	10	Partial thickness articular cartilage defect	Gross appearance, magnetic resonance imaging, histology, immunohistochemistry
Umbilical cord	Single intraarticular, allogeneic	–	1	Surgical manipulation of articular cartilage	Magnetic resonance imaging, radiography, ultrasonography, blood test, scanning electron microscope



types of the liver (hepatocytes, Kuffer cells, and hepatic stellate cells) and the secretion of various cytokines that modulates the immune system, apoptosis, and vasculogenesis. In canines, the liver fibrosis model to establish with autologous transplantation of bone-marrow-derived MSCs and documented improved liver fibrosis without adverse effect (Matsuda et al. 2017). However, caution should be kept in mind that prolongs immunomodulation and anti-inflammatory conditions to treat liver fibrosis may have adverse effects and thus, alternative strategies must be taken to improve outcomes of stem cell therapy for prolong use in treating chronic conditions. Strategies to improve liver stem cell success may include modification of stem cell niche, pretreatment or co-treatment of the recipient and gene modifications (Hu et al. 2019). Intravenous injection of adipose tissue-derived MSCs experimentally induced liver injury and caused restoration of liver function (Teshima et al. 2017), indicating the possibility of stem cell therapy for hepatic disease in dogs.

***Kidney Diseases*** Chronic diseases of kidney is one of the common issues of old aged cats and dogs. The disease is manifested by fibrosis, nephritis, and tubular atopy. Currently, only the renal transplantation is the therapeutic solution to the chronic kidney disease (Voga et al. 2020). Very few studies conducted the same investigator (Quimby et al. 2016; Quimby and Dow 2015) and reported various strategies to look critically for MSCs-based therapy for the feline chronic kidney diseases. More research is required to know about the best source of MSCs (bone-marrow vs adipose tissue-derived cells), route of cell administration, dose of cells, stages of MSCs including the impact of age/health/status of tissue donor animals (SAGE 2018).

***Joint Diseases*** To study the effects of stem cells therapy in bovine, caprine could be the alternative model animal. Bovines are heavy and especially grazing animals have locomotor issues. One such study aiming to test the efficacy of autologous MSCs for superior cartilage repair, bone marrow harvested cells were injected intra-articularly and demonstrated hyaline like cartilage regeneration in goats (Nam et al. 2013), suggesting potential applicability of stem cell therapy in livestock animals. In horse, bone spavin is a degenerative joint disease of horse and cattle evidence by bony outgrowth within the lower hock joint caused by osteoarthritis. Result of a study conducted in horses with intraarticular autologous transplantation of adipose tissue-derived MSCs suggested positive and long-lasting effect of stem cell therapy and had no signs of lameness after 180 days of the treatment (Nicpon et al. 2013), indicating long-lasting effects of stem cell therapy. There are many other studies in pet animals showing improved and promising results of stem cell treatment in joint diseases.

***Digestive Tract Diseases*** Application of stem cell therapy has gained importance in treating inflammatory conditions of the digestive disorders especially inflammatory bowel diseases (IBD). In canine, a single dose (at two million/kg body weight) of adipose tissue-derived MSCs has reduced gastrointestinal inflammation (Pérez-Merino et al. 2015). Not much work has been done on this aspect in other animals.

Robust stem cell isolation and in vitro propagations protocol, characterization of different subsets of stem cells for various diseases of digestive system and understanding of digestive disorders are the keys to establish stem cell therapy.

***Eye Diseases*** Dry eye disease or keratoconjunctivitis sicca (KCS) is one of the prevalent eye diseases affecting up to 35% of humans and 4–20% of dogs. The mechanisms of the development of the disease are not really understood. However, the most common cause of KCS is an immune-mediated inflammatory response targeted against the production of tear by affecting lacrimal glands. It causes discomforts, visual disturbances, and damage to the ocular surface. Treatment of KCS is difficult and effective and safe treatment of this disease is lacking (Villatoro et al. 2015). However, in 2016 Bittencourt et al. reported that even a single dose of allogeneic administration of MSCs in canine has positive effects on KCS over the long period (up to a year).

***Heart Diseases*** Cardiac diseases in animals include myocardial infarction, dilated cardiomyopathy, degenerative valvular disease, congestive heart failure, and other heart problems. Research has indicated the ability of cardiomyocytes (cells of the heart) to regenerate after myocardial injury, indicating the possibility of therapeutic interventions. In an interesting study, one study claims for improved heart functions in dogs suffering with chronic valvular disease when administered with puppy's deciduous teeth stem cells (Petchdee and Sompeewong 2016). In a study conducted on Dobermans with retrograde coronary venous, allogeneic administration of adipose tissue-derived MSCs did not increase days of survival but the method of stem cell treatment was safe (Pogue et al. 2013). In addition to intracoronary infusion and intramyocardial injection of stem cells, retrograde coronary venous infusion (RCVI) is one of the methods of stem cell administration aimed for improved cardiac functions (Gathier et al. 2018). A combination of two stem cells—mesenchymal (MSCs) and cardiac stem cells (CSC)—administered intra-cardially to immunosuppressed swine (model animal for human research) showed (Quijada et al. 2015) twofold reduction in scar tissue formation, improved left ventricular compliance and contractibility in comparison to placebo animals.

***Skin and Wound Healing*** Skin wound healing is a complex process and well-orchestration of inflammation, matrix formation and tissue remodelling. Cell therapies bring combined actions of immune modulation, growth factors, angiogenesis, extracellular matrix production, and cell differentiation (Giles et al. 2015). Chronic wounds are difficult to heal, as they are associated with other underlying diseases. Generally, in a normal wound healing process, skin infection is prevented and tissue integrity and functions are restored. Faster rate of healing to prevent infection, but compromised healing process resulting in scar formation. Cell-based therapy has actively been used for the treatment of dermal wounds in dogs. Multipotent stem cells and progenitor cells have high proliferative potential, ability to differentiate and secrete different types of growth factors and cytokines that improves wound healing. Cells like endothelial progenitor cells, bone marrow-

derived mesenchymal stem cells (BM-MSCs), and adipose-derived stem cells (ASCs) are being used for cellular therapy. Interestingly, allogenic transplantation of ASCs in canine is possible and results in improved wound healing (Enciso et al. 2020). In a recent study conducted using BM-MSCs, both fresh and frozen cells were capable of promoting wound healing (Bharti et al. 2020), suggesting stem cells have promising potential to treat the cutaneous wound in clinical cases.

**Mastitis** Economic loss due to mastitis is huge. In the United States alone, mastitis causes approximately \$2 billion dollars annual loss to the dairy industry (Donovan et al. 2005). The current treatment of mastitis is to use broad-spectrum antibiotics to control bacterial infection but do not address the tissue regeneration. In vitro study suggests antibiotics induce hypoxia inducing factor 1 alpha (HIF1A) and causes oxidative damage (Elliott and Jiang 2019) that may even slow down the growth of mammary epithelial cells and tissue regeneration. Although antibiotic is the best available method to control mastitis, but has been associated with antibiotic resistance and milk residue. Recent study showed the potential application of stem cells in treating bovine mastitis. *S. aureus* induced mastitis in Holstein Friesian cows treated intramammarily with antibiotics (control group: days 4 and 5) or a suspension of adipose tissue-derived stem cells ( $2.5 \times 10^7$  cells: day 4 and 5) showed reduced colony formation unit (CFU/ml of milk) of bacterial in stem cells quarters (Peralta et al. 2020). Interestingly inoculation of repeated doses of allogenic stem cells was safe to use in healthy cows demonstrating possible cell-based therapy for mastitis treatment. In another study, conditioned DPBS from amniotic membrane stem cells injected into the induced mastitis mammary gland of cow showed improved milk quality of milk of the treated cow in comparison to the antibiotic-treated cow. Upon injection of conditioned media into the teat canal, pH value and titratable acidity of milk were significantly different in the experimental group (Ting et al. 2020), indicating another possibility of using AMSC-derived secretome as an alternative therapy in replacing antibiotics in treating bovine mastitis.

**Neuromuscular Diseases** Spinal cord injury due to trauma is one of the common injuries in dogs. Sometimes, naturally occurring intervertebral disc disease in larger breeds of dogs like German Shepherd has also been observed whose conventional treatment of the disease has very limited success. Injury to the spinal cord is because neuromuscular disease is manifested by the altered gait, pain, and sometimes permanent locomotor disability. Autologous and allogenic bone marrow MSCs therapy alone or in combination with conventional therapy showed some locomotor recovery in dogs. However, naturally occurring degenerative disc disease in German Shepherd dogs did not show positive clinical outcome after 1, 6, and 12 months of MSCs treatment (Steffen et al. 2017).

## 1.7 Conclusions

With advancement in understanding biology of stem cells and their applications in animal diseases and trial results, it seems possible to use healing properties, immunomodulatory activities, and low immunogenicity of adult stem cells, in particular mesenchymal stem cells, for therapeutic applications in various diseases of livestock species and pet animals.

---

## References

- Asgarpour K, Shojaei Z, Amiri F, Ai J, Mahjoubin-Tehran M, Ghasemi F, ArefNezhad R, Hamblin MR, Mirzaei H (2020) Exosomal microRNAs derived from mesenchymal stem cells: cell-to-cell messages. *Cell Commun Signal* 18(1):149. <https://doi.org/10.1186/s12964-020-00650-6>
- Becker AJ, McCulloch EA, Till JE (1963) Cytological demonstration of the clonal nature of spleen colonies derived from transplanted mouse marrow cells. *Nature* 197:452–454
- Bharti MK, Bhat IA, Pandey S, Shabir U, Peer BA, Indu B, Bhat AR, Kumar GS, Amarpal CV, Sharma GT (2020) Effect of cryopreservation on therapeutic potential of canine bone marrow derived mesenchymal stem cells augmented mesh scaffold for wound healing in guinea pig. *Biomed Pharmacother* 121:109573
- Bittencourt MK, Barros MA, Martins JF et al (2016) Allogeneic mesenchymal stem cell transplantation in dogs with keratoconjunctivitis sicca. *Cell Med* 8(3):63–77. <https://doi.org/10.3727/215517916X693366>
- Black LL, Gaynor J, Gahring C, Adams C, Aron D, Harman S, Gingerich DA, Harman R (2007) Effect of adipose-derived mesenchymal stem and regenerative cells on lameness in dogs with chronic osteoarthritis of the coxofemoral joints: a randomized, double-blinded, multicenter, controlled trial. *Vet Ther* 8(4):272–284
- Broeckx S, Zimmerman M, Crocetti S, Suls M, Mariën T, Ferguson SJ, Chiers K, Duchateau L, Franco-Obregón A, Wuertz K, Spaas JH (2014) Regenerative therapies for equine degenerative joint disease: a preliminary study. *PLoS One* 9:e85917
- Broeckx SY, Seys B, Suls M, Vandenberghe A, Mariën T, Adriaensen E, Declercq J, Van Hecke L, Braun G, Hellmann K, Spaas JH (2019) Equine allogeneic chondrogenic induced mesenchymal stem cells are an effective treatment for degenerative joint disease in horses. *Stem Cells Dev* 28(6):410–422. <https://doi.org/10.1089/scd.2018.006>
- Cahuascanco B, Bahamonde J, Huaman O et al (2019) Bovine fetal mesenchymal stem cells exert antiproliferative effect against mastitis causing pathogen *Staphylococcus aureus*. *Vet Res* 50:25. <https://doi.org/10.1186/s13567-019-0643-1>
- Chang CL, Leu S, Sung HC, Zhen YY, Cho CL, Chen A et al (2012) Impact of apoptotic adipose-derived mesenchymal stem cells on attenuating organ damage and reducing mortality in rat sepsis syndrome induced by cecal puncture and ligation. *J Transl Med* 10:244. <https://doi.org/10.1186/1479-5876-10-244>
- Chen Q, Shou P, Zhang L, Xu C, Zheng C, Han Y, Li W, Huang Y, Zhang X, Shao C, Roberts AI, Rabson AB, Ren G, Zhang Y, Wang Y, Denhardt DT, Shi Y (2014) An osteopontin-integrin interaction plays a critical role in directing adipogenesis and osteogenesis by mesenchymal stem cells. *Stem Cells* 32(2):327–337. <https://doi.org/10.1002/stem.1567>
- Cortés-Araya Y, Amilon K, Rink BE, Black G, Lisowski Z, Donadeu FX, Esteve CL (2018) Comparison of antibacterial and immunological properties of mesenchymal stem/stromal cells from equine bone marrow, endometrium, and adipose tissue. *Stem Cells Dev* 27(21):1518–1525. <https://doi.org/10.1089/scd.2017.0241>

- Costa CRM, Feitosa MLT, Rocha AR, Bezerra DO, Leite YKC, Argolo Neto NM et al (2019) Adipose stem cells in reparative goat mastitis mammary gland. *PLoS One* 14:e0223751. <https://doi.org/10.1371/journal.pone.0223751>
- Crain SK, Robinson SR, Thane KE, Davis AM, Meola DM, Barton BA et al (2019) Extracellular vesicles from Wharton's Jelly mesenchymal stem cells suppress CD4 expressing T cells through transforming growth factor beta and adenosine signaling in a canine model. *Stem Cells Dev* 28:212–226. <https://doi.org/10.1089/scd.2018.0097>
- Crisan M, Yap S, Casteilla L, Chen CW, Corselli M, Park TS et al (2008) A perivascular origin for mesenchymal stem cells in multiple human organs. *Cell Stem Cell* 3(3):301–313
- Deome KB, Faulkin LJ Jr, Bern HA, Blair PB (1959) Development of mammary tumors from hyperplastic alveolar nodules transplanted into gland-free mammary fat pads of female C3H mice. *Cancer Res* 19(5):515–520
- Donovan DV, Kerr DE, Wall RJ (2005) Engineering disease resistant cattle. *Transgenic Res* 14:563–567
- Elliott RL, Jiang X-P (2019) The adverse effect of gentamicin on cell metabolism in three cultured mammary cell lines: "are cell culture data skewed?". *PLoS One* 14(4):e0214586. <https://doi.org/10.1371/journal.pone.0214586>
- Enciso N, Avedillo L, Fermín ML et al (2020) Cutaneous wound healing: canine allogeneic ASC therapy. *Stem Cell Res Ther* 11:261
- Evans MJ, Kaufman M (1981) Establishment in culture of pluripotential stem cells from mouse embryos. *Nature* 292:151–156
- Friedenstein AJ, Chailakhyan RK, Latsinik NV, Panasyuk AF, Keiliss-Borok IV (1974) Stromal cells responsible for transferring the microenvironment of the hemopoietic tissues: cloning in vitro and retransplantation in vivo. *Transplantation* 17:331–340
- Gadkari R, Zhao L, Teklemariam T, Hantash BM (2014) Human embryonic stem cell derived-mesenchymal stem cells: an alternative mesenchymal stem cell source for regenerative medicine therapy. *Regen Med* 9(4):453–465. <https://doi.org/10.2217/rme.14.13>
- Gathier WA, van Ginkel DJ, van der Naald M, van Slochteren FJ, Doevendans PA, Chamuleau SAJ (2018) Retrograde coronary venous infusion as a delivery strategy in regenerative cardiac therapy: an overview of preclinical and clinical data. *J Cardiovasc Transl Res* 11(3):173–181. <https://doi.org/10.1007/s12265-018-9785-1>
- Giles TS, Kirby MSJ, Cowin AJ, Smith LE (2015) Stem cells for cutaneous wound healing. *Biomed Res Int* 2015:285869
- Grady ST, Watts AE, Thompson JA, Penedo MCT, Konganti K, Hinrichs K (2019) Effect of intra-ovarian injection of mesenchymal stem cells in aged mares. *J Assist Reprod Genet* 36:543–556. <https://doi.org/10.1007/s10815-018-1371-6>
- Gugjoo MB, Amarpal, Makhdoomi DM, Sharma GT (2019a) Equine mesenchymal stem cells: properties, sources, characterization, and potential therapeutic applications. *J Equine Vet Sci* 72:16–27. <https://doi.org/10.1016/j.jevs.2018.10.007>
- Gugjoo MB, Amarpal, Sharam GT (2019b) Mesenchymal stem cell basic research and applications in dog medicine. *J Cell Physiol* 234(10):16779–16811. <https://doi.org/10.1002/jcp.28348>
- Guo Y, Chi X, Heng BC, Wei Y, Zhang X, Zhao H, Yin Y, Deng X (2020) Mitochondria transfer enhances proliferation, migration, and osteogenic differentiation of bone marrow mesenchymal stem cell and promotes bone defect healing. *Stem Cell Res Ther* 11:245. <https://doi.org/10.1186/s13287-020-01704-9>
- Hill ABT, Bressan FF, Murphy BD, Garcia JM (2019) Applications of mesenchymal stem cell technology in bovine species. *Stem Cell Res Ther* 10(44):1145. <https://doi.org/10.1186/s13287-019-1145-9>
- Hsiao C, Ji AT, Chang C et al (2019) Mesenchymal stem cells restore the sperm motility from testicular torsion-detorsion injury by regulation of glucose metabolism in sperm. *Stem Cell Res Ther* 10:270. <https://doi.org/10.1186/s13287-019-1351-5>

- Hu C, Zhao L, Duan J, Li L (2019) Strategies to improve the efficiency of mesenchymal stem cell transplantation for reversal of liver fibrosis. *J Cell Mol Med* 23(3):1657–1670. <https://doi.org/10.1111/jcmm.14115>
- Hur YH, Cerione RA, Antonyak MA (2020) Extracellular vesicles and their roles in stem cell biology. *Stem Cells* 38(4):469–476. <https://doi.org/10.1002/stem.314>
- Hyvarinen K, Holopainen M, Skirdenko V, Ruhanen H, Lehenkari P, Korhonen M et al (2018) Mesenchymal stromal cells and their extracellular vesicles enhance the anti-inflammatory phenotype of regulatory macrophages by downregulating the production of interleukin (IL)-23 and IL-22. *Front Immunol* 9:771. <https://doi.org/10.3389/fimmu.2018.00771>
- Jiang D, Gao F, Zhang Y, Wong DS, Li Q, Tse HF et al (2016) Mitochondrial transfer of mesenchymal stem cells effectively protects corneal epithelial cells from mitochondrial damage. *Cell Death Dis* 7:e2467
- Kalhari Z, Azadbakht M, Soleimani MM, Shariatzadeh MA (2018) Improvement of the folliculogenesis by transplantation of bone marrow mesenchymal stromal cells in mice with induced polycystic ovary syndrome. *Cytotherapy* 20:1445–1458
- Kamihata H, Matsubara H, Nishiue T et al (2001) Implantation of bone marrow mononuclear cells into ischemic myocardium enhances collateral perfusion and regional function via side supply of angioblasts, angiogenic ligands, and cytokines. *Circulation* 104(9):1046–1052
- Kim SE, Pozzi A, Yeh J-C, Lopez-Velazquez M, Yong JAA, Townsend S, Dunlap AE, Christopher SA, Lewis DD, Johnson MD, Petrucci K (2019) Intra-Articular umbilical cord derived mesenchymal stem cell therapy for chronic elbow osteoarthritis in dogs: A double-blinded, placebo-controlled clinical trial. *Front Vet Sci* 6:474. <https://doi.org/10.3389/fvets.2019.00474>
- Konari N, Nagaishi K, Kikuchi S, Fujimiya M (2019) Mitochondria transfer from mesenchymal stem cells structurally and functionally repairs renal proximal tubular epithelial cells in diabetic nephropathy *in vivo*. *Sci Rep* 9:5184. <https://doi.org/10.1038/s41598-019-40163-y>
- Krasnodembskaya A, Song Y, Feng X, Gupta N, Serikov V, Lee J-W, Matthay MA (2010) Antibacterial effect of human mesenchymal stem cells is mediated in part from secretion of the antimicrobial peptide LL-37. *Stem Cells* 28:2229–2238
- Ledet MM, Vasquez AK, Rauner G, Bichoupan AA, Moroni P, Nydam DV et al (2018) The secretome from bovine mammosphere-derived cells (MDC) promotes angiogenesis, epithelial cell migration, and contains factors associated with defense and immunity. *Sci Rep* 8(1):5378
- Lee KS, Kang HW, Lee HT, Kim HJ, Kim CL, Song JY, Lee KW, Cha SH (2014) Sequential sub-passage decreases the differentiation potential of canine adipose-derived mesenchymal stem cells. *Res Vet Sci* 96(2):267–275
- Li C, Cheung MKH, Han S, Zhang Z, Chen L, Chen J, Zeng H, Qiu J (2019) Mesenchymal stem cells and their mitochondrial transfer: a double-edged sword. *Biosci Rep* 39(5):BSR20182417
- Markoski MM (2016) Advances in the use of stem cells in veterinary medicine: from basic research to clinical practice. *Scientifica*. <https://doi.org/10.1155/2016/4516920>
- Martin GR (1981) Isolation of a pluripotent cell line from early mouse embryos cultured in medium conditioned by teratocarcinoma stem cells. *Proc Natl Acad Sci U S A* 78:7634–7638
- Marx C, Silveira MD, Nardi NB (2015) Adipose-derived stem cells in veterinary medicine: characterization and therapeutic applications. *Stem Cells Dev* 24(7):803–813
- Matsuda T, Takami T, Sasaki R, Nishimura T, Aibe Y, Paredes BD, Quintanilha LF, Matsumoto T, Ishikawa T, Yamamoto N, Tani K, Terai S, Taura Y, Sakaida I (2017) A canine liver fibrosis model to develop a therapy for liver cirrhosis using cultured bone marrow-derived cells. *Hepatol Commun* 1(7):691–703. <https://doi.org/10.1002/hep4.1071>
- Melzer C, Ohe J, Hass R (2018) Concise Review: Crosstalk of mesenchymal stroma/stem-like cells with cancer cells provides therapeutic potential. *Stem Cells* 36(7):951–968. <https://doi.org/10.1002/stem.2829>
- Murphy JM, Fink DJ, Hunziker EB, Barry FP (2003) Stem cell therapy in a caprine model of osteoarthritis. *Arthritis Rheum* 48:3464–3474
- Nam HY, Karunanithi P, Loo WC, Naveen S, Chen H, Hussin P, Chan L, Kamarul T (2013) The effects of staged intra-articular injection of cultured autologous mesenchymal stromal cells on